

## Responses of cotton and wheat photosynthesis and growth to cyclic variation in carbon dioxide concentration

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### Abstract

The carbon dioxide concentration in free air carbon dioxide enrichment (FACE) systems typically has rapid fluctuations. In our FACE system, power spectral analysis of CO<sub>2</sub> concentration measured every second with an open path analyzer indicated peaks in variation with a period of about one minute. I used open-top chambers to expose cotton and wheat plants to either a constant elevated CO<sub>2</sub> concentration of 180  $\mu\text{mol mol}^{-1}$  above that of outside ambient air, or to the same mean CO<sub>2</sub> concentration, but with the CO<sub>2</sub> enrichment cycling between about 30 and 330  $\mu\text{mol mol}^{-1}$  above the concentration of outside ambient air, with a period of one minute. Three short-term replicate plantings of cotton were grown in Beltsville, Maryland with these CO<sub>2</sub> concentration treatments imposed for 27-day periods over two summers, and one winter wheat crop was grown from sowing to maturity. In cotton, leaf gas-exchange measurements of the continuously elevated treatment and the fluctuating treatment indicated that the fluctuating CO<sub>2</sub> concentration treatment consistently resulted in substantial down-regulation of net photosynthetic rate ( $P_N$ ) and stomatal conductance ( $g_s$ ). Total shoot biomass of the vegetative cotton plants in the fluctuating CO<sub>2</sub> concentration treatment averaged 30% less than in the constantly elevated CO<sub>2</sub> concentration treatment at 27 days after planting. In winter wheat, leaf gas-exchange measurements also indicated that down-regulation of  $P_N$  and  $g_s$  occurred in flag leaves in the fluctuating CO<sub>2</sub> concentration treatment, but the effect was not as consistent in other leaves, nor as severe as found in cotton. However, wheat grain yields were 12% less in the fluctuating CO<sub>2</sub> concentration treatment compared with the constant elevated CO<sub>2</sub> concentration treatment. Comparison with wheat yields in chambers without CO<sub>2</sub> addition indicated a nonsignificant increase of 5% for the fluctuating elevated CO<sub>2</sub> concentration treatment, and a significant increase of 19% for the constant elevated treatment. The results suggest that treatments with fluctuating elevated CO<sub>2</sub> concentrations could underestimate plant growth at projected future atmospheric CO<sub>2</sub> concentrations.

*Additional key words:* acclimation; down-regulation; stomatal conductance.

### Introduction

FACE systems have some advantages over other enrichment systems for exposing crop plants to anticipated future atmospheric concentrations of CO<sub>2</sub>. One advantage is the absence of enclosures which alter wind speed, radiation, temperature, and humidity. The long-term average CO<sub>2</sub> concentration enrichment achieved in FACE systems can be very consistent, and 1-min averages of daytime CO<sub>2</sub> concentration in FACE systems are generally within 10% of the target CO<sub>2</sub> concentration 80 to 90% of the time. However, large rapid fluctuations in CO<sub>2</sub> concentration often occur (Hendrey *et al.* 1999, Okada *et al.* 2001, Bunce 2011). The importance of these rapid fluctuations in CO<sub>2</sub> concentration to plant function remains uncertain.

From leaf chlorophyll fluorescence measurements on wheat leaves Hendrey *et al.* (1997) concluded that fluctuations in CO<sub>2</sub> concentration with periods of less than one minute were unlikely to affect photosynthesis. However, Holtum and Winter (2003) measured  $P_N$  and found significantly lower mean rates when the CO<sub>2</sub> concentration varied with a period of 40 s compared to rates measured at a constant mean CO<sub>2</sub> concentration. In the experiments described here I tested whether the long-term growth and  $P_N$  of cotton and wheat plants were affected by 1-min cycles of CO<sub>2</sub> concentration.

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*Abbreviations:*  $C_i$  – CO<sub>2</sub> concentration in the substomatal (intercellular) airspace; FACE – free air carbon dioxide enrichment;  $g_s$  – stomatal conductance;  $P_N$  – net photosynthetic rate.

*Acknowledgements:* Dr. Bruce Kimball suggested the use of open-top chambers and a solenoid valve system to achieve the cyclic elevated carbon dioxide treatment.

