

## Short- versus long-term effects of elevated CO<sub>2</sub> on night-time respiration of needles of Scots pine (*Pinus sylvestris* L.)

M.E. JACH\* and R. CEULEMANS

*Department of Biology, University of Antwerpen (UIA), Universiteitsplein 1, B-2610 Wilrijk, Belgium*

### Abstract

Dark respiration rate in the night ( $R_D$ ) was measured in five-year-old Scots pine (*Pinus sylvestris* L.) trees grown for two years under ambient (AC) and elevated (AC + 400  $\mu\text{mol mol}^{-1}$  = EC) CO<sub>2</sub> concentrations in open top chambers. Two needle age classes (*i.e.*, current-year and one-year-old) were measured at AC and EC in both AC- and EC-grown pines. Additionally different chemical characteristics were determined on the needles, such as nitrogen (N), carbon (C), starch, and soluble sugar concentrations as well as specific leaf area. The direct, short-term and indirect, long-term effects of EC on  $R_D$  for the two needle age classes were examined.  $R_D$  was expressed on a per needle area, needle mass, N, C, and C/N bases. Direct effects were only pronounced in the AC treatment where inhibition of  $R_D$  was found at EC in both current- and one-year-old needles. Indirect effects were only significant in one-year-old needles where a decrease was found in the EC grown trees as compared with AC ones when  $R_D$  was expressed per unit needle mass, C, or C/N.  $R_D$  per unit needle area and needle N were not sensitive to long-term EC, in any needle age class. Long-term EC treatment also influenced the response of the two needle age classes. One-year-old needles from the EC treatment had significantly lower  $R_D$  than current-year needles, but no such response was observed in the AC treatment. Our experiment re-emphasised the importance of expressing  $R_D$  on different bases for a correct interpretation of the responses to EC. Moreover, we showed that different needle age classes can respond differently to a CO<sub>2</sub> enrichment.

*Additional key words:* carbon; C/N; direct and indirect effects; nitrogen; soluble sugars; starch; specific leaf area.

### Introduction

Respiration in trees may consume half or more of the carbon fixed in photosynthesis (Farrar 1985, Amthor 1995). Moreover, leaf  $R_D$  in trees may constitute more than half of the total whole-plant respiration (Hagihara and Hozumi 1991) and changes in this component in response to atmospheric CO<sub>2</sub> enrichment could have a significant effect on forest carbon budgets. On a global scale, about 10 times more CO<sub>2</sub> is released by plant respiration as is released by fossil fuel burning; therefore any effects of increasing CO<sub>2</sub> on the amount of CO<sub>2</sub> released to the atmosphere by plant respiration are important to the global C cycling (Amthor 1995). Given the importance of plant respiration, it is important to

improve our knowledge of the effects of rising atmospheric CO<sub>2</sub> concentration on respiratory carbon release.

While the effects of atmospheric CO<sub>2</sub> on photosynthesis and photorespiration are well documented (Gunderson and Wullschlegel 1994, Norby *et al.* 1999, Saralabai *et al.* 1997), much less is known about the response of dark respiration. The measurements of  $R_D$  are, however, essential to extrapolate the gas exchange measurements to biomass production (Mousseau and Saugier 1992). It is useful to distinguish direct and indirect effects in identifying mechanisms by which EC might affect respiration (Amthor 1991, 1995, Ryan

Received 28 December 1999, accepted 29 December 1999.

\*Fax: 32-3-8202271, e-mail: rceulem@uia.ua.ac.be

**Acknowledgements:** This research was supported by the EC through its Environment R&D Programme under contract n° ENV4-CT95-0077 for "Research on the Likely Impact of Rising CO<sub>2</sub> and Temperature on European Forests" (co-ordinated by the University of Edinburgh, Scotland) and by the Belgian Prime Minister's Office, Federal Services for Scientific, Technical and Cultural Affairs (contract n° GC/DD/05B). The authors greatly acknowledge B. De Cuyper and J. Van Slycken (Institute for Forestry and Game Management, Belgium) for providing the pine seedlings, A. Grey and P.G. Jarvis (University of Edinburgh, Scotland) for sugar analysis, E. De Bruyn for starch analysis, R. Jach for help with field measurements, N. Calluy and F. Kockelbergh for technical assistance. This study contributes to the Global Change and Terrestrial Ecosystems (GCTE) core project of the International Geosphere Biosphere Programme (IGBP).

1991, Wullschleger *et al.* 1994). Only few studies, however, have examined both direct and indirect effects of EC on  $R_D$  so far (Bunce 1990, Thomas and Griffin 1994, Wullschleger *et al.* 1994, Griffin *et al.* 1996, Amthor 1997, Marek *et al.* 1997, Will and Ceulemans 1997, Urban and Marek 1999).

Mechanisms underlying short-term and long-term reductions in respiratory  $CO_2$  are believed to be different and are still the subject of discussion. Leaf  $R_D$ 's may be directly and reversibly inhibited by EC (Bunce and Caulfield 1991, El Kohen *et al.* 1991, Bunce 1994, 1995, Wullschleger *et al.* 1994, Ziska and Bunce 1994, Griffin *et al.* 1996). Direct effects of  $CO_2$  on  $R_D$  are short-term, resulting in a depression of the apparent  $R_D$  of leaf tissue with instantaneous changes in  $CO_2$  concentration (Drake *et al.* 1999). Direct effects might involve specific interactions of  $CO_2$  inside a leaf with regulatory enzymes of the respiratory pathway (Amthor 1997). The direct effect of  $CO_2$  on  $R_D$  is in particular important in the case of forest canopies, where daily and seasonal fluctuations can be large at the forest floor, depending on soil activity and canopy structure (Bazzaz and Williams 1991).

In addition to direct effects,  $R_D$  values may be altered through a longer-term acclimation response to EC (Bunce 1994, Wullschleger *et al.* 1994, Norby *et al.* 1999). Indirect effects of  $CO_2$  on  $R_D$  are due to long-term EC specific changes in tissue quality and construction costs (Poorter *et al.* 1992, Curtis 1996, Poorter *et al.* 1997, Wullschleger *et al.* 1997). EC generally enhances photosynthesis (Gunderson and Wullschleger 1994, Ceulemans and Mousseau 1994, Saxe *et al.* 1998) stimulating the levels of soluble saccharides (Barnes *et al.* 1995). Enhanced levels of soluble saccharides can potentially increase respiration by stimulating growth and different processes consuming respiratory products. In contrast, decreased concentrations of soluble protein (Ziska and Bunce 1994) may affect the specific maintenance respiration in a negative way. Maintenance respiration seems to be more sensitive to  $CO_2$  concentration than growth respiration (Ryan 1991). Moreover, since maintenance respiration is strongly related to N concentration of the tissue (Ryan 1991), it is expected to decline with EC, because N concentration is

generally lower than in AC leaves. To better explain the results, construction and maintenance respiration components of respiration are frequently reported (Ryan 1991, Sprugel and Benecke 1991).

There is a variability in the way  $R_D$  is determined: from response to photosynthetically active radiation (PAR) (Nijs *et al.* 1988, Wang *et al.* 1995, Kubiske and Pregitzer 1996), from  $CO_2$  response, or by darkening leaves during the day (Ziska *et al.* 1990, Teskey 1995). However, the best estimate of  $R_D$  is obtained when measured at night because the impact of sugar synthesis and transport is then minimised (Amthor 1989, Will and Ceulemans 1997). This is of particular relevance in EC experiments, where C assimilation and consequently sugar production are higher. At night, however, stomatal closure and low night temperatures may restrict the  $CO_2$  evolution from needles (Sprugel *et al.* 1995).

