

# Effect of ectomycorrhizal infection on growth and photosynthetic characteristics of *Pinus densiflora* seedlings grown under elevated CO<sub>2</sub> concentrations

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## Abstract

The effect of ectomycorrhizal *Pisolithus tinctorius* (*Pt*) infection was studied on the growth and photosynthetic characteristics of *Pinus densiflora* seedlings grown at ambient (360  $\mu\text{mol mol}^{-1}$ , AC) and elevated (720  $\mu\text{mol mol}^{-1}$ , EC) CO<sub>2</sub> concentrations. After 18 weeks, *Pt* inoculation had led to significantly increased dry mass and stem diameter of *P. densiflora* at both CO<sub>2</sub> concentrations, relative to non-inoculated seedlings. Moreover, EC significantly increased the ectomycorrhizal development. The phosphate content in needles inoculated with *Pt* was about three times higher than without inoculation at both CO<sub>2</sub> concentrations. The PAR saturated net photosynthetic rates ( $P_{\text{sat}}$ ) of *P. densiflora* inoculated with *Pt* were clearly higher than for control seedlings at both CO<sub>2</sub> concentrations, and the maximum net photosynthetic rate ( $P_N$ ) at saturated CO<sub>2</sub> concentration ( $P_{\text{max}}$ ) was higher than in controls. Moreover, the carboxylation efficiency (CE) and RuBP regeneration rate of the  $P_N/C_i$  curve for *P. densiflora* inoculated with *Pt* were significantly higher than for non-inoculated seedlings at both CO<sub>2</sub> concentrations, especially at EC. The water use efficiency (WUE) of seedlings inoculated with *Pt* grown at EC was significantly raised. Allocation of photosynthates to roots was greater in *Pt* inoculated pine seedlings, because of the enhanced activity of ectomycorrhiza associated with seedlings at EC. Moreover,  $P_N$  of non-inoculated seedlings grown for 18 weeks at EC tended to be down regulated; in contrast, *Pt* inoculated seedlings showed no down-regulation at EC. The activity of ectomycorrhiza may therefore be enhanced physiological function related to water and phosphate absorption in *P. densiflora* seedlings at EC.

**Additional key words:** down-regulation; dry mass; phosphorus; *Pinus densiflora*; *Pisolithus tinctorius*; seedling diameter; stomatal limitation; water use efficiency.

## Introduction

Red pine (*Pinus densiflora*) is the dominant conifer in infertile granite regions in the Korean peninsula and Japan. *P. densiflora* can survive and grow on the nutrient-poor ridges of mountains after disturbances. This species usually has a symbiotic interaction with ectomycorrhiza such as *Pisolithus tinctorius* (Pers.) Coker *et* Couch (*Pt*), allowing for vigorous growth. Where there have been several disturbances, *P. densiflora* was used for forest rehabilitation practices in nutrient-poor soils (*e.g.* Moroto *et al.* 1987), because they grow fast to mature trees in about 20 years.

The pattern and amount of precipitation at middle and

high latitudes have recently been predicted to undergo great changes with the increase in atmospheric CO<sub>2</sub> concentration [CO<sub>2</sub>] (IPCC 1996). Precipitation could decrease significantly. Moreover, photosynthetic adjustment is frequently observed in plants grown under high [CO<sub>2</sub>] because of dilution effects of nutrient, enhanced accumulation of photosynthates in photosynthetic organs, and sink strength (Wardlaw 1990, Farrar and Williams 1991, Berntson and Bazzaz 1996).

Symbiotic microorganisms such as ectomycorrhiza usually act as an efficient root system for absorbing water and essential nutrients (nitrogen, phosphate) (Bolan 1991,

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Smith and Read 1997, Quoreshi *et al.* 2003). The activity of the host plant (*P. densiflora*) should be enhanced at high  $[\text{CO}_2]$  because symbiotic ectomycorrhiza plays an essential role in absorbing water and important elements such as nitrogen and phosphate, and also act as a large carbon sink (Ceulemans and Mousseau 1994, Smith and Read 1997). Therefore, no down-regulation of photosynthesis is predicted following *P. densiflora* inoculation with ectomycorrhiza at high  $[\text{CO}_2]$ .

## Materials and methods

**Plants:** Seeds of red pine (*Pinus densiflora* Sieb. *et* Zucc.) were collected at the Yongmun Experimental Field of the Korea Forest Service (37.3°N, 127.2°E). These seeds were surface sterilized with 30 %  $\text{H}_2\text{O}_2$  for 20 min and rinsed 4–5 times with sterile de-ionized water. They were then germinated on sterilized media, black sand : vermiculate : peat moss = 3 : 2 : 1 (volume), in a glasshouse at day/night temperatures 25/20 °C with a 16-h photoperiod.