## Materials and methods

**Design criteria:** An open path CO<sub>2</sub> analyzer (LI-7500, LI-Cor, Inc., Lincoln, NE, USA) operating at 5 Hz mounted at canopy height near the center of an area distributed FACE plot (Bunce 2011) was used to record CO<sub>2</sub> concentration once per second for two hour periods on two days. Four independent sequences of 1,000-s duration were randomly selected from this data set, and time series analysis (JMP v. 5.1, SAS Institute, NC, USA) was used to develop power spectra for each 1,000-s sequence. All four power spectra had distinct peaks at periods of approximately 20 to 80 s, and all samples had a large peak very near 60 s (Fig. 1). Based on these observations, it was decided to use a period of 60 s for the fluctuating CO<sub>2</sub> concentration treatments. The daytime CO<sub>2</sub> concentration in our FACE system, when operated with a mean enrichment of 1.4 times ambient, was frequently little enriched above ambient, but was only seldom enriched to more than twice the ambient concentration (Bunce 2011, Fig. 3). For this study, a system was designed to expose plants to CO<sub>2</sub> concen-

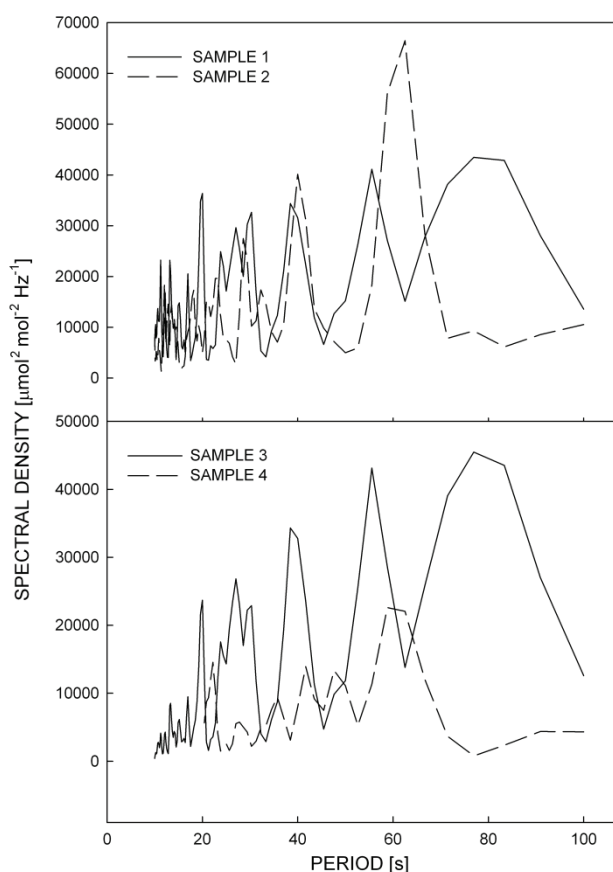


Fig. 1. Power spectra of CO<sub>2</sub> concentrations for four randomly selected 1,000-s periods for an area distributed free air carbon dioxide enrichment system (Bunce 2011). CO<sub>2</sub> concentrations were recorded at 1 Hz from an open path analyzer operating at 5 Hz.

trations ranging from about 30 to 330 μmol mol<sup>-1</sup> above the concentration of outside air, with a period of one minute. This “fluctuating” elevated CO<sub>2</sub> concentration treatment was compared with a “constant” enrichment of 180 μmol mol<sup>-1</sup> above the concentration of the outside air, which averaged 370 μmol mol<sup>-1</sup> in the daytime.

**CO<sub>2</sub> control:** A solenoid valve was placed in the line supplying CO<sub>2</sub> to an open-top chamber equipped with a blower injecting air and CO<sub>2</sub> into the bottom of the chamber through a 10 cm diameter perforated plastic pipe running the whole length of the center of the chamber. For the fluctuating CO<sub>2</sub> concentration treatment, the flow rate of CO<sub>2</sub> to the inlet of the blower was twice the normal rate, and the solenoid valve was turned off for the first 30 s of every minute with an electronic timer. The range of CO<sub>2</sub> concentration achieved in the chamber was dependent on the air-volume turnover time of the chamber, which was nominally 0.6 min in both sizes of open-top chambers used in these experiments. Representative time courses of CO<sub>2</sub> concentration measured with the open path analyzer in “fluctuating” and “constant” elevated CO<sub>2</sub> concentration chambers, and in chambers with no CO<sub>2</sub> added are presented in Fig. 2. The sensor of the open path analyzer was mounted horizontally near the centers of the chambers, at the height of the upper canopy leaves, when plants were about 50 cm in height. This pattern of exposure with CO<sub>2</sub> concentration gradually oscillating between minimum and maximum values is similar to that of Holtum and Winter (2003), and is different from the pattern of sharp transitions between minimum and maximum exposures of Cardon *et al.* (1994) and Hendrey *et al.* (1997).