Studies on the effects of EC on respiration have yielded contrasting results. The response of respiration may differ among species, among studies, and likely among plant tissues. Leaf or needle  $R_D$  per dry mass may either increase (Gifford *et al.* 1985, Hrubec *et al.* 1985, Williams *et al.* 1992, Thomas *et al.* 1993, Thomas and Griffin 1994), or decrease (Bunce and Caulfield 1991, Wullschleger and Norby 1992, Bunce 1992, 1995, Williams *et al.* 1992, Wullschleger *et al.* 1992, Ziska and Bunce 1993, Azcón-Bieto *et al.* 1994, Teskey 1995, Wang *et al.* 1995, Kubiske and Pregitzer 1996), or remain constant (Gifford *et al.* 1985, Baker *et al.* 1992, Hrubec *et al.* 1985, Ceulemans and Mousseau 1994) under EC.

The objectives of this investigation were to examine the direct and indirect effects of a doubled  $CO_2$  concentration on the night-time respiration of Scots pine needles. We examined whether needle  $R_D$  is altered on the short- or long-term by EC, and in which direction. Explanations of the results have been sought by measuring needle chemical characteristics such as N, C, soluble sugar, and starch concentrations, and morphological characteristics such as specific leaf area (SLA).  $R_D$  was expressed on the bases of needle area, needle mass, N, C, and C/N, so that comparisons could be made.

## Materials and methods

**Plants and growth conditions:** Three-years-old, pot-grown and dormant Scots pine (*Pinus sylvestris* L.) seedlings obtained from the Institute for Forestry and Game Management in Groenendaal (Belgium), were planted in the ground in four open top fumigation chambers (OTC) situated on the campus of the University of Antwerp (UIA), on 21 March 1996. Mean annual

temperature and rainfall at the site are 11.98 °C and 769 mm, respectively. All seedlings were from the same Belgian provenance (south from Samber and Maas) and were about 0.4 m tall at the time of planting. Eleven trees per chamber were planted in a circular planting pattern, 70 cm apart from each other and from the walls. To reduce the boundary effect, each OTC was surrounded by

seedlings of the same seed lot. The original heavy loam soil was excavated to the depth of 0.5 m and replaced by forest soil (approx. 0.12 % N on a dry mass basis). No nutrients or water were applied during the course of experiment.

Each decagonal OTC (diameter 3 m, height 4 m) was made of 1-m wide *Perspex* acrylic sheets, and had a usable ground area of 7.1 m<sup>2</sup>. The four OTC's were divided into ambient, AC (*ca.* 350 µmol mol<sup>-1</sup> or the current background level) and elevated, EC (AC + 400 µmol mol<sup>-1</sup>). The CO<sub>2</sub> enrichment started on 1 April, 1996, and the treatment was continuously applied since then on a 24 h basis, also in wintertime. For a more detailed description of the experimental conditions see Jach and Ceulemans (1999). Results of the second year (1997) of treatment are reported.

**Night-time  $R_D$  measurements:** Steady-state  $R_D$  measurements were made in the night using an open system infrared gas analyser (*CIRAS 1*, *PP Systems*, Herts., UK) and a standard cuvette of 2.5 cm<sup>2</sup> in diameter. Air temperature in the cuvette was measured using *BETATHERM* thermistors (type 200K6), and leaf temperature was calculated from the energy balance. Input air for the gas exchange system was supplied by passing building air through a *Drierite* column and a soda lime column attached to the *CIRAS* unit, to remove water vapour and CO<sub>2</sub>, respectively. Air of the desired CO<sub>2</sub> concentration was supplied by pressurized *Sparklets* CO<sub>2</sub> bulbs (*Hemisiphon*, Solingen, Germany) attached to an automatic CO<sub>2</sub> flow regulation system. A fan inside the cuvette stirred the air to maintain boundary layer resistance to H<sub>2</sub>O diffusion less than 0.2 m<sup>2</sup> s<sup>-1</sup> mol<sup>-1</sup>. The relative humidity of the air in the cuvette was set at 80 % and the flow rate at 3.33 cm<sup>3</sup> s<sup>-1</sup>.

All measurements were made in July 1997. Night-time measurements started 2 h after sunset, at 00:00 h, and ended at about 04:30 h. Measurements were made *in situ* in the OTC's on current-year and one-year-old shoots, all from the third whorl, situated on the south side of the tree. Two fully expanded and attached fascicles (four needles) per tree were enclosed in the cuvette for each  $R_D$  measurement. Needles were stretched completely across the cuvette. For each CO<sub>2</sub> treatment, measurements were made at actual night temperatures and at two CO<sub>2</sub> partial pressures (350 and 700 µmol mol<sup>-1</sup>), beginning with the growth concentration. The record was taken only after CO<sub>2</sub> efflux had reached a steady-state, usually requiring 10 to 15 min. Needle projected area was estimated prior to enclosure in the cuvette, from total needle width and cuvette width measured with a digital micrometer (*Mitutoyo*, Germany). After  $R_D$  measurements the fascicles were harvested, dried, and SLA, N and C concentrations were

determined as described below.

**Measurements of N, C, soluble sugar, and starch concentrations:** Immediately after  $R_D$  measurements, needles were destructively sampled for measurements of needle area, dry mass (DM), and foliar N and C concentrations. Needle area was calculated from average needle width and needle length. Each set of four needles was freeze-dried and SLA was determined as the ratio of projected needle area to dry mass [cm<sup>2</sup> kg<sup>-1</sup>], in order to calculate N and C concentrations on an area basis ( $N, C$ )<sub>area</sub> = [(N, C)<sub>mass</sub>/SLA]. Following drying, needles were ground in a mill (*Cyclotec 1093 Sample Mill*, Sweden) to fine powder and analysed for N and C by a Dynamic Flush Combustion Method with a *NC 2100 Soil Analyser* (*Carlo Erba*, Rodano, Italy).

Following  $R_D$  measurements, additional needles were collected for analysis of starch and soluble sugars. For this purpose, needles growing close to those used for  $R_D$  measurements and N and C analyses were sampled. Needles for saccharide analysis (a batch of approx. 30 needles) were harvested in the morning and put in a freeze-drier within 30 min. After drying, the needles were ground to fine powder and shipped to the University of Edinburgh, Scotland where they were analysed for sugars (inositol, sorbitol, galactase, glucose, fructose, sucrose) with a *Dionex DX-500 HPLC* (Sunnyvale, CA, USA) fitted with *GP40* gradient pump, *PA-1 CarboPac*, and *ED40* electrochemical detector. On a sub-sample of the needle powder, starch concentration was determined at the University of Antwerp enzymatically based on the formation of NADPH which is stoichiometric to the amount of D-glucose formed by the hydrolysis of starch (Boehringer 1997).

**Statistical methods:** A two-way ANOVA was used to investigate the long-term (indirect) effect of CO<sub>2</sub> treatment (*Treat.C*) and needle age (*Age*) on  $R_D$  expressed on various bases, and on chemical needle characteristics. A three-way ANOVA was used to investigate the short-term effects of measurement CO<sub>2</sub> concentration in the cuvette (*Cuv.C*), effects of CO<sub>2</sub> treatment (*Treat.C*) and needle age (*Age*) on  $R_D$  expressed on different bases. In each analysis also chamber (nested within treatment) and individual plant (nested within chamber) were added as random effects to the model. Adding individual as random effect to the model and the use of Satterthwaite's procedure to obtain the denominator degrees of freedom adjust the analysis for its repeated measures design. In this way a compound symmetry correlation structure was assumed. Analyses were performed in *SAS* (version 6.12) using the *MIXED* procedure (Littell *et al.* 1996).

