

# Partitioning of photosynthetic electron flow between CO<sub>2</sub> assimilation and O<sub>2</sub> reduction in sunflower plants under water deficit

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## Abstract

In sunflower (*Helianthus annuus* L.) grown under controlled conditions and subjected to drought by withholding watering, net photosynthetic rate ( $P_N$ ) and stomatal conductance ( $g_s$ ) of attached leaves decreased as leaf water potential ( $\Psi_w$ ) declined from  $-0.3$  to  $-2.9$  MPa. Although  $g_s$  decreased over the whole range of  $\Psi_w$ , nearly constant values in the intercellular CO<sub>2</sub> concentrations ( $C_i$ ) were observed as  $\Psi_w$  decreased to  $-1.8$  MPa, but  $C_i$  increased as  $\Psi_w$  decreased further. Relative quantum yield, photochemical quenching, and the apparent quantum yield of photosynthesis decreased with water deficit, whereas non-photochemical quenching ( $q_{NP}$ ) increased progressively. A highly significant negative relationship between  $q_{NP}$  and ATP content was observed. Water deficit did not alter the pyridine nucleotide concentration but decreased ATP content suggesting metabolic impairment. At a photon flux density of  $550 \mu\text{mol m}^{-2} \text{s}^{-1}$ , the allocation of electrons from photosystem (PS) 2 to O<sub>2</sub> reduction was increased by 51 %, while the allocation to CO<sub>2</sub> assimilation was diminished by 32 %, as  $\Psi_w$  declined from  $-0.3$  to  $-2.9$  MPa. A significant linear relationship between mean  $P_N$  and the rate of total linear electron transport was observed in well watered plants, the correlation becoming curvilinear when water deficit increased. The maximum quantum yield of PS2 was not affected by water deficit, whereas  $q_p$  declined only at very severe stress and the excess photon energy was dissipated by increasing  $q_{NP}$  indicating that a greater proportion of the energy was thermally dissipated. This accounted for the apparent down-regulation of PS2 and supported the protective role of  $q_{NP}$  against photoinhibition in sunflower.

*Additional key words:* chlorophyll fluorescence; electron transport; fluorescence quenching; *Helianthus annuus*; intercellular CO<sub>2</sub> concentration; net photosynthetic rate; stomatal conductance.

## Introduction

Water deficit is the main factor limiting plant growth and yield in the world. There is a big controversy as to whether water deficit mainly limits photosynthesis by stomatal closure (Cornic 2000, Flexas *et al.* 2006) or metabolic damage (Lawlor 1995, Lawlor and Cornic 2002).

Water deficit induces stomatal closure diminishing net CO<sub>2</sub> assimilation and growth (Lawlor 1995). However, production capacity of reducing power is little affected by drought. There is evidence that the pools of reduced pyridine nucleotides are little affected (NADPH) or increased (NADH) in stressed leaves; NADP<sup>+</sup> increases and then decreases, while NAD<sup>+</sup> decreases with

electrons for the reduction of acceptors and supply of progressive stress. This suggests that both supply of oxidized nucleotides are adequate and the process is efficient even under severe stress (Lawlor and Khanna-Chopra 1984, Stuhlfauth *et al.* 1991). However, the available data for NADPH are scarce. Electron transport is, therefore, considered to continue at a significant rate even in severely stressed tissues, and the electrons are passed on to the normal physiological acceptors NADP<sup>+</sup> and ferredoxin and to alternative acceptors, such as oxygen in the Mehler reaction (Biehler and Fock 1996, Heber 2002, Kitao *et al.* 2003), and eventually used in

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*Abbreviations:*  $C_a$  – ambient CO<sub>2</sub> concentration;  $C_i$  – intercellular CO<sub>2</sub> concentration;  $F_m$ ,  $F'_m$  – maximum fluorescence after 1 h dark adaptation and under steady-state irradiation;  $F_s$  – fluorescence at steady state in the light;  $F_v$  – variable fluorescence after 1 h dark adaptation;  $F_v/F_m$  – maximum quantum yield of photosystem 2;  $g_s$  – stomatal conductance;  $J_C$  – electron flow to CO<sub>2</sub> fixation;  $J_O$  – electron flow to O<sub>2</sub>;  $J_T$  – total electron transport;  $P_N$  – net photosynthetic rate; PFD – photon flux density; PS – photosystem;  $q_p$ ,  $q_{NP}$  – coefficients for photochemical and non-photochemical quenching of fluorescence; RWC – relative water content;  $\Psi_w$  – leaf water potential;  $\Phi_{CO_2}$  – apparent quantum yield of CO<sub>2</sub> assimilation;  $\Phi_{PS2}$  – relative quantum yield of PS2 electron transport.