**Ectomycorrhiza** fungi, *Pisolithus tinctorius* (Pers.) Coker *et* Couch (*Pt*), were obtained from *Horticultural Alliance*, USA. The *Pt* spores were dissolved in distilled water, then inoculated directly to roots of *P. densiflora* seedlings, and mixed with prepared soil media (see below). The seedlings inoculated with *Pt* were naturalized in a rhizo-box (100×230×18 mm) filled with sterilized medium consisting of vermiculite : black sand : peat moss = 2 : 2 : 1 (volume).

**Experimental design:** Our experiments with *P. densiflora* were conducted at the Hokkaido Research Center, Forestry and Forest Products Research Institute (FFPRI), Sapporo, Japan in 2002. The seedlings were grown in a FFPRI phytotron with a natural sunlight, day/night temperature range of 26/16 °C, and humidity range of 55–75 % during the study period lasting 18 weeks. Six cabinets were used, with two differing  $[\text{CO}_2]$ . The rhizo-boxes were allocated at random such that half of the seedlings experienced ambient  $[\text{CO}_2]$ , AC (360  $\mu\text{mol mol}^{-1}$ ) and the other half experienced elevated  $[\text{CO}_2]$ , EC (720  $\mu\text{mol mol}^{-1}$ ) (Koike 1995). At each  $\text{CO}_2$  concentration (3 rooms of phytotron), half of the seedlings were *Pt* inoculated and the other half were not inoculated (control). The seedling number of each treatment in a room was sixteen. Each treatment was repeated in three rooms.

**Measurement of photosynthesis:** The net photosynthetic rate/intercellular  $\text{CO}_2$  concentration ( $P_N/C_i$ ) curves were examined using an open gas exchange system (*LI-6400*, *Li-Cor*, Lincoln, NE, USA) between 09:00 and 15:00 local time. From five to seven cm of above ground part was covered with a conifer chamber. The change in  $P_N$  was measured at saturating photosynthetic photon flux

Several ectomycorrhizas form symbiotic interactions. Of these, *Pt* is the most common species in pine forests world-wide (Marx *et al.* 1984, Allen 1991). We studied the effect of *Pt* on the physiological responses of *P. densiflora* seedlings under high  $[\text{CO}_2]$ . Our aim was to estimate the effect of *Pt* development on the down-regulation of photosynthesis responses and the growth of *P. densiflora* at enhanced  $[\text{CO}_2]$  through water use efficiency and P concentration in needles.

density (PPFD) of 1 000–1 200  $\mu\text{mol m}^{-2} \text{s}^{-1}$  that was provided by a cool halogen lamp (Walz, Effeltrich, Germany). The leaf temperature was 25 °C and the relative humidity was 50–70 %. Leaves were allowed to acclimate to their surroundings for 10 min prior to measurement, following which we began the determination with 150–1 500  $\mu\text{mol}(\text{CO}_2) \text{mol}^{-1}$ .

The initial slope of the  $P_N/C_i$  curve is proportional to the carboxylation activity of ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBPCO), *i.e.* the carboxylation efficiency CE [ $\mu\text{mol Pa}^{-1}$ ]. CE was calculated from the increment in  $C_i$  and  $P_N$  of the initial slope of the  $P_N/C_i$  curve (Terashima 1992). The maximum  $P_N$  at  $\text{CO}_2$  saturation,  $P_{\text{max}}$  [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ] was determined as 1 200  $\mu\text{mol}(\text{CO}_2) \text{mol}^{-1}$ . Water use efficiency, WUE [ $\mu\text{mol mmol}^{-1}$ ] was taken as the ratio of  $P_N$  to transpiration rate at PPFD saturation and constant water vapour pressure deficit (Field *et al.* 1983). The relative stomatal limitation of photosynthesis,  $l_s$  [%] was calculated as follows, based on  $P_N/C_i$  curves:  $l_s = (P_0 - P_N)/P_0$ , where  $P_0$  is  $P_N$  when stomatal resistance to  $\text{CO}_2$  diffusion is zero (Sharkey 1985a). After the gas exchange measurement, we measured the needle projection area with an image scanner (*FB636U*, *Canon*, Japan), and then calculated  $P_N$  per unit area.

**Infection rate:** We observed the root tips and counted both infected and non-infected roots (Quoreshi 2003). The inoculation rate of ectomycorrhiza (IRE) was determined according to the following formula:  $\text{IRE} [\%] = \text{ER}/(\text{ER} + \text{NR}) \times 100$ , where ER and NR respectively denote the number of ectomycorrhizal and non-ectomycorrhizal roots.

**Growth rate:** Stem diameter of each seedling was measured immediately by digital caliper (*CD-15B*, *Mitutoyo Corp.*, Japan) take out from rhizo-box. Shoot and root dry masses of each seedling were measured after drying at 60 °C for 1 week using an electronic balance (*HR-202*, *A&D*, Japan).