## Results

**Direct effects of short-term CO<sub>2</sub> enrichment** on  $R_D$  were significant only in the AC for both needle age classes (Table 1). Short-term EC decreased  $R_D$  whether expressed per unit needle area, mass, N, C, or C/N. The EC caused changes in the direct response of  $R_D$  to short-term EC in both needle age classes and independent of the reference basis of expression. Area-based  $R_D$  of trees grown in AC compared with EC-trees was much more responsive to short-term changes in cuvette CO<sub>2</sub> as  $R_D$  decreased by 65 % in current-year and by 79 % in one-year-old needles when CO<sub>2</sub> was increased from 350 to 700  $\mu\text{mol mol}^{-1}$ , compared with reductions of 31 % in current-year and of 32 % in one-year-old needles in the EC treatment.

One-year-old needles had significantly lower  $R_D$  than current-year needles, irrespective of the treatment and cuvette CO<sub>2</sub> concentrations, when  $R_D$  was expressed per unit mass or per unit N (Table 1). When measured at AC, in both treatments  $R_D$  did not differ between the treatments (Table 1). However, when measured at EC, in both treatments  $R_D$  was higher in the EC-trees than AC-trees.

**Indirect effects of long-term CO<sub>2</sub> treatment:** A summary of the indirect effects of EC on  $R_D$  (measurements made at the growth CO<sub>2</sub> concentration) for two needle age classes, expressed per units of needle area, needle mass, N, C, and C/N, is presented in Table 2. Long-term EC treatment resulted in overall reductions in  $R_D$ . Differences were, however, only significant in one-year-old needles and only in mass-, C-, and C/N-based measurements (Table 2).

When expressed on a needle mass basis,  $R_D$  was 27 % lower in the EC treatment in current-year needles and 33 % lower in one-year-old needles as compared with the AC treatment. When expressed per unit C, the relative difference in  $R_D$  between the EC and AC treatments was higher in one-year-old needles (by 32 %) than in current-year needles (by 26 %). And when expressed on a unit C/N basis,  $R_D$  was 29 % lower in the EC treatment in current-year needles and 48 % lower in one-year-old needles as compared with the AC treatment.

Age effects were only significant in the EC-treatment, with higher  $R_D$  in current-year needles than in one-year-old needles when expressed on a needle mass or C and C/N bases.

**Chemical and morphological needle characteristics:** N concentration expressed on a mass basis ( $N_{\text{mass}}$ ) was significantly lower in needles from EC as compared with AC, but only in one-year-old needles (Fig. 1). The opposite was found when N was expressed per unit

needle area ( $N_{\text{area}}$ ), namely EC resulted in significantly higher  $N_{\text{area}}$  in the current-year needles, in contrast to one-year-old needles having lower, although not significantly,  $N_{\text{area}}$  concentrations (Fig. 1). Age effects were only significant for area-based N, with current-year needles having lower  $N_{\text{area}}$  concentrations as compared with one-year-old needles, independent of the CO<sub>2</sub> treatment (Fig. 1).

No significant differences in C between the treatments were found when C was expressed on a dry mass basis ( $C_{\text{mass}}$ ) (Fig. 1). Also needle age effects on  $C_{\text{mass}}$  were not significant (Fig. 1). When expressed per unit needle projected area, C concentration ( $C_{\text{area}}$ ) was significantly higher in EC treatment as compared with AC, but the difference was only significant in current-year needles. In both treatments, current-year needles had significantly less  $C_{\text{area}}$  than one-year-old needles (Fig. 1).

The response of both C and N concentrations expressed on a needle area basis resulted from the response of SLA to the EC treatment (Fig. 1). SLA was lower in the EC treatment as compared with AC in both needle age classes, but the difference was only significant in current-year needles. Age effects on SLA were significant in both treatments, one-year-old needles having lower SLA than current-year needles. Since no differences in  $C_{\text{mass}}$  were found in either needle age class, significantly higher  $C_{\text{area}}$  in current-year needles resulted from lower SLA. Similarly, in current-year needles higher  $N_{\text{area}}$  in EC resulted exclusively from lower SLA (Fig. 1). The significant reduction in  $N_{\text{mass}}$  in EC in one-year-old needles as compared with AC was partly counterbalanced by an increase in mesophyll tissue (SLA higher, although not significantly). This resulted in similar values of  $N_{\text{area}}$  of one-year-old needles for both treatments.

EC resulted in a significantly higher C/N ratio as compared with the AC treatment, but only in one-year-old needles (Fig. 1). The age effect was significant only in the AC treatment, where one-year-old needles had a lower C/N ratio as compared with current-year needles (Fig. 1).

The concentration of total soluble sugars was not affected by the CO<sub>2</sub> treatments (Table 3). When different soluble sugars were considered separately, the EC treatment resulted in significantly higher (more than doubled) glucose concentrations in both needle age classes as compared with AC. Also sorbitol concentrations were significantly enhanced by EC, but only in current-year needles (Table 3). One-year-old needles had generally higher concentrations of soluble sugars and this in both treatments, but the differences

Table 1. Direct (short-term) effects of CO<sub>2</sub> partial pressure (cuvette CO<sub>2</sub>) on dark respiration rate ( $R_D$ ) of current-year and one-year-old Scots pine needles grown for two years in either ambient, AC (360  $\mu\text{mol mol}^{-1}$ ) or elevated, EC (700  $\mu\text{mol mol}^{-1}$ ) atmospheric CO<sub>2</sub> concentrations (growth CO<sub>2</sub>). Results are expressed on a projected needle area [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ], needle dry mass [ $\text{mol kg}^{-1} \text{s}^{-1}$ ], needle nitrogen (N) [ $\text{mmol kg}^{-1} \text{s}^{-1}$ ], needle carbon (C) [ $\text{mol kg}^{-1} \text{s}^{-1}$ ], and needle C/N [ $\text{nmol s}^{-1}$ ] bases. Values shown are means  $\pm$  SE for each treatment where  $n = 8$ . Statistical significance of the single and interactive effects of treatment growth CO<sub>2</sub> concentration (*Treat.C*), needle age (*Age*), and measurement CO<sub>2</sub> concentration in the cuvette (*Cuv.C*) (three-factor ANOVA) is shown as: \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , ns =  $p > 0.05$ .  $R_D$  was measured at ambient temperature ( $T \approx 17.6^\circ\text{C}$ ) during the night.