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photorespiration (Lawlor 1995, Lawlor and Cornic 2002). The absorption of excess photon energy by leaves, which potentially occurs when stomatal closure reduces CO<sub>2</sub> assimilation, is a source of damage in water-stressed plants (Björkman and Powles 1984, Björkman and Demmig-Adams 1994). The ability of a plant species to efficiently use its capabilities for quantum energy dissipation provides the investigator with a measure of water deficit tolerance (Scheuermann *et al.* 1991).

During water deficit, restricted CO<sub>2</sub> availability due to stomatal closure may lead to increased susceptibility to photodamage (Powles 1984). Sometimes such damage does not occur under natural and greenhouse conditions (Epron and Drier 1992, Tezara *et al.* 1999, Lawlor and Cornic 2002), suggesting that the mechanisms of protection against an excess of absorbed excitation energy are efficient. The major process involved in protection against photodamage is probably the increase in non-photochemical quenching ( $q_{NP}$ ), which reduces the quantum yield of photosystem 2 ( $\Phi_{PS2}$ ) in order to maintain an adequate balance between photosynthetic electron transport and carbon metabolism (Weis and Berry 1987, Krause and Weis 1991). Furthermore, photorespiration in C<sub>3</sub> plants may be an alternative sink for light-induced electron flow and is often presented as a process that may help to consume a considerable proportion of the electron flow during periods of high irradiance and restricted CO<sub>2</sub> availability in the chloroplasts (Stuhlfauth *et al.* 1990). Considerable oxygen uptake occurs during photorespiration of C<sub>3</sub> plants (Heber 2002).

There is good evidence that a large proportion of the

electron flux is diverted from CO<sub>2</sub> assimilation to O<sub>2</sub> reduction under stress at low intercellular CO<sub>2</sub> concentration ( $C_i$ ) (Osmond 1981). Maintenance of a high capacity for photosynthetic and photorespiratory carbon metabolism is the primary means of protection against photo-inhibition when stomata close (Osmond 1981, Powles 1984). Cornic and Briantais (1991) concluded from measurements of fluorescence parameters and calculation of the quantum yield of electron flow that the allocation of electrons to O<sub>2</sub> reduction increased, particularly at the more severe stress (Cornic 1994), as a consequence of the decrease in  $C_i$  and stomatal conductance ( $g_s$ ).

In some studies water deficit did not inhibit O<sub>2</sub> evolution at very high CO<sub>2</sub> (Chaves 1991, Quick *et al.* 1992), leading to the effects of stress on CO<sub>2</sub> assimilation being ascribed to stomatal limitation. There is little evidence supporting that water splitting is inhibited by water deficit within the physiological range. No inhibition of O<sub>2</sub> evolution and PS2 activity in mildly stressed leaves of sunflower was observed (Tezara *et al.* 1999). However, rates of O<sub>2</sub> evolution by stressed leaves have been measured with oxygen electrodes under elevated CO<sub>2</sub> and in some cases a decrease has been found (Lawlor and Khanna-Chopra 1984, Havaux *et al.* 1987).

Our main objective was to examine the effects of water deficit on the partitioning of photosynthetic electron flow between CO<sub>2</sub> assimilation and O<sub>2</sub> reduction by simultaneous measurements of gas exchange and fluorescence in attached leaves of water-stressed sunflower plants. ATP and pyridine nucleotide content were also determined.

## Materials and methods

**Plants:** Sixty sunflower plants (*Helianthus annuus* L. cv. Avante) were grown in a glasshouse in pots containing 5 000 cm<sup>3</sup> of organic soil under controlled environmental conditions: temperatures 20/18 °C day/night, 360  $\mu\text{mol}(\text{CO}_2) \text{ mol}^{-1}$ , 500  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  photon flux density (PFD, 400–700 nm), and 50–70 % relative humidity. When irradiance was lower than 300  $\mu\text{mol m}^{-2} \text{ s}^{-1}$ , additional irradiation provided at the top of the canopy by lamps gave a maximum PFD of 700  $\mu\text{mol m}^{-2} \text{ s}^{-1}$ . Plants were watered daily and supplied weekly with 100 cm<sup>3</sup> of a complete nutrient solution (*Vitax* 4 : 3 : 1). All measurements were performed on the third pair of leaves. Water deficit was induced in one-month-old plants by withholding irrigation over 12 d.

**Leaf water status:** Relative water content (RWC) of leaf discs was determined as  $\text{RWC} = (\text{FM} - \text{DM}) / (\text{TM} - \text{DM})$ , where FM is the fresh mass, TM is water saturated mass obtained after re-hydrating the disc for 6 h in distilled water at 25 °C and DM is the dry mass after oven-drying for 24 h at 80 °C. Predawn leaf water potential ( $\Psi_w$ ) was measured on three individual leaves using a custom-made pressure chamber (Rothamsted, U.K.).