**Phosphorus analysis:** The dried samples were then ground to a fine powder in a vibrating sample mill (*Wonder Blender*, *Osaka Chemical Co.*, Osaka, Japan). To determine the content of P, the samples were digested

by a microwave digestion system (*O-I Analytical*, College Station, TX, USA) and underwent ICP analysis.

**Statistical methods:** Mean values of the inoculation rate,

## Results

**Dry mass and stem diameter:** *Pt* inoculation significantly increased the shoot dry mass relative to non-inoculated seedlings grown at AC, from 187.7 to 293.7 ( $p<0.05$ ). Moreover, the shoot dry mass of inoculated seedlings tended to increase at EC from 280.0 to 293.3 mg. The root dry mass also increased significantly relative to non-inoculated seedlings, from 169.3 to 437.0 mg at AC and from 276 to 464 mg at EC ( $p<0.05$ ) (Fig. 1). Moreover, the stem diameter was significantly greater in seedlings infected with *Pt* at both CO<sub>2</sub> concentrations ( $p<0.05$ ) (Fig. 1). *Pt* inoculation increased all growth parameters for *P. densiflora* seedlings, i.e. shoot and root dry mass and stem diameter, especially at AC. The dry masses and stem diameters of control seedlings grown at EC were significantly greater than at AC.

**Mycorrhizal development and P content in needles** (Fig. 1): After 18 weeks the ectomycorrhizal development was significantly affected by the CO<sub>2</sub> concentration ( $p<0.05$ ). The inoculation rates with *Pt* grown at AC and EC were 55.1 and 66.5 %, respectively. We did not find *Pt* or any other ectomycorrhiza development in the non-inoculated seedlings grown at both CO<sub>2</sub> concentrations. The P content in needles inoculated with *Pt* was about 3 times higher than without inoculation at both CO<sub>2</sub> concentrations ( $p<0.001$ ).

**Photosynthetic response** (Fig. 2):  $P_N$  of *P. densiflora* inoculated with *Pt* was significantly higher than for control seedlings in both [CO<sub>2</sub>] treatments. The maximum  $P_N$  at saturating CO<sub>2</sub> concentration ( $P_{max}$ ) of seedlings inoculated with *Pt* was higher than for controls ( $p<0.05$ ). Moreover, CE and ribulose-1,5-bisphosphate (RuBP)

the P content in needles,  $P_N$ , WUE,  $l_s$ , the shoot length, root growth, and stem diameter were examined by the *t*-test (Li 1964) using the *Stat View 5.0* software (SAS Institute, Cary, NC, USA).

regeneration rate of the  $P_N/C_i$  curve for *P. densiflora* inoculated with *Pt* were significantly higher than for non-inoculated seedlings at both CO<sub>2</sub> concentrations, and especially at EC (Table 1,  $p<0.05$ ).

Table 1. Parameter estimates as functions of the  $P_N/C_i$  curve. Maximum net photosynthetic rate at saturated CO<sub>2</sub> concentration ( $P_{max}$ ), carboxylation efficiency (CE), and RuBP regeneration rate of photosynthesis in the needles of *P. densiflora* seedlings grown at ambient (AC) or enhanced (EC) CO<sub>2</sub> concentrations. Means±SE, \* $p<0.05$ , \*\* $p<0.01$ , \*\*\* $p<0.001$ . (*Pt* means seedlings infected with *Pt*).

		CE	RuBP regeneration	$P_{max}$
AC	Control	0.21±0.11	14.33± 5.42	7.86±1.37
	<i>Pt</i>	0.43±0.12*	54.94± 8.05**	11.78±0.65*
EC	Control	0.17±0.02	20.18± 8.36	6.55±0.74
	<i>Pt</i>	0.44±0.06***	60.80±13.12**	13.22±1.93**

**WUE and stomatal limitation:** WUE of *Pt* inoculated seedlings at EC was significantly higher than for uninoculated seedlings ( $p<0.05$ ). However, no difference in WUE of seedlings at AC was found with *Pt* infection. In contrast, the WUE of non-inoculated seedlings grown at EC was significantly decreased, but in *Pt* inoculated seedlings the increase was not significant. Moreover, *P. densiflora* seedlings inoculated with *Pt* had lower stomatal limitation in photosynthesis ( $l_s$ ) than non-inoculated seedlings. This tendency was clearer in the *Pt* inoculated seedlings at EC ( $p<0.05$ ) (Fig. 1). The stomatal limitation in photosynthesis of *Pt* inoculated seedlings grown at EC was significantly decreased (Table 1).