**Cotton** (*Gossypium hirsutum* L., var. Delta Pine 555) seeds were planted in four square open-top chambers each covering 1.9 m<sup>2</sup> of ground. The chambers were 2 m in height, and the walls were clear acrylic plastic. Two of the chambers had constantly elevated CO<sub>2</sub> concentrations and two had fluctuating elevated CO<sub>2</sub> concentrations, with CO<sub>2</sub> added 24 h per day. These treatments were rotated among chambers in three replicate runs over two summers. In the third replicate run, an additional two chambers were planted with cotton, but were operated with no CO<sub>2</sub> addition. Cotton was planted on days 197 and 236 in 2010, and 201 in 2011. The soil of the field plot containing the chambers was a Codorus silt loam, a fine-loamy, mixed, mesic Fluvaquentic Dystrochrept soil, and the prior crop was *Phaseolus vulgaris* grown with a 10-10-10, N, P and K fertilizer. No fertilizer was added to the soil for the cotton. Cotton seedlings were thinned to 24 plants per chamber, in two border and two interior rows. Plants sampled for leaf gas exchange and biomass were from the center of the interior rows of each chamber, *i.e.* bordered by other cotton plants on all sides.

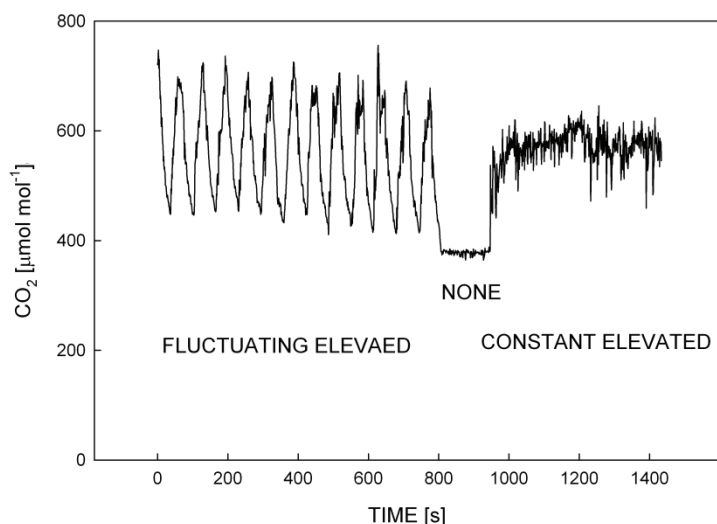


Fig. 2. Time sequences of measurements of CO<sub>2</sub> concentration in three CO<sub>2</sub> treatment chambers: fluctuating elevated, no added CO<sub>2</sub>, and constant elevated. CO<sub>2</sub> concentrations were recorded at 1 Hz from an open path analyzer operating at 5 Hz. Mean concentrations were 559, 378, and 569  $\mu\text{mol mol}^{-1}$  for the three treatments, respectively.

Air samples from each chamber were pumped to an infrared CO<sub>2</sub> analyzer in a nearby shelter, and mean CO<sub>2</sub> concentration of air from each chamber, and the CO<sub>2</sub> concentration of outside air were each recorded once per hour. CO<sub>2</sub> flow rates to each chamber were adjusted daily, as necessary. Mean air temperatures for the three replicate experiments were 26.5, 21.4, and 26.3°C. The plot was not irrigated, but no significant soil water deficits occurred.