**Gas exchange** was measured on fully expanded leaves, using a six-chamber computerized infra-red open gas-exchange system (WA-225-MK3; *Analytical Development Co.*, Hoddesdon, UK) as described by Jacob and Lawlor (1991). Areas of 10 cm<sup>2</sup> of leaves attached to the plant were sealed into the chambers. During measurements incident PFD was 1 100  $\mu\text{mol}(\text{photon}) \text{ m}^{-2} \text{ s}^{-1}$ , leaf temperature 25 °C, O<sub>2</sub> concentration 0.21 mol mol<sup>-1</sup>, and ambient CO<sub>2</sub> concentration ( $C_a$ ) 350  $\mu\text{mol mol}^{-1}$ . Vapour pressure deficit between the leaf and chamber was 1.0–1.4 kPa.

Photosynthetic O<sub>2</sub> evolution was measured with a leaf-disc oxygen electrode (*Hansatech*, Norfolk, U.K.) at 10 mmol mol<sup>-1</sup> of CO<sub>2</sub>. Leaf discs of 10 cm<sup>2</sup> were irradiated at 1 100  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  PFD from a slide projector bulb; temperature was controlled at 25 °C.

Irradiance-response curves ( $P_N/\text{PFD}$ ) at constant  $C_a$  (350  $\mu\text{mol mol}^{-1}$ ) and a leaf temperature of 25 °C were done by irradiating the leaf at different PFD until  $P_N$  was constant. The magnitude of PFD was modified using neutral density filters (*Lee Filters*; *A.C. Lighting*, Bucks., UK). Measurements of gas exchange were taken during steady state photosynthesis after a period of 1-h

adjustment by the leaf to the chamber conditions. Calculations of  $P_N$ ,  $g_s$ , and  $C_i$  were done after Farquhar and Sharkey (1982). Leaf absorbance was determined using an integrating sphere (*Applied Physics*), as described by Rackham and Wilson (1968). The apparent quantum yield of CO<sub>2</sub> ( $\Phi_{CO_2}$ ) was calculated as the slope of the linear portion of the  $P_N$ /PFD curve at 0–150  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PFD.

**Chlorophyll *a* fluorescence** of PS 2 and gas exchange were measured simultaneously at room temperature on intact attached leaves in a temperature-controlled metal chamber with a glass window, as described by Habash *et al.* (1995). Fluorescence was measured using a modulated fluorometer (*MSMF*; *Hansatech*, Kings Lynn, UK) providing a low-intensity modulated beam of less than 0.5  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PFD set at 580 nm using a narrow-band pass filter *585DF 44* (*Omega Optical*, Brattleboro, VT, USA) and selectively measuring the resulting fluorescence at 695 nm using a band pass filter *695 DF 30* (*Omega Optical*). Actinic irradiation was provided by metal-halide lamps used to drive photosynthesis for both gas-exchange and fluorescence measurements. To fully reduce  $Q_A$  a 2-s saturating flash of 7 000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PFD of 400–635 nm was shone through a short-pass filter from a pulse source *PLS2* (*Hansatech*, Kings Lynn, UK). Far-red radiation of 15 W  $\text{m}^{-2}$  was supplied by the *PLS2* through a far-red filter *RG 715* (*Schott*, Mainz, Germany) for the determination of minimum fluorescence ( $F'_0$ ) at the steady state photosynthesis. The protocol for fluorescence measurements was similar to that described by Genty *et al.* (1989), except that the measurements were done on attached leaves. The relative quantum yield of PS2 at steady state photosynthesis is defined as  $\Phi_{PS2} = (F'_m - F'_s)/F'_m$  according to Genty *et al.* (1989), where  $F'_s$  and  $F'_m$  are fluorescence signal at steady state photosynthesis and maximum fluorescence under irradiation, respectively.

**Determination of ATP and pyridine nucleotides:** The amount of ATP was determined by an enzymatic method

## Results

**Leaf water status:** Decreasing water supply decreased RWC and  $\Psi_w$  of sunflower leaves. There was a strong linear positive relationship between RWC and  $\Psi_w$  ( $\text{RWC} = 18.9 \Psi_w + 95.4$ ;  $r^2 = 0.90$ ;  $p < 0.05$ ). A decrease of  $\Psi_w$  to −3.0 MPa with a 40 % reduction in RWC was observed.

**CO<sub>2</sub> exchange:**  $P_N$  and  $g_s$  decreased with water deficit by 98 and 97 %, respectively, as  $\Psi_w$  declined from −0.3 to −3.0 MPa (Fig. 1A,B). Although  $g_s$  decreased over the whole range of  $\Psi_w$ , the  $C_i/C_a$  ratio showed little response to  $\Psi_w$ , remaining at an average value of 0.8 as  $\Psi_w$  decreased to −2.0 MPa, but  $C_i/C_a$  increased as  $\Psi_w$  decreased further (Fig. 1C).

of Stitt *et al.* (1989). Oxidized ( $\text{NAD}^+$ ,  $\text{NADP}^+$ ) and reduced ( $\text{NADH}$ ,  $\text{NADPH}$ ) pyridine nucleotides were determined by the method of Carrier and Neve (1979).