## Discussion

Inoculation with *Pt* significantly increased the dry mass and stem diameter of seedlings grown at both CO<sub>2</sub> concentrations after 18 weeks (Fig. 1). Previous research on the effect of ectomycorrhiza and EC on the biomass growth of forest tree seedlings revealed a varied response (Tissue *et al.* 1996, Gavito *et al.* 2000). Growth improvement in ectomycorrhizal plants has been found in several species (Allen 1992, Mousseau *et al.* 1996, Norby *et al.* 1996, Tissue *et al.* 1996, Smith and Read 1997, Rouhier and Read 1998, 1999). However, even at EC the dry mass of the shoot of seedlings inoculated with *Pt* was similar to that for non-inoculated seedlings. This growth pattern suggests that more photosynthates were allocated to those

roots for increased activity of root cells and ectomycorrhiza, and pool accumulation of saccharides (*e.g.* Norby *et al.* 1987, Wright *et al.* 2000); this could also increase the production of fine roots and ectomycorrhiza.

The infection rate of *Pt* in *P. densiflora* seedlings grown at EC was significantly higher than at AC. The significantly enhanced P content in needles of *Pt*-inoculated seedlings was therefore due to the development of ectomycorrhiza (Fig. 1). Previous studies have also found that ectomycorrhizal development in seedlings of several tree species at EC was greater than at AC (Seegmüller and Rennenberg 1994, Ineichen *et al.* 1995, Rey and Jarvis 1997, Runion *et al.* 1997, Rouhier and

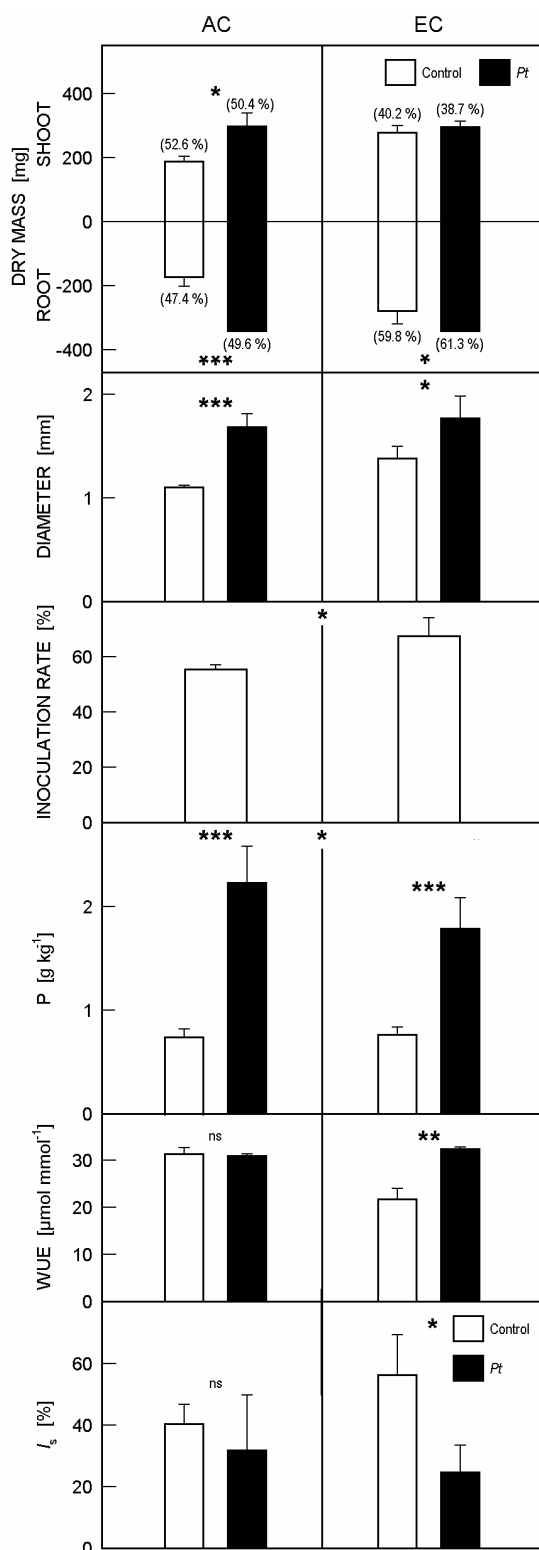


Fig. 1. Effect of ectomycorrhizal inoculation by *Pt* on the shoot and root dry mass, diameter, inoculation rate, P content, water use efficiency (WUE), and stomatal limitation ( $I_s$ ) of *P. densiflora* seedlings grown at 360  $\mu\text{mol mol}^{-1}$  (AC) or 720  $\mu\text{mol mol}^{-1}$  (EC). % values give partition rate. Means  $\pm$  SE, \* $p$  < 0.05, \*\* $p$  < 0.01, \*\*\* $p$  < 0.001.