Leaf gas exchange was measured on four dates, one date for each of the two 2010 crops, and on two dates in the 2011 crop. In 2010, leaf gas-exchange measurements were made 26 d after planting. In 2011, leaf gas-exchange measurements were made at 23 and 26 d after planting. Leaf gas exchange was measured near mid-day on clear days, using a *CIRAS-1* portable photosynthesis system (*PP-Systems*, Amesbury, Massachusetts, USA) with CO<sub>2</sub> concentration control. Two mature upper canopy leaves from each open-top chamber were measured at the ambient air temperature and water vapor content, in full sunlight on each occasion. Leaves from chambers with constant elevated CO<sub>2</sub> concentration were measured only at the growth CO<sub>2</sub> concentrations, but leaves from the fluctuating elevated CO<sub>2</sub> concentration were measured at the average minimum, mean, and maximum CO<sub>2</sub> concentration during growth. In the 2011 crop, leaves from chambers without added CO<sub>2</sub> were measured at the same three CO<sub>2</sub> concentrations as leaves from the fluctuating elevated CO<sub>2</sub> concentration chambers. In cases where measurements were to be made at three CO<sub>2</sub> concentrations, the first measurement was made at the mean growth CO<sub>2</sub> concentration. No significant change in stomatal conductance ( $g_s$ ) with measurement CO<sub>2</sub> concentration occurred during these measurements, probably because leaves were kept at the different CO<sub>2</sub> concentration only long enough (about a minute) for  $P_N$  to become stable. Whole shoots of eight plants per chamber

were harvested 27 d after sowing to determine shoot dry mass.

**Winter wheat** (*Triticum aestivum* L., var. Choptank) was planted on day of year 282 in 2010 in twelve rectangular open-top chambers, each covering 2.8 m<sup>2</sup> of ground. The chamber height was 2.5 m, and the chamber walls were clear acrylic plastic. There were four rows of plants per chamber, with 30 cm between rows. The soil was the same as described for the cotton experiments, and 60 g of urea was added to each chamber when wheat growth resumed in the spring. Three CO<sub>2</sub> concentration treatments, constant elevated CO<sub>2</sub> concentration, fluctuating CO<sub>2</sub> concentration, and no added CO<sub>2</sub> were randomly assigned to each of four chambers. CO<sub>2</sub> was added 24 h per day, except when the ground was covered with snow. Air from each chamber was pumped to an infrared CO<sub>2</sub> analyzer and mean CO<sub>2</sub> concentration from each chamber was recorded once an hour, with CO<sub>2</sub> flow rates adjusted daily, as necessary.

Leaf gas-exchange measurements, as described for the cotton experiment, were conducted on ten days with wheat, once in the fall of 2010, and nine times in spring, which included four days of measurements on flag leaves. A harvest of two plants from border rows in each of the four corners of each chamber was made 44 d after sowing in 2010, before shoots were damaged by low winter temperatures. At crop maturity in June 2011, 3 m of interior rows were harvested from each chamber to determine shoot and seed dry mass.

**Statistics:** Chambers were treated as the experimental units. Treatments were compared using *ANOVA*, except that leaf gas exchange in wheat was analyzed using repeated measures *ANOVA*, because the same experimental units were measured on multiple occasions.

## Results

**Cotton:** For comparisons of the two elevated CO<sub>2</sub> concentration treatments, the measurement date affected  $P_N$  and  $g_s$ , but there was no significant interaction between measurement date and CO<sub>2</sub> concentration treatment for  $P_N$  or  $g_s$ . Mean values for the four measurement dates (Table 1) indicated substantially lower  $P_N$  and  $g_s$ , but similar substomatal carbon dioxide concentrations ( $C_i$ ) for the plants grown with fluctuating elevated CO<sub>2</sub> concentration compared with constant elevated CO<sub>2</sub> concentration.

For the 2011 data on leaf gas exchange, plants grown without added CO<sub>2</sub> had higher rates of  $P_N$  than plants grown with fluctuating elevated CO<sub>2</sub> concentration when measured at the lower, but not at the higher CO<sub>2</sub> concentration, and low  $g_s$  when measured at both CO<sub>2</sub> concentrations (Table 2).  $C_i$  during these measurements did not differ significantly with growth conditions (Table 2). When measured at the mean elevated CO<sub>2</sub> concentration, plants grown without added CO<sub>2</sub> and those grown with constant elevated CO<sub>2</sub> concentration had mean  $P_N$  of 35.3 and 34.0 mmol m<sup>-2</sup> s<sup>-1</sup>, respectively, and  $g_s$  of 668 and 574 mmol m<sup>-2</sup> s<sup>-1</sup>, neither of which was different at  $P=0.05$ .