| $R_D$ per   |    | Current-year needles |              | One-year-old needles |              | ANOVA statistics  |
|-------------|----|----------------------|--------------|----------------------|--------------|---|
|             |    | 350                  | 700          | 350                  | 700          |   |
| needle area | AC | 1.69 (0.19)          | 0.48 (0.13)  | 2.08 (0.15)          | 0.38 (0.07)  | <i>Treat.C</i> *  |
|             | EC | 2.09 (0.28)          | 1.56 (0.07)  | 2.30 (0.11)          | 1.55 (0.19)  | <i>Age</i> ns<br><i>Cuv.C</i> ***<br><i>Age</i> × <i>Treat.C</i> ns<br><i>Age</i> × <i>Cuv.C</i> ns<br><i>Treat.C</i> × <i>Cuv.C</i> ***<br><i>Treat.C</i> × <i>Age</i> × <i>Cuv.C</i> ns |
| dry mass    | AC | 11.74 (1.60)         | 3.20 (0.82)  | 9.79 (0.70)          | 1.78 (0.33)  | <i>Treat.C</i> *  |
|             | EC | 11.42 (1.40)         | 8.59 (0.40)  | 9.98 (1.04)          | 6.52 (0.79)  | <i>Age</i> *<br><i>Cuv.C</i> ***<br><i>Age</i> × <i>Treat.C</i> ns<br><i>Age</i> × <i>Cuv.C</i> ns<br><i>Treat.C</i> × <i>Cuv.C</i> ***<br><i>Treat.C</i> × <i>Age</i> × <i>Cuv.C</i> ns  |
| N           | AC | 1.50 (0.25)          | 0.41 (0.12)  | 1.00 (0.09)          | 0.18 (0.03)  | <i>Treat.C</i> *  |
|             | EC | 1.44 (0.19)          | 1.09 (0.07)  | 1.23 (0.13)          | 0.82 (0.12)  | <i>Age</i> **<br><i>Cuv.C</i> ***<br><i>Age</i> × <i>Treat.C</i> ns<br><i>Age</i> × <i>Cuv.C</i> ns<br><i>Treat.C</i> × <i>Cuv.C</i> **<br><i>Treat.C</i> × <i>Age</i> × <i>Cuv.C</i> ns  |
| C           | AC | 25.50 (3.44)         | 7.01 (1.79)  | 20.86 (1.52)         | 3.77 (0.69)  | <i>Treat.C</i> *  |
|             | EC | 25.07 (3.10)         | 18.84 (0.87) | 21.32 (2.10)         | 14.12 (1.73) | <i>Age</i> ns<br><i>Cuv.C</i> **<br><i>Age</i> × <i>Treat.C</i> ns<br><i>Age</i> × <i>Cuv.C</i> ns<br><i>Treat.C</i> × <i>Cuv.C</i> *<br><i>Treat.C</i> × <i>Age</i> × <i>Cuv.C</i> ns    |
| C/N         | AC | 0.21 (0.03)          | 0.06 (0.01)  | 0.21 (0.02)          | 0.04 (0.01)  | <i>Treat.C</i> *  |
|             | EC | 0.20 (0.03)          | 0.15 (0.01)  | 0.17 (0.02)          | 0.11 (0.01)  | <i>Age</i> ns<br><i>Cuv.C</i> **<br><i>Age</i> × <i>Treat.C</i> ns<br><i>Age</i> × <i>Cuv.C</i> ns<br><i>Treat.C</i> × <i>Cuv.C</i> ***<br><i>Treat.C</i> × <i>Age</i> × <i>Cuv.C</i> ns  |

were more pronounced in EC, except for inositol (Table 3). Thus, in the EC treatment, concentrations of total soluble sugars, inositol, glucose, and fructose were significantly higher in one-year-old than current-year needles. Starch concentrations were significantly higher

under EC as compared to AC, but only in current-year needles. Consequently, starch accumulation in current-year needles under EC resulted in significantly lower SLA as described above.

Table 2. Indirect (long-term) effects of the growth CO<sub>2</sub> concentration on the rate of dark respiration ( $R_D$ ) of current-year and one-year-old Scots pine needles grown for two years in either ambient, AC (360  $\mu\text{mol mol}^{-1}$ ) or elevated, EC (700  $\mu\text{mol mol}^{-1}$ ) atmospheric CO<sub>2</sub> concentrations and measured at the growth CO<sub>2</sub>. Results are expressed per unit of projected needle area [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ], needle dry mass [ $\mu\text{mol kg}^{-1} \text{s}^{-1}$ ], needle nitrogen (N) [ $\text{mmol kg}^{-1} \text{s}^{-1}$ ], needle carbon (C) [ $\mu\text{mol kg}^{-1} \text{s}^{-1}$ ], and needle C/N [ $\text{nmol s}^{-1}$ ]. Values shown are means  $\pm$ SE for each treatment where  $n = 8$ . Statistical significance of the single and interactive effects of treatment growth CO<sub>2</sub> concentration (*Treat.C*) and needle age (*Age*) (two-factor ANOVA) is shown as: \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , ns =  $p \geq 0.05$ .  $R_D$  was measured at ambient temperature ( $T \approx 17.6^\circ\text{C}$ ) during the night.

| $R_D$ per | Current-year needles         |              |       | One-year-old needles         |              |       | ANOVA statistics  |
|-----------|------------------------------|--------------|-------|------------------------------|--------------|-------|---|
|           | Growth CO <sub>2</sub><br>AC | EC           | EC/AC | Growth CO <sub>2</sub><br>AC | EC           | EC/AC |   |
| leaf area | 1.69 (0.19)                  | 1.56 (0.07)  | 0.92  | 2.08 (0.15)                  | 1.55 (0.19)  | 0.74  | <i>Treat.C</i> ns<br><i>Age</i> ns<br><i>Treat.C</i> $\times$ <i>Age</i> ns |
| dry mass  | 11.74 (1.60)                 | 8.59 (0.40)  | 0.73  | 9.79 (0.70)                  | 6.52 (0.79)  | 0.66  | <i>Treat.C</i> ***<br><i>Age</i> *<br><i>Treat.C</i> $\times$ <i>Age</i> ** |
| N         | 1.50 (0.25)                  | 1.09 (0.07)  | 0.72  | 1.00 (0.09)                  | 0.82 (0.12)  | 0.82  | <i>Treat.C</i> ns<br><i>Age</i> ns<br><i>Treat.C</i> $\times$ <i>Age</i> ns |
| C         | 25.50 (3.44)                 | 18.84 (0.87) | 0.74  | 20.86 (1.52)                 | 14.12 (1.73) | 0.67  | <i>Treat.C</i> *<br><i>Age</i> *<br><i>Treat.C</i> $\times$ <i>Age</i> **   |
| C/N       | 0.21 (0.03)                  | 0.15 (0.01)  | 0.71  | 0.21 (0.02)                  | 0.11 (0.01)  | 0.52  | <i>Treat.C</i> ***<br><i>Age</i> *<br><i>Treat.C</i> $\times$ <i>Age</i> ** |

## Discussion

In this study on Scots pine, we demonstrated evidence not only for a direct, short-term inhibition of needle  $R_D$  by EC, but also for some indirect, long-term effects of EC. Short-term exposure to different CO<sub>2</sub> concentrations was sufficient to lower or enhance  $R_D$ , and this response was fully reversible. So, leaf  $R_D$  of AC-trees decreased when measured in EC, while  $R_D$  of EC-trees was enhanced when measured in AC, similarly to the results of El Kohen *et al.* (1991) on sweet chestnut and Will and Ceulemans (1997) on poplar. The magnitude of this change was, however, affected by the growth CO<sub>2</sub> concentration. Short-term effect was only significant in the AC treatment, irrespective of needle age and of the way the results were expressed. A significant interaction between the direct and indirect effects is in agreement with the findings of Bunce (1990) and Thomas and Griffin (1994). In contrast, no differences between the AC and EC growth treatments in the relative reduction of  $R_D$  in response to an instantaneous increase in CO<sub>2</sub> concentration were found by Griffin *et al.* (1996).

Based on literature, leaf  $R_D$  is on average reduced by 15 to 20 % when CO<sub>2</sub> is increased from 350 to 700  $\mu\text{mol}$

$\text{mol}^{-1}$  (González-Meler and Siedow 1999). The metabolic explanation of the direct response remains hypothetical and the precise manner in which CO<sub>2</sub> affects  $R_D$  is still unknown. The inhibition appears, however, to be a classic case of feed-back control by an end-product (Amthor *et al.* 1992, González-Meler *et al.* 1996, González-Meler and Siedow 1999).