Read 1998). Ectomycorrhizal development enlarges the absorptive surface of the root, with widely ramified hyphae allowing the release of phosphatases, which enhance the availability of organic phosphate and exude organic acids. These interactions between host plant and ectomycorrhiza increase the use efficiency of limited soluble phosphate and organic N in soil (Smith and Read 1997, Lambers *et al.* 1998). The  $P_N$ , CE, RuBP regeneration rate, and  $P_{\text{max}}$  of *Pt* inoculated *P. densiflora* seedlings grown at both  $\text{CO}_2$  concentrations were significantly higher than those of non-inoculated seedlings, and the P content in needles was also significantly greater (see Table 1, Figs. 1 and 2). These photosynthetic changes are regulated by the capacity of RuBPCO to fix  $\text{CO}_2$  and the capacity to regenerate RuBP (Farquhar *et al.* 1980, Farquhar and Sharkey 1982). Under some conditions, phosphate ( $P_i$ ) may limit photosynthesis if  $P_i$  utilization during  $\text{CO}_2$  assimilation and RuBP regeneration exceeds the capacity for  $P_i$  release during starch and sucrose synthesis (Sharkey 1985b, Lambers *et al.* 1998).

Ectomycorrhizal plants enhance the rate of photosynthesis over non-mycorrhizal plants as a result of improved plant nutritional status, including N and P (Reid *et al.* 1983, Bolan 1991, Finlay 1992, Smith and Read 1997, Qureshi 2003, Qureshi *et al.* 2003), even if the P content is the same in needles (Rousseau and Reid 1990). Moreover, ectomycorrhiza improves WUE (Guehl *et al.* 1990, Duñabeitia *et al.* 2004) by enlarging the soil volume exploited, via widely distributed hyphae (Smith and Read 1997, Lambers *et al.* 1998). In general, the P content in needles affects photosynthesis through RuBP regeneration (Sharkey 1985a, Fredeen *et al.* 1990, Kirschbaum and Tompkins 1990, Lewis *et al.* 1994), through thylakoids (Conroy *et al.* 1986, Lambers *et al.* 1998), and through the peak carboxylation velocity or peak capacity of electron transport (Harley and Sharkey 1991). We conclude that ectomycorrhiza development in inoculated seedlings and the increased P content in needles enhance the  $P_N$  and WUE and reduce stomatal limitation relative to non-inoculated seedlings (Smith and Read 1997, Hensen *et al.* 1998) (Table 1, Figs. 1 and 2).

$P_N$  of non-inoculated seedlings grown at EC for 18 weeks tends to be down regulated, *i.e.* the seedlings exert reduced CE,  $P_{\text{max}}$ , and  $P_N/C_i$  curve. In contrast, *Pt* inoculated seedlings showed no down-regulation at EC. This phenomenon has been reported in other studies with adequate soil N fertility (Idso and Kimball 1991), with  $\text{N}_2$  fixing species even when grown in poor soils (Vogel and Curtis 1995), or with mycorrhizal plants (Staddon *et al.* 1999). Phosphorus or nitrogen limitation (Conroy *et al.* 1986, Jach and Ceulemans 2000, Koike *et al.* 2000, Eguchi *et al.* 2004) and reduced sink strength (*e.g.* Stitt 1991, Rogers *et al.* 1998, Griffin *et al.* 2000) have both been proposed as mechanisms inducing down-regulation, via reduction of RuBP regeneration capacity and reduced RuBP carboxylase/oxygenase activity (Sharkey *et al.* 1994, Adam *et al.* 2004). We therefore suggest that *Pt*

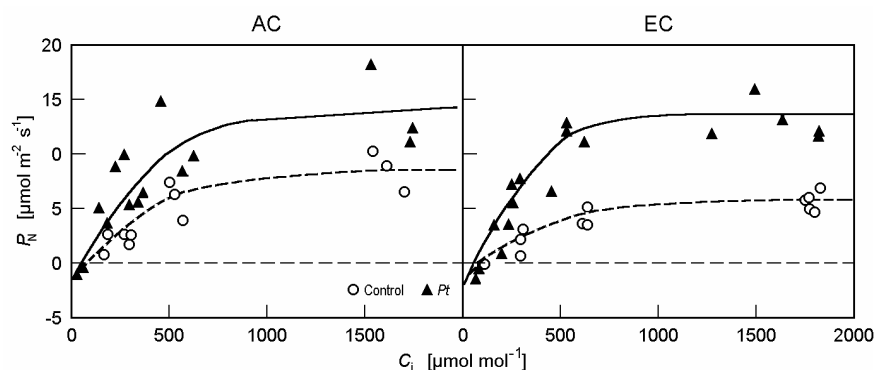


Fig. 2. Effect of ectomycorrhiza inoculation by *Pt* on the net photosynthetic rate ( $P_N$ ) response curve to intercellular  $\text{CO}_2$  concentration ( $C_i$ ) of *P. densiflora* seedlings grown at  $360 \mu\text{mol mol}^{-1}$  (AC) or  $720 \mu\text{mol mol}^{-1}$  (EC).  $\blacktriangle$  = inoculated *Pt* seedlings,  $\circ$  = control. \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .

inoculated seedlings have increased nutrient and water uptake, leading to improved plant nutritional status and giving rise to more vigorous physiological response, in particular photosynthetic activity; and that these responses delay down-regulation at EC.