**Electron-transport rate in leaves:** Whole chain electron transport rate in the leaves ( $J_T$ ) was estimated by the method of Krall and Edwards (1992) from the equation:

$$J_T = \Phi_{PS2} \text{ PFD } a f$$

where  $a$  is the fraction of incident PFD absorbed by the leaf, and  $f$  the absorption of PS2 divided by the absorption of PS1+PS2. It is assumed that the two photosystems are equally involved in linear electron transport, so a value of 0.5 was used for  $f$ .

Electron transport allocation to CO<sub>2</sub> and O<sub>2</sub> was calculated as described by Peterson (1989). Assuming that the linear electron flow is uniquely devoted to carboxylation and oxygenation by ribulose-1,5-bisphosphate carboxylase/oxygenase, *i.e.* all the other processes consuming electrons are either negligible or at least constant (Cornic and Briantais 1991), partitioning of electron flow was calculated as

$$J_T = J_C + J_O$$

where  $J_C$  and  $J_O$  are the electron flows attributable to carboxylation and oxygenation reactions, respectively. Assuming that four electrons are consumed per CO<sub>2</sub> molecule fixed,  $J_C$  was estimated as

$$J_C = 1/3 [J_T + 8 (P_N + R_D)]$$

$$J_O = 2/3 [J_T - 4 (P_N + R_D)]$$

where  $R_D$  (day respiration) is the rate of CO<sub>2</sub> evolution in the light from processes other than photorespiration.  $R_D$  was determined as described by Jacob and Lawlor (1993).

**Statistics:** The statistical analyses were done using the statistical packages *Statistica* and *Sigmaplot*. All linear single regressions, correlations, and one way ANOVA were tested for significance at  $p < 0.05$ . The data shown are the results of at least three different experiments. Results are presented as means of a minimum of five or maximum of ten measurements plus one standard.

**Response of  $P_N$  and fluorescence to PFD:** Under well-watered conditions,  $P_N$  was not saturated by photon energy at a PFD of 1 200  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . However, as  $\Psi_w$  decreased to −2.9 MPa, energy saturation was evident and maximum  $P_{N\text{max}}$  progressively decreased (Fig. 2).  $\Phi_{CO_2}$  also declined from 0.035 to 0.004 as  $\Psi_w$  decreased from −0.3 to −2.9 MPa. The fraction of PFD absorbed by the leaf was  $0.89 \pm 0.15$ , and did not change with  $\Psi_w$ .

The  $\Phi_{CO_2}$  was more affected by moderate stress than  $\Phi_{PS2}$ , which decreased only at severe water deficit (Fig. 3A). There was a strong positive, curvilinear relationship between  $\Phi_{CO_2}$  and  $\Phi_{PS2}$  ( $r^2 = 0.80$ ,  $p < 0.05$ ). The  $q_p$  followed a similar trend as  $\Phi_{PS2}$ , decreasing only

at very severe water deficit (Fig. 3B), whereas  $q_{NP}$  increased progressively in a linear fashion with declining  $\Psi_w$  ( $r^2 = 0.91$ ;  $p < 0.05$ ) (Fig. 3B).  $F_v/F_m$  was not affected by decreasing  $\Psi_w$  (Fig. 3C).

ATP content decreased significantly (Table 1), while  $q_{NP}$  increased as  $\Psi_w$  decreased. A non-linear negative

relationship between  $q_{NP}$  and ATP was observed ( $r^2 = 0.64$ ;  $p < 0.05$ ; Fig. 4).

Water deficit did not alter pyridine nucleotide contents (Table 1). The NADPH and NADH contents were maintained, so that the ratio of total reduced to oxidised nucleotides was unaffected by water deficit.

Table 1. ATP [ $\mu\text{mol m}^{-2}$ ] and pyridine nucleotide contents [ $\mu\text{mol m}^{-2}$ ] of fully expanded leaves of sunflower measured at control and water deficit.