In response to the long-term EC, there was a considerable difference in  $R_D$  between the two needle age classes (for the effects of leaf age on photosynthetic characteristics see Šesták 1985). The indirect response of  $R_D$  to EC was larger in one-year-old needles, since they experienced a longer exposition to the treatment. Similar results were found in poplar clone Beaupré where the CO<sub>2</sub> effect was minimal in very young leaves and increased with leaf age (Will and Ceulemans 1997, Jarvis 1998). In one-year old needles  $R_D$  significantly decreased under EC as compared with AC, whether expressed on a mass, C, or C/N basis.

Similar to other studies (Poorter *et al.* 1992, Thomas *et al.* 1993, Azcón-Bieto *et al.* 1994, Thomas and Griffin 1994, Mitchell *et al.* 1995, Griffin *et al.* 1996) the

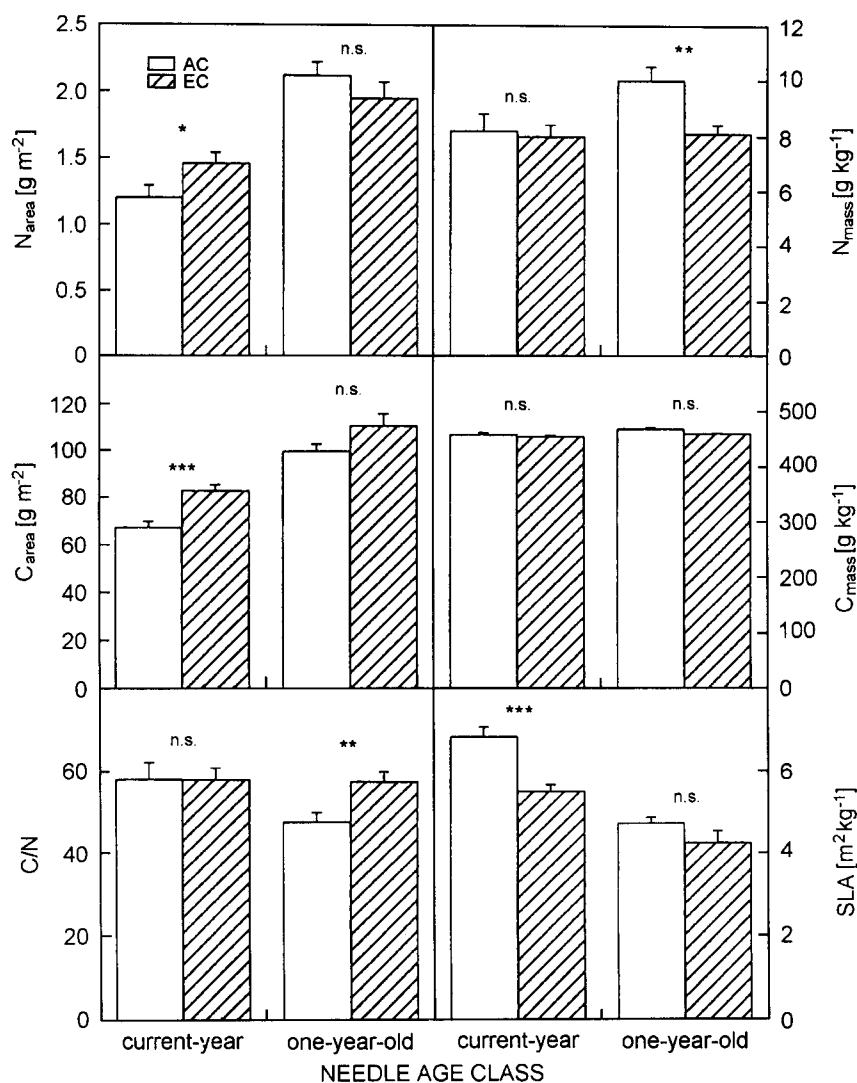


Fig. 1. Needle nitrogen (N) and carbon (C) concentrations, both on an area and a mass basis, carbon to nitrogen ratio (C/N), and specific leaf area (SLA) for current-year and one-year-old needles during the second growing season in ambient (AC) and elevated (EC) CO<sub>2</sub>. Mean values  $\pm$  SE are shown, with  $n = 8$ . Only treatment differences within the needle age class are shown; differences between the needle age classes within the treatments are described in the text. Levels of significance are indicated as: \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , ns  $p > 0.05$ .

present study re-emphasizes the importance of a multiple expression of values for the interpretation of the indirect effects of CO<sub>2</sub> on  $R_D$ . Changes in  $R_D$  in response to EC have often been described in terms of changes in sugar accumulation in the leaves and of a limited transport or sink activity. This could lead to excessive saccharide accumulation and consequently to a decrease in SLA, resulting in a decreased  $R_D$  on a mass basis, but increase area-based  $R_D$  (Thomas and Griffin 1994).

Nevertheless, no long-term CO<sub>2</sub> treatment effect was found when  $R_D$  was expressed on a needle area or N basis. Azcón-Bieto *et al.* (1983) suggested that

respiration is limited by a supply of glucose. Since glucose concentration on an area basis was significantly higher in both needle age classes in the EC treatment as compared with AC (Table 3) we could expect that this contributed to an absence of treatment differences in the area-based  $R_D$ . On an area basis, most studies on the long-term effects of EC on  $R_D$  report decreases, although also an increase or no change has been found (Jarvis 1998).  $R_D$  per unit projected needle area was similar to those reported in the literature for Scots pine needles measured at comparable ambient temperatures (Leverenz 1988).

Table 3. Soluble sugar and starch concentrations expressed on a projected needle area [ $\text{g m}^{-2}$ ] basis in current-year (CY) and one-year-old (OY) needles of Scots pine trees sampled in July of the second growing season under ambient (AC) and elevated (EC)  $\text{CO}_2$  conditions. Mean values of 22 plants per treatment  $\pm$  SE are shown.  $\Delta\%$  = relative difference between the treatments calculated as 100 [(EC – AC)/AC]. Statistical significance is shown as: \*\*\*  $p \leq 0.001$ , \*\*  $p \leq 0.01$ , \*  $p \leq 0.05$ , ns  $p > 0.05$ .

|              | Needle age  | AC             | EC             | Sign. diff. | $\Delta\%$ |
|--------------|-------------|----------------|----------------|-------------|------------|
| Inositol     | CY          | 1.562 (0.074)  | 1.779 (0.088)  | ns          | + 14       |
|              | OY          | 2.027 (0.087)  | 2.383 (0.141)  | ns          | + 18       |
|              | Sign. diff. | ***            | **             |             |            |
| Sorbitol     | CY          | 0.931 (0.386)  | 0.105 (0.047)  | *           | - 88       |
|              | OY          | 0.910 (0.415)  | 0.454 (0.212)  | ns          | - 50       |
|              | Sign. diff. | ns             | ns             |             |            |
| Glucose      | CY          | 1.900 (0.233)  | 3.874 (0.265)  | ***         | +104       |
|              | OY          | 2.067 (0.309)  | 4.684 (0.230)  | ***         | +127       |
|              | Sign. diff. | ns             | *              |             |            |
| Fructose     | CY          | 4.330 (0.456)  | 4.371 (0.382)  | ns          | + 1        |
|              | OY          | 4.724 (0.576)  | 5.832 (0.479)  | ns          | + 23       |
|              | Sign. diff. | ns             | *              |             |            |
| Sucrose      | CY          | 1.543 (0.571)  | 1.277 (0.194)  | ns          | - 17       |
|              | OY          | 3.427 (1.127)  | 1.099 (0.149)  | ns          | - 68       |
|              | Sign. diff. | ns             | ns             |             |            |
| Total sugars | CY          | 10.270 (1.000) | 11.410 (0.640) | ns          | + 11       |
|              | OY          | 13.160 (1.900) | 14.650 (0.850) | ns          | + 11       |
|              | Sign. diff. | ns             | **             |             |            |
| Starch       | CY          | 1.09 (0.19)    | 5.28 (0.82)    | *           | +384       |
|              | OY          | 2.20 (0.37)    | 2.84 (0.52)    | ns          | + 29       |
|              | Sign. diff. | ns             | ns             |             |            |

In one-year-old needles, the lower  $R_D$  per unit mass under EC resulted from a combination of both a lower (although not significantly) SLA and area-based  $R_D$ . In current-year needles, the significant decrease in SLA under EC as compared with AC did not counterbalance the absence of treatment differences in area-based  $R_D$ , with consequentially no differences in mass-based  $R_D$ . The significantly lower SLA in current-year needles in EC resulted from a significant starch accumulation in those needles. Indirect effects on needle  $R_D$  on a mass basis were also found in *Pinus palustris* (Mitchell *et al.* 1995) and *Pinus ponderosa* (Griffin *et al.* 1996).