Our present results indicate that ectomycorrhiza inoculation increases uptake of nutrients and water, and

the resulting vigorous physiological response significantly increases the dry mass and stem diameter of seedlings grown at AC and EC. Moreover, ectomycorrhizal seedlings moderated down-regulation at EC. Ectomycorrhiza therefore might have strong positive effects on *P. densiflora* seedlings at both  $\text{CO}_2$  concentrations.

## Reference

- Adam, N.R., Wall, G.W., Kimball, B.A., Idso, S.B., Webber, A.N.: Photosynthetic down-regulation over long-term  $\text{CO}_2$  enrichment in leaves of sour orange (*Citrus aurantium*) trees. – *New Phytol.* **163**: 341–347, 2004.
- Allen, M.F.: The Ecology of Mycorrhizae. – Cambridge University Press, New York 1991.
- Allen, M.F. (ed.): Mycorrhizal Functioning. – Chapman & Hall, New York 1992.
- Berntson, G.M., Bazzaz, F.A.: Below-ground positive and negative feedbacks on  $\text{CO}_2$  growth enhancement. – *Plant Soil* **187**: 119–131, 1996.
- Bolan, N.S.: A critical review on the role of mycorrhizal fungi in the uptake of phosphorus by plants. – *Plant Soil* **134**: 189–207, 1991.
- Ceulemans, R., Mousseau, M.: Effects of elevated atmospheric  $\text{CO}_2$  on woody plants. – *New Phytol.* **127**: 425–446, 1994.
- Conroy, J.P., Smillie, R.M., Küppers, M., Bevege, D.J., Barlow, E.W.: Chlorophyll *a* fluorescence and photosynthetic and growth response of *Pinus radiata* to phosphorus deficiency, drought stress, and high  $\text{CO}_2$ . – *Plant Physiol.* **81**: 423–429, 1986.
- Duñabeitia, M.K., Hormilla, S., Garcia-Plazaola, J.I., Txarterina, K., Arteché, U., Becerril, J.M.: Differential responses of three fungal species to environmental factors and their role in the mycorrhization of *Pinus radiata* D. Don. – *Mycorrhiza* **14**: 11–18, 2004.
- Eguchi, N., Fukatsu, E., Funada, R., Tobita, H., Kitao, M., Maruyama, Y., Koike, T.: Changes in morphology, anatomy, and photosynthetic capacity of needles of Japanese larch (*Larix kaempferi*) seedlings grown in high  $\text{CO}_2$  concentrations. – *Photosynthetica* **42**: 173–178, 2004.
- Farquhar, G.D., Caemmerer, S. von, Berry, J.A.: A biochemical model of photosynthetic  $\text{CO}_2$  assimilation in leaves of  $\text{C}_3$  species. – *Planta* **149**: 78–90, 1980.
- Farquhar, G.D., Sharkey, T.D.: Stomatal conductance and photosynthesis. – *Annu. Rev. Plant Physiol.* **33**: 317–345, 1982.
- Farrar, J.F., Williams, M.L.: The effects of increased atmospheric carbon dioxide and temperature on carbon partitioning, source-sink relations and respiration. – *Plant Cell Environ.* **14**: 819–830, 1991.
- Field, C., Merino, J., Mooney, H.A.: Compromises between water-use efficiency and nitrogen-use efficiency in five species of California evergreens. – *Oecologia* **60**: 384–389, 1983.
- Finlay, R.D.: Uptake and translocation of nutrients by ectomycorrhizal fungal mycelia. – In: Read, D.J., Lewis, D.H., Fitter, A.H., Alexander, I.J. (ed.): *Mycorrhizas in Ecosystems*. Pp. 92–97. CAB International, Wallingford 1992.
- Fredeen, A.L., Raab, T.K., Rao, I.M., Terry, N.: Effects of phosphorus nutrition on photosynthesis in *Glycine max* (L.) Merr. – *Planta* **181**: 399–405, 1990.
- Gavito, M.E., Curtis, P.S., Mikkelsen, T.N., Jakobsen, I.: Atmospheric  $\text{CO}_2$  and mycorrhiza effects on biomass allocation and nutrient uptake of nodulated pea (*Pisum sativum* L.) plants. – *J. exp. Bot.* **51**: 1931–1938, 2000.
- Griffin, K.L., Tissue, D.T., Turnbull, M.H., Whitehead, D.: The onset of photosynthetic acclimation to elevated  $\text{CO}_2$  partial pressure in field-grown *Pinus radiata* D. Don. after 4 years. – *Plant Cell Environ.* **23**: 1089–1098, 2000.
- Guehl, J.M., Mousain, D., Falconnet, G., Gruez, J.: Growth, carbon dioxide assimilation capacity and water-use efficiency of *Pinus pinea* L. seedlings inoculated with different ectomycorrhizal fungi. – *Ann. Sci. forest.* **47**: 91–100, 1990.
- Harley, P.C., Sharkey, T.D.: An improved model of  $\text{C}_3$  photosynthesis at high  $\text{CO}_2$ : Reversed  $\text{O}_2$  sensitivity explained by lack of glycerate reentry into the chloroplast. – *Photosynth. Res.* **27**: 169–178, 1991.