$\Psi_w$ [MPa]	ATP	NADH	NADPH	NAD <sup>+</sup>	NADP <sup>+</sup>
$-0.36 \pm 0.09$	$18.20 \pm 1.50$ a	$0.78 \pm 0.12$ a	$1.40 \pm 0.30$ a	$6.50 \pm 0.70$ a	$8.14 \pm 1.01$ a
$-1.96 \pm 0.15$	$8.30 \pm 1.20$ b	$0.76 \pm 0.05$ a	$1.10 \pm 0.30$ a	$6.10 \pm 0.20$ a	$8.21 \pm 2.70$ a

**Total electron transport rate in watered and droughted leaves:** The calculated rates of total electron flow ( $J_T$ ) decreased by 71 % with decreasing  $\Psi_w$  (Fig. 5). Water deficit had a stronger effect on  $J_C$  than on  $J_O$ . At a PFD of  $550 \mu\text{mol m}^{-2} \text{s}^{-1}$   $J_C$  decreased by 32 % with a decline of  $\Psi_w$  from  $-0.3$  to  $-2.9$  MPa, whilst  $J_O$  increased by 51 % in the same range of  $\Psi_w$  (Fig. 5). There was a positive linear relationship between  $J_T$  and  $P_N$  in control plants ( $r^2 = 0.99$ ;  $p < 0.05$ ), the relationship became curvilinear as  $\Psi_w$  declined from  $-0.3$  to  $-2.9$  MPa

(Fig. 6A). When  $P_N$  was completely suppressed at severe stress, a relatively high  $J_T$  [ $50 \mu\text{mol(e}^-) \text{m}^{-2} \text{s}^{-1}$ ] was still observed. A strong linear relationship between  $J_T$  and  $O_2$  evolution (measured at an external  $\text{CO}_2$  concentration of  $10 \text{ mmol mol}^{-1}$ ) ( $r^2 = 0.97$ ;  $p < 0.05$ ) was observed during the development of water deficit (Fig. 6B). The values of  $O_2$  evolution measured at  $10 \text{ mmol}(\text{CO}_2) \text{mol}^{-1}$  at low  $\Psi_w$  were significantly and substantially lower than in control leaves (Fig. 6B).

## Discussion

The parameters characterizing plant water status (RWC,  $\Psi_w$ ) decreased with increasing water deficit in the sunflower plants, in agreement with earlier reports (Tezara *et al.* 1999, 2002). Water deficit provoked a reduction in RWC and  $\Psi_w$ , which markedly diminished  $P_N$  and  $g_s$  (Lawlor and Cornic 2002).

The decline in  $P_N$  with decreasing  $\Psi_w$  is correlated with a reduction in  $g_s$  (Cornic 1994, 2000) which reduces diffusion of  $\text{CO}_2$  into the leaf ( $C_i$  decreases). However, in the present study,  $C_i$  remained nearly constant up to  $-3.0$  MPa, in spite of decreased  $g_s$ . Moreover, severe water deficit increased  $C_i$  possibly due to metabolic causes: respiratory metabolism dominates, and therefore  $C_i$  and compensation  $\text{CO}_2$  concentration ( $\Gamma$ ) rises (Lawlor and Cornic 2002). The ratio  $C_i/C_a$  associated with maximum  $P_N$  did not decrease during drought in sunflower suggesting metabolic impairment in accordance with an earlier report (Tezara *et al.* 1999). Similar effects of water deficit on  $C_i/C_a$  have been reported (Lawlor 1995) whereas a reduction in  $C_i/C_a$  was observed in *Prunus avium* (Centritto 2005). Precaution must be taken on the conclusions relating  $C_i/C_a$  ratio with stomatal and metabolic limitations of  $P_N$ , which in some cases may be misleading due to the erroneous calculation of  $C_i$  because of stomatal patchiness (Terashima *et al.* 1988, Buckley *et al.* 1997) or to an increase in cuticular conductance to water vapour but not to  $\text{CO}_2$  (Boyer *et al.* 1997). In addition, water deficit may cause changes in the mesophyll conductance of  $\text{CO}_2$  (Flexas *et al.* 2002, 2006, Centritto

*et al.* 2003, Centritto 2005).

Both apparent quantum yield and photon energy-saturated  $P_N$  decreased with increasing water deficit, suggesting that either energy harvesting and/or linear electron transport were affected by water deficit or that enzymatic processes were responsible for decreasing  $\text{CO}_2$  fixation or all factors functioned together. Similar results were reported in the desert shrub *Encelia farinosa* (Ehleringer and Cook 1984) and in *Lycium nodosum* (Tezara *et al.* 2003). With decreasing  $\Psi_w$ ,  $\Phi_{\text{CO}_2}$  decreased more than  $\Phi_{\text{PS}_2}$ , which decreased only at severe stress and  $F_v/F_m$  was not affected by water deficit, in agreement with other studies (Lawlor and Cornic 2002, Centritto 2005), suggesting that PS2 activity is resistant to water deficit.

The  $q_p$  under photorespiratory conditions followed  $\Phi_{\text{PS}_2}$ , decreasing only at very severe stress which indicates a higher reduction state of primary acceptors (the  $Q_A$  pool) in stressed vs. control plants. In contrast,  $q_p$  was not affected by water deficit in sunflower (Scheuermann *et al.* 1991) and in wheat (Biehler and Fock 1993, 1996).