Averaged over the three replicate runs, shoot dry mass per plant averaged 1.31 g for the constant elevated CO<sub>2</sub>

concentration treatment and 0.92 g for the fluctuating elevated CO<sub>2</sub> concentration treatment at 27 days after planting. These means were statistically different at  $P=0.05$ . In 2011, shoot dry mass for the constant elevated CO<sub>2</sub> concentration treatment was 1.47 times that of the treatment without added CO<sub>2</sub>, compared with 1.25 times for the fluctuating elevated CO<sub>2</sub> concentration treatment. Mean masses in the three treatments were all different from each other at  $P=0.05$  in 2011.

Table 1. Mean net photosynthetic rate ( $P_N$ ), stomatal conductance ( $g_s$ ), and substomatal concentration of carbon dioxide ( $C_i$ ) of leaves of cotton plants grown with fluctuating and constant elevated carbon dioxide concentrations. Values are averaged over four measurement dates, and leaves were measured at the mean daytime elevated CO<sub>2</sub> concentration of 550  $\mu\text{mol mol}^{-1}$ . Values within columns followed by *different letters* differed between CO<sub>2</sub> concentration treatments at  $P=0.05$ , using ANOVA.

Treatment	$P_N$ [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ]	$g_s$ [mmol m <sup>-2</sup> s <sup>-1</sup> ]	$C_i$ [ $\mu\text{mol mol}^{-1}$ ]
Constant	41.8 <sup>A</sup>	1180 <sup>A</sup>	418 <sup>A</sup>
Fluctuating	34.5 <sup>B</sup>	745 <sup>B</sup>	413 <sup>A</sup>
Ratio (F/C)	0.83	0.69	0.99

Table 2. Mean net photosynthetic rate ( $P_N$ ), stomatal conductance ( $g_s$ ), and substomatal concentration of carbon dioxide ( $C_i$ ) of cotton plants grown with no added CO<sub>2</sub> or with a fluctuating elevated CO<sub>2</sub> concentration. Values are averaged over two 2011 measurement dates, and leaves were measured at two CO<sub>2</sub> concentrations. Values within columns followed by *different letters* differed between growth CO<sub>2</sub> concentration treatments at  $P=0.05$ , using ANOVA.

Measurement CO <sub>2</sub> [ $\mu\text{mol mol}^{-1}$ ]	$P_N$ [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ]		$g_s$ [mmol m <sup>-2</sup> s <sup>-1</sup> ]		$C_i$ [ $\mu\text{mol mol}^{-1}$ ]	
	370	730	370	730	370	730
Treatment						
None	25.0 <sup>A</sup>	42.1 <sup>A</sup>	683 <sup>A</sup>	633 <sup>A</sup>	249 <sup>A</sup>	533 <sup>A</sup>
Fluctuating	19.9 <sup>B</sup>	38.8 <sup>A</sup>	493 <sup>B</sup>	456 <sup>B</sup>	248 <sup>A</sup>	537 <sup>B</sup>
Ratio (F/N)	0.80	0.92	0.72	0.72	1.00	1.01

Table 3. Mean photosynthetic rate ( $P_N$ ), stomatal conductance ( $g_s$ ), and substomatal concentration of carbon dioxide ( $C_i$ ) of flag leaves of wheat plants grown without added CO<sub>2</sub>, and with fluctuating or constant elevated CO<sub>2</sub> concentration. Values are averaged over four measurement dates, and leaves were measured at the mean daytime elevated CO<sub>2</sub> concentration of 550  $\mu\text{mol mol}^{-1}$ . Values within columns followed by *different letters* differed between CO<sub>2</sub> concentration treatments at  $P=0.05$ , using ANOVA.

Treatment	$P_N$ [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ]	$g_s$ [mmol m <sup>-2</sup> s <sup>-1</sup> ]	$C_i$ [ $\mu\text{mol mol}^{-1}$ ]
None	39.4 <sup>A</sup>	1338 <sup>A</sup>	472 <sup>A</sup>
Constant	37.1 <sup>B</sup>	1180 <sup>B</sup>	477 <sup>A</sup>
Fluctuating	34.4 <sup>C</sup>	1037 <sup>C</sup>	467 <sup>A</sup>