Foliage N is a good predictor of  $R_D$ , because both growth respiration (Will and Ceulemans 1997) and maintenance respiration (Ryan 1991) are highly correlated with N concentration. A decreased  $N_{\text{mass}}$  as commonly observed for leaf tissues grown under EC (Curtis 1996, Cotrufo *et al.* 1998, Medlyn *et al.* 1999) may have contributed to the decreased  $R_D$  per mass basis (Amthor 1991, Wullschlegel *et al.* 1994) since a lower  $N_{\text{mass}}$  leads to lower construction costs and maintenance requirements (Mitchell *et al.* 1995). The absence of a treatment effect on  $R_D$  per unit N may reflect that there

was no difference in growth or saccharide transport at a given level of foliar N (Mitchell *et al.* 1995). Indeed, growth rates (Jach and Ceulemans 1999) and soluble saccharide concentrations were similar in both treatments at the time of  $R_D$  measurements. In contrast to our results,  $R_D$  per unit foliar N were higher in the EC treatment than in AC in *Pinus palustris* (Mitchell *et al.* 1995). However, in some studies a decreased  $R_D$  per unit leaf N, following a long-term EC treatment, has been found (Ziska and Bunce 1993, Ceulemans and Mousseau 1994).

There was less N per unit C in the one-year-old EC needles as compared with AC (Fig. 1). EC generally increases C/N ratio (Norby *et al.* 1992, Field *et al.* 1992) and thus may alter the relationship between  $R_D$  per unit dry mass and per unit N. An increased C/N ratio tends to reduce tissue specific respiration ( $R_D$  per unit of tissue mass) because more saccharides are stored in tissue grown under EC at little cost in maintenance respiration (Saxe *et al.* 1998). The C/N basically represents a balance between the supply of respiratory products (C) and the demand for respiratory products (N) and that is why the C/N ratio may also be a good indicator of



respiration (Griffin *et al.* 1996). In the present experiment,  $R_D$  per unit C/N was significantly decreased under EC, but only in one-year-old needles. In contrast to our findings, no indirect effects on  $R_D$  per unit C/N basis were found in *Pinus ponderosa* (Griffin *et al.* 1996).

The significantly lower  $R_D$  in one-year-old needles grown in EC as compared with current-year needles from the same treatment could possibly be attributed to a reduced SLA, but also to a lower demand for ATP and a reduced respiratory machinery (Sprugel *et al.* 1995). As leaves age, they become increasingly shaded and also more distant from active sinks (Sprugel *et al.* 1995). In the present experiment, shading effects became more pronounced in EC treatment than in AC (Jach and Ceulemans 1999). How  $R_D$  changes with leaf age in response to EC has not been, however, addressed in many studies. Young needles have higher  $R_D$  because they are still growing and construction respiration is occurring, but with maturation  $R_D$  decreases dramatically (Sprugel *et al.* 1995). Generally, conifers have lower  $R_D$  than broadleaf plants and  $R_D$  of conifers can vary considerably within the vertical canopy profile and with needle age (Sprugel *et al.* 1995). Because age related effects on  $R_D$  were only present in the EC treatment, the CO<sub>2</sub> treatment might have enhanced needle ontogeny.

There is some evidence in the literature that  $R_D$  varies with time due to leaf and plant ontogeny (Šesták 1985, Poorter *et al.* 1992, Mousseau 1993). In *Castanea sativa* leaf  $R_D$  in EC-trees were lower than in the AC ones in spring, but the effect disappeared with the prolongation of growing season (El Kohen *et al.* 1991). In the present study,  $R_D$  derived from the PAR response curves during the day was not affected by the EC treatment during the growing season (Jach and Ceulemans 2000), but a large seasonal variation in the response of  $R_D$  was found, with highest  $R_D$  during the

phase of rapid growth. The needle age effect was significant only in mid-summer and disappeared later. So, for a good understanding of the impact of EC on  $R_D$  it is essential to measure  $R_D$  throughout the growing season and for different leaf age classes.

In conclusion, both direct and indirect effects of EC on  $R_D$  were reported. The direct effects were only pronounced in the AC treatment, where a suppression of  $R_D$  was found with a short-term CO<sub>2</sub> increase. The indirect effects were only significant in one-year-old needles, where a decreased  $R_D$  was found in EC grown needles as compared with the AC ones, when  $R_D$  was expressed on a mass, C, or C/N bases. Long-term age-related effects on  $R_D$  were only significant in the EC treatment, where current-year needles had higher  $R_D$  than one-year-old needles on a mass, C, or C/N bases. Area- and N-based  $R_D$  seemed not to be very sensitive to long-term CO<sub>2</sub> exposure. Mass-, C-, and C/N-based  $R_D$  mirrored best both treatment- and age- related effects of long-term CO<sub>2</sub> exposure.

Since respiration may consume half of the CO<sub>2</sub> fixed in photosynthesis and since atmospheric CO<sub>2</sub> concentrations are rising in the range of the inhibitory effects, a better understanding of the long-term effects of increasing CO<sub>2</sub> on  $R_D$  is necessary to model plant and ecosystem responses to future climate. The highly variable and inconsistent response of  $R_D$  to CO<sub>2</sub> reported in the literature may be caused by the different ways of expressing the results, by leaf age, saccharide and nitrogen concentrations as well as by temperature at the time of measurement. These factors have seldom been considered in measurements of leaf  $R_D$  at either AC or EC. To resolve uncertainties about the response of leaf  $R_D$  to EC, both short-term direct effects and long-term acclimation effects should be considered in future experiments.

## References

- Amthor, J.S.: Respiration and Crop Productivity. - Springer-Verlag, New York - Berlin - Heidelberg - London - Paris - Tokyo 1989.
- Amthor, J.S.: Respiration in a future, higher-CO<sub>2</sub> world. - *Plant Cell Environ.* **14**: 13-20, 1991.
- Amthor, J.S.: Terrestrial higher-plant response to increasing atmospheric [CO<sub>2</sub>] in relation to the global carbon cycle. - *Global Change Biol.* **1**: 243-274, 1995.
- Amthor, J.S.: Plant respiratory responses to elevated carbon dioxide partial pressure. - In: Allen, L.H., Kirkham, M.B., Olszyk, D.M., Whitman, C. (ed.): *Advances in Carbon Dioxide Effects Research*. Pp. 35-77. Amer. Soc. Agron., Madison 1997.
- Amthor, J.S., Koch, G.W., Bloom, A.J.: CO<sub>2</sub> inhibits respiration in leaves of *Rumex crispus* L. - *Plant Physiol.* **98**: 757-760, 1992.
- Azcón-Bieto, J., Gonzalez-Meler, M.A., Doherty, W., Drake, B.G.: Acclimation of respiratory O<sub>2</sub> uptake in green tissues of field-grown native species after long-term exposure to elevated atmospheric CO<sub>2</sub>. - *Plant Physiol.* **106**: 1163-1168, 1994.
- Azcón-Bieto, J., Lambers, H., Day, D.A.: Effect of photosynthesis and carbohydrate status on respiratory rates and the involvement of the alternative pathway in leaf respiration. - *Plant Physiol.* **72**: 598-603, 1983.
- Baker, J.T., Laugel, F., Boote, K.J., Allen, L.H., Jr.: Effects of daytime carbon dioxide concentration on dark respiration in rice. - *Plant Cell Environ.* **15**: 231-239, 1992.