- Hensen, C.W., Lynch, J., Ottosen, C.O.: Response to phosphorus availability during vegetative and reproductive growth of *Chrysanthemum*: I. Whole-plant carbon dioxide exchange. – J. amer. Soc. horticult. Sci. **123**: 215-222, 1998.
- Idso, S.B., Kimball, B.A.: Downward regulation of photosynthesis and growth at high CO<sub>2</sub> levels. No evidence for either phenomenon in three-year study of sour orange trees. – Plant Physiol. **96**: 990-992, 1991.
- Ineichen, K., Wiemken, V., Wiemken, A.: Shoots, roots and ectomycorrhizal formation of pine seedlings at elevated atmospheric carbon dioxide. – Plant Cell Environ. **18**: 703-707, 1995.
- IPCC: Climate Change 1995: The Science of Climate Change, Summary for Policymakers. – Cambridge University Press, New York 1996.
- Jach, M.E., Ceulemans, R.: Effects of season, needle age and elevated atmospheric CO<sub>2</sub> on photosynthesis in Scots pine (*Pinus sylvestris*). – Tree Physiol. **20**: 145-157, 2000.
- Kirschbaum, M.U.F., Tompkins, D.: Photosynthetic responses to phosphorus nutrition in *Eucalyptus grandis* seedlings. – Aust. J. Plant Physiol. **17**: 527-535, 1990.
- Koike, T.: Effects of CO<sub>2</sub> in interaction with temperature and soil fertility on the foliar phenology of alder, birch, and maple seedlings. – Can. J. Bot. **73**: 149-157, 1995.
- Koike, T., Yazaki, K., Funada, R., Kitao, M., Maruyama, Y., Takahashi, K., Maximov, T.C., Ivanov, B.I.: Photosynthetic characteristics of Dahurian larch, Scotch pine and white birch seedlings native to eastern Siberia raised under elevated CO<sub>2</sub>. – Eurasian J. Forest Res. **1**: 31-37, 2000.
- Lambers, H., Chapin, F.S., III, Pons, T.L.: Plant Physiological Ecology. – Springer-Verlag, New York 1998.
- Lewis, J.D., Griffin, K.L., Thomas, R.B., Strain, B.R.: Phosphorus supply affects the photosynthetic capacity of loblolly pine grown in elevated carbon dioxide. – Tree Physiol. **14**: 1229-1244, 1994.
- Li, J.C.R.: Analysis of variance versus Chi-square test. – In: Statistical Inference. Pp. 471-484. Edwards Brothers, Ann Arbor 1964.
- Marx, D.H., Cordell, C.E., Kenney, D.S., Mexal, J.G., Artman, J.D., Riffle, J.W., Molina, R.J.: Commercial vegetative inoculum of *Pisolithus tinctorius* and inoculation techniques for development of ectomycorrhizae on bare-root seedlings. – Forest Sci. Monogr. **25**: 1-101, 1984.
- Moroto, M., Mashimo, Y., Harata, Y.: [Soil properties and growth of Japanese red pine (*Pinus densiflora*) in the hilly and low-mountainous region of central Japan.] – J. jap. Forestry Soc. **69**: 371-378, 1987. [In Jap.]
- Mousseau, M., Dufrêne, E., El Kohen, A., Epron, D., Godard, D., Liozon, R., Pontailier, J.Y., Saugier, B.: Growth strategy and tree response to elevated CO<sub>2</sub>: A comparison of beech (*Fagus sylvatica*) and sweet chestnut (*Castanea sativa* Mill.). – In: Koch, G.W., Mooney, G.A. (ed.): Carbon Dioxide and Terrestrial Ecosystems. Pp. 71-86. Academic Press, San Diego – New York – Boston – London – Sydney – Tokyo – Toronto 1996.
- Norby, R.J., O'Neill, E.G., Hood, W.G., Luxmoore, R.J.: Carbon allocation, root exudation and mycorrhizal colonization of *Pinus echinata* seedlings grown under CO<sub>2</sub> enrichment. – Tree Physiol. **3**: 203-210, 1987.
- Norby, R.J., Wullschlegel, S.D., Gunderson, C.A.: Tree responses to elevated CO<sub>2</sub> and implications for forests. – In: Koch, G.A., Mooney, H.A. (ed.): Carbon Dioxide and Terrestrial Ecosystems. Pp. 1-21. Academic Press, San Diego – New York – Boston – London – Sydney – Tokyo – Toronto 1996.
- Quoreshi, A.M.: Nutritional preconditioning and ectomycorrhizal formation of *Picea mariana* (Mill.) B.S.P. seedlings. – Eurasian J. Forest Res. **6**: 1-63, 2003.