**Wheat:** The leaf gas-exchange data as a whole showed significant effects of treatment, date, and treatment by date interactions for both  $P_N$  and  $g_s$ . However, restricting the statistical analysis to measurements on flag leaves (four dates) the treatment by date interaction term became nonsignificant for both gas-exchange parameters. For flag leaves,  $P_N$  and  $g_s$  measured at the mean elevated CO<sub>2</sub> concentration was highest in leaves from chambers without CO<sub>2</sub> addition, intermediate in leaves from chambers with constant elevated CO<sub>2</sub> concentration, and lowest in leaves from the fluctuating elevated CO<sub>2</sub> concentration chambers (Table 3). Similar patterns and magnitudes of treatment effects also occurred on some other measurement dates, but there were also measurement dates when no significant treatment effects

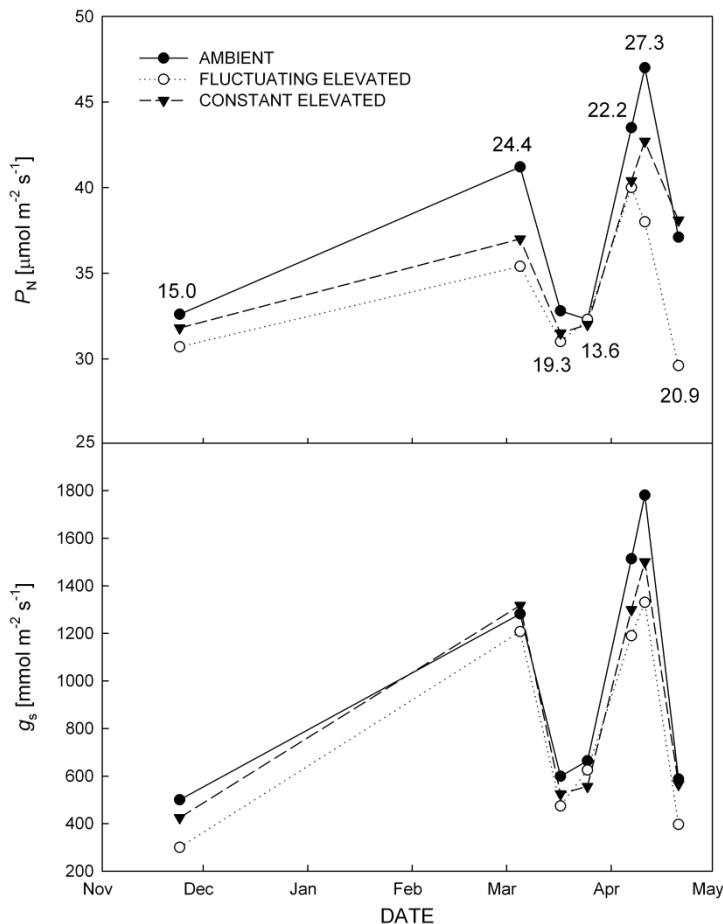


Fig. 3. Mean values of leaf net photosynthetic rate ( $P_N$ ) and stomatal conductance ( $g_s$ ) in leaves of vegetative wheat plants on different measurement dates, for three CO<sub>2</sub> treatments. Measurements were made at 370  $\mu\text{mol mol}^{-1}$  for the “ambient” CO<sub>2</sub> concentration treatment and at 550  $\mu\text{mol mol}^{-1}$  for the “elevated” CO<sub>2</sub> concentration treatments. Numbers indicate mean leaf temperatures during the measurements on each date. The treatment, date, and treatment by date interaction terms were all significant at  $P=0.05$  for both  $P_N$  and  $g_s$ , using repeated measures ANOVA.

Table 4. Harvest data for winter wheat crops grown in open-top chambers without added CO<sub>2</sub>, with constant elevated CO<sub>2</sub> concentration, and with fluctuating elevated CO<sub>2</sub> concentration. Values are means for four chambers per treatment. Values within columns followed by different letters differed between growth CO<sub>2</sub> concentration treatments at  $P=0.05$ , using ANOVA. DM – dry mass.

Treatment	At day 44 Total shoot [g(DM) plant <sup>-1</sup> ]	At crop maturity Total shoot [g(DM) m <sup>-2</sup> ]	Seed [g(DM) m <sup>-2</sup> ]
None	0.98 <sup>C</sup>	373 <sup>B</sup>	125 <sup>B</sup>
Constant	1.43 <sup>A</sup>	445 <sup>A</sup>	149 <sup>A</sup>
Fluctuating	1.20 <sup>B</sup>	395 <sup>B</sup>	131 <sup>B</sup>

## Discussion

The results of this study indicated that the cyclically varying elevated CO<sub>2</sub> concentration treatment reduced the long-term growth of both species compared with a more constant CO<sub>2</sub> concentration treatment with the same mean CO<sub>2</sub> concentration. Lower  $P_N$  also occurred, and could have been responsible for the slower dry matter production. As pointed out by Hendrey *et al.* (1997) and Holtum and Winter (2003), because  $P_N$  has a saturating response to CO<sub>2</sub> concentration, mean  $P_N$  would always be less for

occurred, without any obvious relationship with growth stage or measurement temperature (Fig. 3).