- Barnes, J.D., Pfirmann, T., Steiner, K., Lütz, C., Busch, U., Küchenhoff, H., Payer, H.-D.: Effects of elevated CO<sub>2</sub>, elevated O<sub>3</sub> and potassium deficiency on Norway spruce [*Picea abies* (L.) Karst.]: seasonal changes in photosynthesis and non-structural carbohydrate content. - *Plant Cell Environ.* **18**: 1345-1357, 1995.
- Bazzaz, F.A., Williams, W.E.: Atmospheric CO<sub>2</sub> concentrations within a mixed forest: implications for seedling growth. - *Ecology* **72**: 12-16, 1991.
- Boehringer Mannheim Biochemicals: Methods of enzymatic bioanalysis and food analysis, using test combinations. UV method for the determination of native starch in foodstuffs and other materials. - Pp. 128-131. Boehringer Mannheim Biochemicals, Mannheim 1997.
- Bunce, J.A.: Short- and long-term inhibition of respiratory carbon dioxide efflux by elevated carbon dioxide. - *Ann. Bot.* **65**: 637-642, 1990.
- Bunce, J.A.: Stomatal conductance, photosynthesis and respiration of temperate deciduous tree seedlings grown outdoors at an elevated concentration of carbon dioxide. - *Plant Cell Environ.* **15**: 541-549, 1992.
- Bunce, J.A.: Responses of respiration to increasing atmospheric carbon dioxide concentrations. - *Physiol. Plant.* **90**: 427-430, 1994.
- Bunce, J.A.: Effects of elevated CO<sub>2</sub> concentration in the dark on the growth of soybean seedlings. - *Ann. Bot.* **75**: 365, 1995.
- Bunce, J.A., Caulfield, F.: Reduced respiratory carbon dioxide efflux during growth at elevated carbon dioxide in three herbaceous perennial species. - *Ann. Bot.* **67**: 325-330, 1991.
- Ceulemans, R., Mousseau, M.: Effects of elevated atmospheric CO<sub>2</sub> on woody plants. - *New Phytol.* **127**: 425-446, 1994.
- Cotrufo, F.M., Ineson, P., Scott, A.: Elevated CO<sub>2</sub> reduces nitrogen concentration of plant tissues. - *Global Change Biol.* **4**: 43-54, 1998.
- Curtis, P.S.: A meta-analysis of leaf gas exchange and nitrogen in trees grown under elevated carbon dioxide. - *Plant Cell Environ.* **19**: 127-137, 1996.
- Drake, B.G., Azcón-Bieto, J., Berry, J., Bunce, J., Dijkstra, P., Farrar, J., Gifford, R.M., Gonzalez-Meler, M.A., Koch, G., Lambers, H., Siedow, J., Wulschleger, S.: Does elevated atmospheric CO<sub>2</sub> concentration inhibit mitochondrial respiration in green plants? - *Plant Cell Environ.* **22**: 649-657, 1999.
- El Kohen, A., Pontailier, J.-Y., Mousseau M.: Effet d'un doublement du CO<sub>2</sub> atmosphérique sur la respiration à l'obscurité des parties aériennes de jeunes châtaigniers (*Castanea sativa* Mill.). - *Compt. rend. Acad. Sci. Paris Sér. III* **312**: 477-481, 1991.
- Farrar, J.F.: The respiratory source of CO<sub>2</sub>. - *Plant Cell Environ.* **8**: 427-438, 1985.
- Field, C.B., Chapin, F.S., Matson, P.A., Mooney, H.A.: Responses of terrestrial ecosystems to the changing atmosphere: a resource-based approach. - *Annu. Rev. Ecol. Syst.* **23**: 201-235, 1992.
- Gifford, R.M., Lambers, H., Morison, J.I.L.: Respiration of crop species under CO<sub>2</sub> enrichment. - *Physiol. Plant.* **63**: 351-356, 1985.
- González-Meler, M.A., Ribas-Carbó, M., Siedow, J.N., Drake, B.G.: Direct inhibition of plant mitochondrial respiration by elevated CO<sub>2</sub>. - *Plant Physiol.* **112**: 1349-1355, 1996.
- González-Meler, M.A., Siedow, J.N.: Direct inhibition of mitochondrial respiratory enzymes by elevated CO<sub>2</sub>: does it matter at the tissue or whole-plant level? - *Tree Physiol.* **19**: 253-259, 1999.
- Griffin, K.L., Ball, J.T., Strain, B.R.: Direct and indirect effects of elevated CO<sub>2</sub> on whole-shoot respiration in ponderosa pine seedlings. - *Tree Physiol.* **16**: 33-41, 1996.
- Gunderson, C.A., Wulschleger, S.D.: Photosynthetic acclimation in trees to rising atmospheric CO<sub>2</sub>: a broader perspective. - *Photosynth. Res.* **39**: 369-388, 1994.
- Hagihara, A., Hozumi, K.: Respiration. - In: Ragavendra, A.S. (ed.): *Physiology of Trees*. Pp. 87-100. John Wiley and Sons, New York 1991.
- Hrubec, T.C., Robinson, J.M., Donaldson, R.P.: Effects of CO<sub>2</sub> enrichment and carbohydrate content on the dark respiration of soybeans. - *Plant Physiol.* **79**: 684-689, 1985.
- Jach, M.E., Ceulemans, R.: Effects of elevated atmospheric CO<sub>2</sub> on phenology, growth and crown structure of Scots pine (*Pinus sylvestris* L.) seedlings after two years of exposure in the field. - *Tree Physiol.* **19**: 289-300, 1999.
- Jach, M.E., Ceulemans, R.: Effects of season, needle age and elevated atmospheric CO<sub>2</sub> on photosynthesis in Scots pine (*Pinus sylvestris* L.). - *Tree Physiol.* **20**: 145-157, 2000.
- Jarvis, P.G.: *European Forests and Global Change: The Likely Impacts of Rising CO<sub>2</sub> and Temperature*. - Cambridge University Press, Cambridge 1998.
- Kubiske, M.E., Pregitzer, K.S.: Effects of elevated CO<sub>2</sub> and light availability on the photosynthetic light response of trees of contrasting shade tolerance. - *Tree Physiol.* **16**: 351-358, 1996.
- Leverenz, J.W.: The effects of illumination sequence, CO<sub>2</sub> concentration, temperature and acclimation on the convexity of the photosynthetic light response curve. - *Physiol. Plant.* **74**: 332-341, 1988.
- Littell, R.C., Milliken, G.A., Stroup, W.W., Wolfinger, R.D.: *SAS System for Mixed Models*. - SAS Institute, Cary 1996.
- Marek, M.V., Šprtová, M., Kalina, J.: The photosynthetic irradiance-response of Norway spruce exposed to a long-term elevation of CO<sub>2</sub> concentration. - *Photosynthetica* **33**: 259-268, 1997.
- Medlyn, B.E., Badeck, F.-W., de Pury, D.G.G., Barton, C.V.M., Broadmeadow, M., Ceulemans, R., de Angelis, P., Forstreuter, M., Jach, M.E., Kellomäki, S., Laitat, E., Marek, M., Philippot, S., Rey, A., Strassmeyer, J., Laitinen, K., Liozon, R., Portier, B., Roberntz, P., Wang, K., Jarvis, P.G.: Effects of elevated CO<sub>2</sub> on photosynthesis in European forest species: a meta-analysis of model parameters. - *Plant Cell Environ.* **22**: 1475-1495, 1999.
- Mitchell, R.J., Runion, G.B., Prior, S.A., Rogers, H.H., Amthor, J.S., Henning, F.P.: Effects of nitrogen on *Pinus palustris* foliar respiratory responses to elevated atmospheric CO<sub>2</sub> concentrations. - *J. exp. Bot.* **46**: 1561-1567, 1995.
- Mousseau, M.: Effects of elevated CO<sub>2</sub> on growth, photosynthesis and respiration of sweet chestnut (*Castanea sativa* Mill.). - *Vegetatio* **104/105**: 413-419, 1993.