Water deficit increased  $q_{NP}$  in plants of sunflower, indicating that a greater proportion of the energy was thermally dissipated, thus accounting for the apparent down-regulation of PS2 and supporting the protective role of the non-photochemical quenching against photoinhibition. In contrast,  $q_{NP}$  was unaffected by water deficit in wheat (Biehler and Fock 1993) and increased slightly in sunflower (Scheuermann *et al.* 1991).

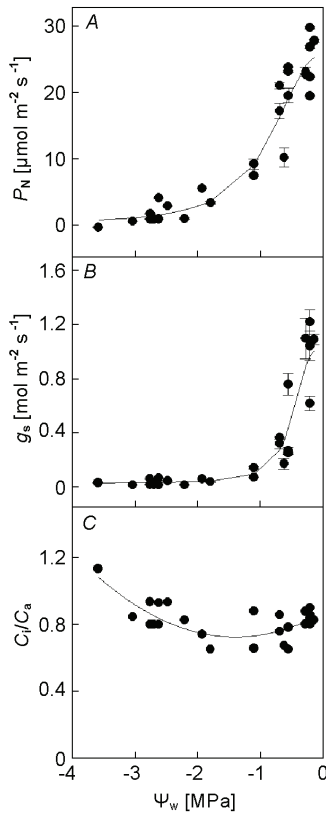


Fig. 1. Effect of leaf water potential ( $\Psi_w$ ) on (A) net photosynthetic rate ( $P_N$ ), (B) stomatal conductance ( $g_s$ ), and (C) ratio of intercellular and ambient CO<sub>2</sub> concentrations ( $C_i/C_a$ ) measured at a photon flux density (PFD) of  $1\,100\,\mu\text{mol m}^{-2} \text{s}^{-1}$ , leaf temperature of  $25\,^{\circ}\text{C}$ , O<sub>2</sub> concentration of  $0.21\,\text{mol mol}^{-1}$ , and  $C_a$  of  $350\,\mu\text{mol mol}^{-1}$ . Means  $\pm$  SE,  $n = 10$ .

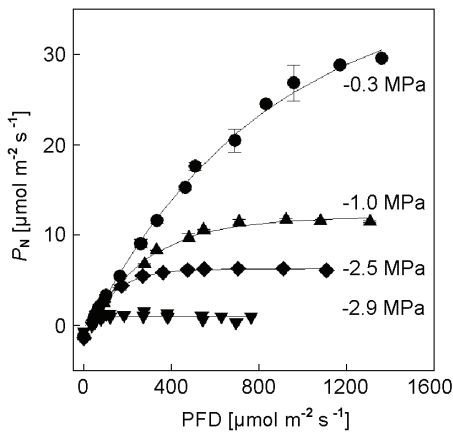


Fig. 2. Photosynthetic responses ( $P_N$ ) to photon flux density (PFD) of attached sunflower leaves at different leaf water potential ( $\Psi_w$ ). Measurements at leaf temperature of  $25\,^{\circ}\text{C}$ , O<sub>2</sub> of  $0.21\,\text{mol mol}^{-1}$ , and ambient CO<sub>2</sub> concentration ( $C_a$ ) of  $350\,\mu\text{mol mol}^{-1}$ . Means  $\pm$  SE,  $n = 6$ .

A negative relationship between  $q_{NP}$  and ATP content in sunflower suggests that increased thylakoid membrane energization was a consequence of the inhibition of either ATP generation or ATP consuming reactions, in agreement with earlier reports (Stuhlfauth *et al.* 1988, Tezara *et al.* 1999). In the intact leaf,  $q_{NP}$  consists mainly of the “energy quenching” which is related to the proton gradient across the thylakoid membrane, and affects PS2 *via* membrane energization (Schreiber *et al.* 1986, Krause

*et al.* 1988). In this study, ATP content decreased in sunflower subjected to water deficit, supporting the hypothesis of impaired metabolism and confirming an earlier report of Tezara *et al.* (1999).