The dry mass of shoots of wheat plants at an early vegetative stage of development (44 days after sowing) was highest in the constant elevated CO<sub>2</sub> concentration treatment and lowest in the treatment without added CO<sub>2</sub> (Table 4). At maturity, total shoot mass and seed mass per ground area were significantly higher for the constant elevated CO<sub>2</sub> concentration treatment than either the fluctuating elevated CO<sub>2</sub> concentration treatment or the treatment without CO<sub>2</sub> addition, which did not differ significantly from each other (Table 4).

leaves exposed only to the maximum and minimum CO<sub>2</sub> concentration than for those exposed only to the mean concentration. In both our and the Holtum and Winter (2003) fluctuating CO<sub>2</sub> concentration treatments, plants were exposed to the whole range of concentrations (Fig. 2), not just to the minimum and maximum, as in the Cardon *et al.* (1994) and the Hendrey *et al.* (1997) treatments, so the anticipated effect on mean  $P_N$  would be smaller. From the frequency distribution of exposure to

different CO<sub>2</sub> concentration, in combination with the observed CO<sub>2</sub> concentration response curves, assuming an instantaneous response of  $P_N$  to CO<sub>2</sub> concentration leads to an estimated reduction in mean  $P_N$  of only  $3 \pm 1\%$  for both cotton and wheat due to this direct effect of variation in CO<sub>2</sub> concentration. In FACE systems which expose plants intermittently to much higher CO<sub>2</sub> concentrations than used here, this effect would be larger, because very high CO<sub>2</sub> concentrations would increase mean CO<sub>2</sub> concentration but have little additional effect on  $P_N$ . In these experiments in open-top chambers, the observed down-regulation of  $P_N$  was far more important in reducing  $P_N$  in the fluctuating CO<sub>2</sub> concentration treatments, especially in cotton, than was this effect due to the curvilinear photosynthetic response.

This is the first report to document down-regulation of  $P_N$  in response to long-term exposure to fluctuating CO<sub>2</sub> concentrations. Both Holtum and Winter (2003) and Hendrey *et al.* (1997) exposed plants to fluctuating CO<sub>2</sub> concentrations only for several minutes. The larger down-regulation of  $P_N$  at lower measurement CO<sub>2</sub> concentration than at high CO<sub>2</sub> concentration documented here in cotton suggests a larger reduction in carboxylation capacity than in RuBP regeneration capacity, because carboxylation capacity is generally limiting at low measurement CO<sub>2</sub> concentrations, and regeneration capacity

becomes limiting at high CO<sub>2</sub> concentrations. Lack of change in  $C_i$  with growth CO<sub>2</sub> concentration treatment suggests that the treatment effects on  $g_s$  were a response to changes in  $P_N$ , rather than the reverse (Bounoua *et al.* 1999). Cardon *et al.* (1994) reported that fluctuations in CO<sub>2</sub> concentration disrupted  $g_s$ , but did not report responses to fluctuations with periods as short as the 1-min cycle used in these experiments.

Although the period of the cyclic variation in CO<sub>2</sub> concentration used here was based on periods observed in a FACE system, FACE systems also expose plants to a much wider range of CO<sub>2</sub> concentration and to more abrupt changes in CO<sub>2</sub> concentration than used here (Hendrey *et al.* 1999, Bunce 2011). The large effects of this limited magnitude, cyclic variation in CO<sub>2</sub> concentration observed here on plant biomass production in both cotton and wheat could conceivably be larger or smaller than possible effects of the more variable fluctuations in CO<sub>2</sub> concentration occurring in FACE systems. Season-long, side-by-side comparisons of plant growth responses to elevated CO<sub>2</sub> concentration in FACE and open-top chambers, like the shorter study by Kimball *et al.* (1997), would be worthwhile, as well as other efforts to evaluate and understand the impacts of rapid fluctuations in CO<sub>2</sub> concentration on plants.

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