- Mousseau, M., Saugier, B.: The direct effect of increased CO<sub>2</sub> on gas exchange and growth of forest tree species. - J. exp. Bot. **43**: 1121-1130, 1992.
- Nijs, I., Impens, I., Behaeghe, T.: Effects of elevated atmospheric carbon dioxide on gas exchange and growth of white clover. - Photosynth. Res. **15**: 163-175, 1988.
- Norby, R.J., Gunderson, C.A., Wullschleger, S.D., O'Neill, E.G., McCracken, M.K.: Productivity and compensatory responses of yellow-poplar trees in elevated CO<sub>2</sub>. - Nature **357**: 322-324, 1992.
- Norby, R.J., Wullschleger, S.D., Gunderson, C.A., Johnson, D.W., Ceulemans, R.: Tree responses to rising CO<sub>2</sub> in field experiments: implications for the future forest. - Plant Cell Environ. **22**: 683-714, 1999.
- Poorter, H., Gifford, R.M., Kriedemann, P.E., Wong, S.C.: A quantitative analysis of dark respiration and carbon content as factors in the growth response of plants to elevated CO<sub>2</sub>. - Aust. J. Bot. **40**: 501-513, 1992.
- Poorter, H., Van Berkel, Y., Baxter, R., Den Hertog, J., Dijkstra, P., Gifford, R.M., Griffin, K.L., Roumet, C., Roy, J., Wong, S.C.: The effects of elevated CO<sub>2</sub> on the chemical composition and construction costs of leaves of 27 C<sub>3</sub> species. - Plant Cell Environ. **20**: 472-482, 1997.
- Ryan, M.G.: Effects of climate change on plant respiration. - Ecol. Appl. **1**: 157-167, 1991.
- Saralabai, V.C., Vivekanandan, M., Suresh Babu, R.: Plant responses to high CO<sub>2</sub> concentration in the atmosphere. - Photosynthetica **33**: 7-37, 1997.
- Saxe, H., Ellsworth, D.S., Heath, J.: Tree and forest functioning in an enriched CO<sub>2</sub> atmosphere. - New Phytol. **139**: 395-436, 1998.
- Šesták, Z. (ed.): Photosynthesis During Leaf Development. - Academia, Praha; Dr W. Junk Publ., Dordrecht - Boston - Lancaster 1985.
- Sprugel, D.G., Benecke, U.: Measuring woody-tissue respiration and photosynthesis. - In: Lassoie, J.P., Hinckley, T.M. (ed.): Techniques and Approaches in Forest Tree Ecophysiology. Pp. 329-355. CRC Press, Boca Raton 1991.
- Sprugel, D.G., Ryan, M.G., Brooks, J.R., Vogt, K.A., Martin, T.A.: Respiration from the organ level to the stand. - In: Smith, W.K., Hinckley, T.M. (ed.): Resource Physiology of Conifers. Acquisition, Allocation, and Utilisation. Pp. 255-298. Academic Press, San Diego 1995.
- Teskey, R.O.: A field study of the effects of elevated CO<sub>2</sub> on carbon assimilation, stomatal conductance and leaf and branch growth of *Pinus taeda* trees. - Plant Cell Environ. **18**: 565-573, 1995.
- Thomas, R.B., Griffin, K.L.: Direct and indirect effects of atmospheric carbon dioxide enrichment on leaf respiration of *Glycine max* (L.) Merr. - Plant Physiol. **104**: 355-361, 1994.
- Thomas, R.B., Reid, C.D., Ybema, R., Strain, B.R.: Growth and maintenance components of leaf respiration of cotton grown in elevated carbon dioxide partial pressure. - Plant Cell Environ. **16**: 539-546, 1993.
- Urban, O., Marek, M.V.: Seasonal changes of selected parameters of CO<sub>2</sub> fixation biochemistry of Norway spruce under the long-term impact of elevated CO<sub>2</sub>. - Photosynthetica **36**: 533-545, 1999.
- Wang, K.-Y., Kellomäki, S., Laitinen, K.: Effects of needle age, long-term temperature and CO<sub>2</sub> treatments on the photosynthesis of Scots pine. - Tree Physiol. **15**: 211-218, 1995.
- Will, R.E., Ceulemans, R.: Effects of elevated CO<sub>2</sub> concentration on photosynthesis, respiration and carbohydrate status of coppice *Populus* hybrids. - Physiol. Plant. **100**: 933-939, 1997.
- Williams, K., Jones, D.G., Baxter, R., Farrar, J.F.: The effect of enhanced concentrations of atmospheric CO<sub>2</sub> on leaf respiration. - In: Lambers, H., van der Plas, L.H.W. (ed.): Molecular, Biochemical and Physiological Aspects of Plant Respiration. Pp. 547-551. SPB Academic Publ., The Hague 1992.
- Wullschleger, S.D., Ziska, L.H., Bunce, J.A.: Respiratory responses of higher plants to atmospheric CO<sub>2</sub> enrichment. - Physiol. Plant. **90**: 221-229, 1994.
- Wullschleger, S.D., Norby, R.J.: Respiratory cost of leaf growth and maintenance in white oak saplings exposed to atmospheric CO<sub>2</sub> enrichment. - Can. J. Forest Res. **22**: 1717-1721, 1992.
- Wullschleger, S.D., Norby, R.J., Gunderson, C.A.: Growth and maintenance respiration in leaves of *Liriodendron tulipifera* L. exposed to long-term carbon dioxide enrichment in the field. - New Phytol. **121**: 515-523, 1992.
- Wullschleger, S.D., Norby, R.J., Gunderson, C.A.: Forest trees and their response to atmospheric carbon dioxide: a compilation of results. - In: Allen, L.H., Kirkham, M.B., Olszyk, D.M., Whitman, C. (ed.): Advances in Carbon Dioxide Effects Research. Amer. Soc. Agron., Madison 1997.
- Ziska, L.H., Bunce, J.A.: Inhibition of whole plant respiration by elevated CO<sub>2</sub> as modified by growth temperature. - Physiol. Plant. **87**: 459-466, 1993.
- Ziska, L.H., Bunce, J.A.: Direct and indirect inhibition of single leaf respiration by elevated CO<sub>2</sub> concentrations: interaction with temperature. - Physiol. Plant. **90**: 130-138, 1994.
- Ziska, L.H., Seemann, J.R., DeJong, T.M.: Salinity-induced limitations in photosynthesis in *Prunus salicina*, a deciduous tree species. - Plant Physiol. **93**: 864-870, 1990.