- Quoreshi, A.M., Maruyama, Y., Koike, T.: The role of mycorrhiza in forest ecosystems under CO<sub>2</sub>-enriched atmosphere. – Eurasian J. Forest Res. **6-2**: 171-176, 2003.
- Reid, C.P.P., Kidd, F.A., Ekwebelam, S.A.: Nitrogen nutrition, photosynthesis and carbon allocation in ectomycorrhizal pine. – Plant Soil **71**: 415-431, 1983.
- Rey, A., Jarvis, P.G.: Growth response of young birch trees (*Betula pendula* Roth.) after four and a half years of CO<sub>2</sub> exposure. – Ann. Bot. **80**: 809-816, 1997.
- Rogers, A., Fischer, B.U., Bryant, J., Frehner, M., Blum, H., Raines, C.A., Long, S.P.: Acclimation of photosynthesis to elevated CO<sub>2</sub> under low-nitrogen nutrition is affected by the capacity for assimilate utilization. Perennial ryegrass under free-air CO<sub>2</sub> enrichment. – Plant Physiol. **118**: 683-689, 1998.
- Rouhier, H., Read, D.J.: Plant and fungal responses to elevated atmospheric carbon dioxide in mycorrhizal seedlings of *Pinus sylvestris*. – Environ. exp. Bot. **40**: 237-246, 1998.
- Rouhier, H., Read, D.J.: Plant and fungal responses to elevated atmospheric CO<sub>2</sub> in mycorrhizal seedlings of *Betula pendula*. – Environ. exp. Bot. **42**: 231-241, 1999.
- Rousseau, J.V.D., Reid, C.P.P.: Effects of phosphorus and ectomycorrhizas on the carbon balance of loblolly pine seedlings. – Forest Sci. **36**: 101-112, 1990.
- Runion, G.B., Mitchell, R.J., Rogers, H.H., Prior, S.A., Counts, T.K.: Effects of nitrogen and water limitation and elevated atmospheric CO<sub>2</sub> on ectomycorrhiza of longleaf pine. – New Phytol. **137**: 681-689, 1997.
- Seegmüller, S., Rennenberg, H.: Interactive effects of mycorrhization and elevated carbon dioxide on growth of young pedunculate oak (*Quercus robur* L.) trees. – Plant Soil **167**: 325-329, 1994.
- Sharkey, T.D.: Photosynthesis in intact leaves of C<sub>3</sub> plants: physics, physiology and rate limitations. – Bot. Rev. **51**: 53-105, 1985a.
- Sharkey, T.D.: Photosynthesis of cotton plants exposed to elevated levels of carbon dioxide in the field. – Photosynth. Res. **12**: 191-203, 1985b.
- Sharkey, T.D., Socias, X., Loreto, F.: CO<sub>2</sub> effects on photosynthetic and product synthesis and feedback. – In: Alscher, R.G., Wellburn, A.R. (ed.): Plant Responses to the Gaseous Environment. Pp. 55-78. Chapman & Hall, London – New York 1994.
- Smith, S.E., Read, D.J.: Mycorrhizal Symbiosis. – Academic Press, San Diego 1997.
- Staddon, P.L., Fitter, A.H., Robinson, D.: Effects of mycorrhizal colonization and elevated atmospheric carbon dioxide on carbon fixation and below-ground carbon partitioning in *Plantago lanceolata*. – J. exp. Bot. **50**: 853-860, 1999.
- Stitt, M.: Rising CO<sub>2</sub> levels and their potential significance for carbon flow in photosynthetic cells. – Plant Cell Environ. **14**: 741-762, 1991.
- Terashima, I.: Anatomy of non-uniform leaf photosynthesis. – Photosynth. Res. **31**: 195-212, 1992.
- Tissue, D.T., Thomas, R.B., Strain, B.R.: Growth and photosynthesis of loblolly pine (*Pinus taeda*) after exposure to elevated CO<sub>2</sub> for 19 months in the field. – Tree Physiol. **16**: 49-59, 1996.
- Vogel, C.S., Curtis, P.S.: Leaf gas exchange and nitrogen dynamics of N<sub>2</sub>-fixing field-grown *Alnus glutinosa* under elevated

- atmospheric CO<sub>2</sub>. – *Global Change Biol.* **1**: 55-61, 1995.
- Wardlaw, I.F.: The control of carbon partitioning in plants. – *New Phytol.* **116**: 341-381, 1990.
- Wright, D.P., Scholes, J.D., Read, D.J., Rolfe, S.A.: Changes in carbon allocation and expression of carbon transporter genes in *Betula pendula* Roth. colonized by the ectomycorrhizal fungus *Paxillus involutus* (Batsch) Fr. – *Plant Cell Environ.* **23**: 39-49, 2000.