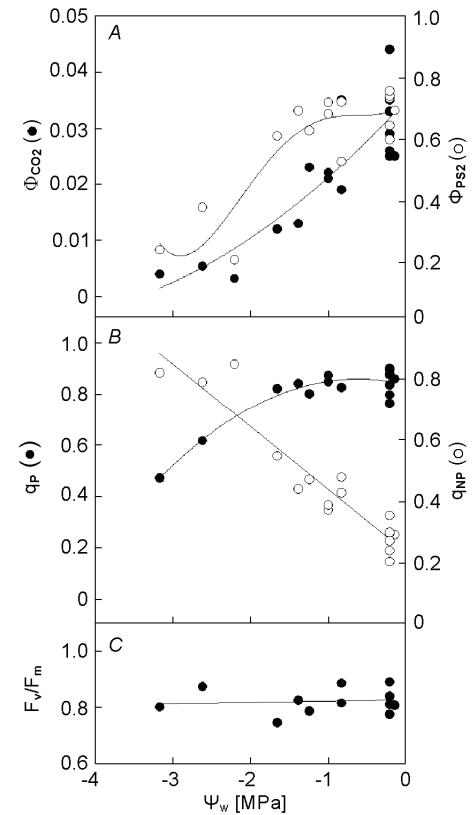


Fig. 3. Changes of fluorescence parameters with leaf water potential ( $\Psi_w$ ) measured at  $550\,\mu\text{mol m}^{-2} \text{s}^{-1}$  of PFD: (A) the relative quantum yield of photosystem 2,  $\Phi_{PS2}$  (○) and the apparent quantum yield of CO<sub>2</sub>,  $\Phi_{CO2}$  (●); (B) the photochemical ( $q_P$ ) and non-photochemical quenching coefficients ( $q_{NP}$ ) (○); and (C) the maximum quantum yield of PS2 ( $F_v/F_m$ ). Means  $\pm$  SE,  $n = 6$ .

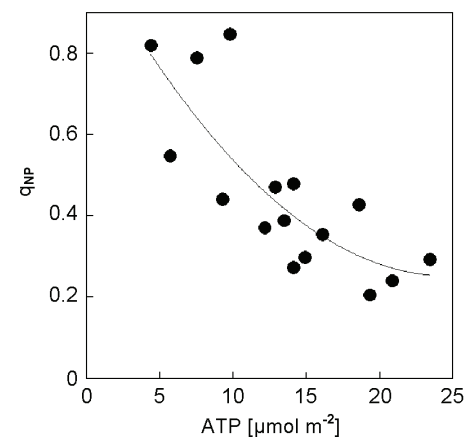


Fig. 4. Relationship between the non-photochemical quenching coefficients ( $q_{NP}$ ) measured at photon flux density (PFD) of  $550\,\mu\text{mol m}^{-2} \text{s}^{-1}$  and ATP content. Means  $\pm$  SE,  $n = 4$ .

Our results confirm that in sunflower plants, when  $P_N$  is inhibited due to water deficit, there is a fraction of PS2 centres which stays open and is able to perform photochemical work. Horton *et al.* (1994) suggested that a certain fraction of PS2 centres stays open when  $P_N$  is very low and the  $\Phi_{PS2}$  is greatly reduced by, for instance, stressful environmental conditions.

Water deficit caused a decline of  $J_T$  in sunflower plants. Lower saturated  $J_T$  and  $\Phi_{PS2}$  during water deficit were observed, suggesting that the photochemical system was down-regulated by changes in leaf water status, and that PS2 activity was resistant to water deficit and not sensitive to photoinhibition. Similarly,  $J_T$  was reduced by approximately 40 % in maize leaves subjected to water deficit (Scheuermann *et al.* 1991). The reduction in  $\Phi_{PS2}$  and  $J_T$  due to water deficit was lower than the decrease in  $P_N$ , possibly due to higher photorespiration at low  $\Psi_w$ , as suggested by Lawlor and Cornic (2002).

The observed reductions in  $\Phi_{PS2}$ ,  $J_T$ , and  $q_P$  may partly contribute with the metabolic limitations of  $P_N$  in sunflower under water deficit (Tezara *et al.* 1999, 2002). The decrease in  $J_T$  of sunflower may have contributed to the increase in metabolic limitations through a reduction in ATP and/or RuBP contents (Tezara *et al.* 1999, Lawlor and Cornic 2002).

In sunflower leaves  $J_C$  was more affected than  $J_O$  suggesting greater effect on electron flux to oxygenation and possible photorespiration increase (Tezara *et al.* 1999),

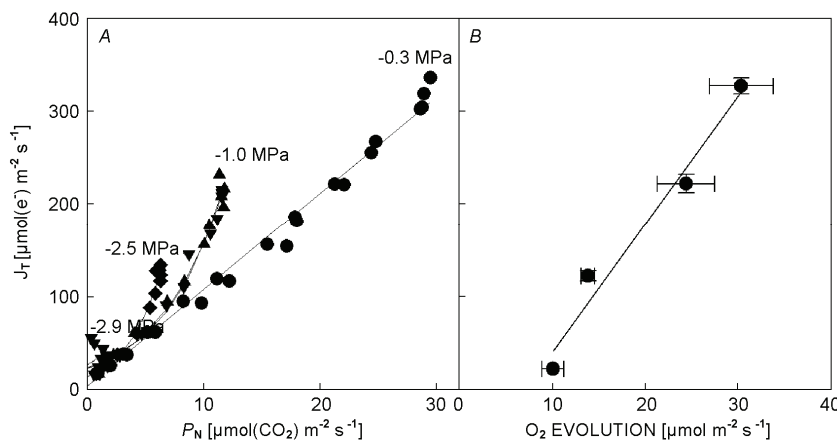


Fig. 6. Relationship between the rate of total linear electron transport ( $J_T$ ) and (A) net photosynthetic rate ( $P_N$ ) in attached leaves of sunflower at different leaf water potential and (B)  $\text{O}_2$  evolution of disc (measured at  $1 \text{ mmol mol}^{-1}$  of  $\text{CO}_2$ , PFD of  $1\,100 \mu\text{mol m}^{-2} \text{ s}^{-1}$ , and  $25^\circ\text{C}$ . Means  $\pm$  SE,  $n = 5$ ).

which supports the protective effect of  $q_{NP}$  against over-reduction and photoinhibition in sunflower under water deficit. Similar results were reported in sunflower and bean plants subjected to water deficit (Scheuermann *et al.* 1991). In contrast, the rate of photorespiration decreased with water deficit in wheat and therefore the  $\text{O}_2$  taken up in the light might be preferentially used by the Mehler-peroxidase reaction (Biehler and Fock 1996).

When  $P_N$  was totally inhibited at low  $\Psi_w$  a residual electron transport of  $50 \mu\text{mol}(e^-) \text{ m}^{-2} \text{ s}^{-1}$  could be measured. This has also been observed in water stressed plants of sorghum (Loreto *et al.* 1995). The positive linear relationship between  $J_T$  and  $P_N$  observed in control

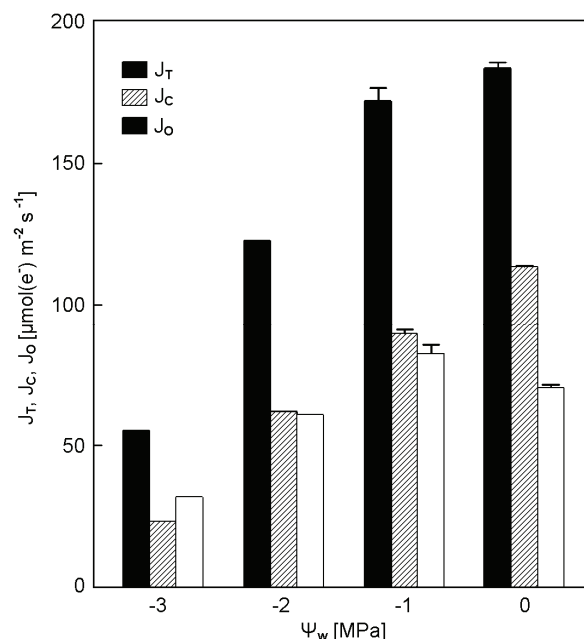


Fig. 5. Effects of leaf water potential ( $\Psi_w$ ) on the rate of total linear electron transport ( $J_T$ ), electron transports allocated to carboxylation ( $J_C$ ) or to oxygenation ( $J_O$ ) in attached leaves of sunflower. Measurements were done at  $550 \mu\text{mol m}^{-2} \text{ s}^{-1}$  of PFD, leaf temperature of  $25^\circ\text{C}$ ,  $\text{O}_2$  of  $0.21 \text{ mol mol}^{-1}$ , and ambient  $\text{CO}_2$  concentration ( $C_a$ ) of  $350 \mu\text{mol mol}^{-1}$ . Means  $\pm$  SE,  $n = 6$ .

plants became curvilinear as  $\Psi_w$  declined, indicating that  $P_N$  was more affected by water deficit than  $J_T$ . Alternative electron sinks may develop when  $P_N$  is very low.

Our results show that the reduction state of the pyridine nucleotides in stressed leaves was not significantly altered as water deficit increased while ATP/NADPH dropped substantially, indicating that carbon metabolism was more affected by ATP content than by NADPH. Similarly, NADPH content decreased only slightly during water deficit (Lawlor and Khanna-Chopra 1984, Stuhlfauth *et al.* 1991).

Measurements of  $\text{O}_2$  evolution, in which diffusion limitations were negligible, showed that  $P_N$  was not sub-

stantially increased by high CO<sub>2</sub> concentrations (10 mmol mol<sup>-1</sup>) as  $\Psi_w$  declined, supporting that dehydrated leaves suffered from more than stomatal limitation. In other plant species, 50–150 mmol(CO<sub>2</sub>) mol<sup>-1</sup> may be required to fully saturate the photosynthetic machinery (Chaves 1991, Quick *et al.* 1992, Cornic 1994).

We conclude that the declines in  $P_N$ ,  $g_s$ , and ATP content associated with a nearly constant  $C_i/C_a$  ratio indicated metabolic impairment in sunflower. During

severe water deficit, the increase in  $q_{NP}$  allowed a greater proportion of the energy to be thermally dissipated. The lower saturated  $J_T$  and  $\Phi_{PS2}$  suggested that the photochemical system was down-regulated, and that leaves were not sensitive to photoinhibition. Changes observed in photochemical activity of PS2 can partly contribute to the metabolic limitations of  $P_N$  in sunflower under water deficit.

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