

# Interrelationships between nitrogen supply and photosynthetic parameters in *Vicia faba* L.

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## Abstract

We determined for *Vicia faba* L the influence of nitrogen uptake and accumulation on the values of photon saturated net photosynthetic rate ( $P_{Nmax}$ ), quantum yield efficiency ( $\alpha$ ), intercellular CO<sub>2</sub> concentration ( $C_i$ ), and carboxylation efficiency ( $C_e$ ). As leaf nitrogen content ( $N_L$ ) increased, the  $\alpha$  converged onto a maximum asymptotic value of  $0.0664 \pm 0.0049 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$ . Also, as  $N_L$  increased the  $C_i$  value fell to an asymptotic minimum of  $115.80 \pm 1.59 \mu\text{mol mol}^{-1}$ , and  $C_e$  converged onto a maximum asymptotic value of  $1.645 \pm 0.054 \mu\text{mol}(\text{CO}_2) \text{m}^{-2} \text{s}^{-1} \text{Pa}^{-1}$  and declined to zero at a  $N_L$ -intercept equal to  $0.596 \pm 0.096 \text{g(N) m}^{-2}$ .  $\alpha$  fell to zero for an  $N_L$ -intercept of  $0.660 \pm 0.052 \text{g(N) m}^{-2}$ . As  $N_L$  increased, the value of  $P_{Nmax}$  converged onto a maximum asymptotic value of  $33.400 \pm 2.563 \mu\text{mol}(\text{CO}_2) \text{m}^{-2} \text{s}^{-1}$ .  $P_N$  fell to zero for an  $N_L$ -intercept of  $0.710 \pm 0.035 \text{g(N) m}^{-2}$ . Under variable daily meteorological conditions the values for  $N_L$ , specific leaf area ( $\delta_L$ ), root mass fraction ( $R_f$ ),  $P_{Nmax}$ , and  $\alpha$  remained constant for a given N supply. A monotonic decline in the steady-state value of  $R_f$  occurred with increasing N supply.  $\delta_L$  increased with increasing N supply or with increasing  $N_L$ .

**Additional key words:** carboxylation efficiency; faba bean; intercellular CO<sub>2</sub> concentration; net photosynthetic rate; quantum yield efficiency; root mass fraction.

## Introduction

In general, the overall relationship between the photosynthetic rate and leaf nitrogen content ( $N_L$ ) follows a curvilinear pattern, with the linear portion of the curve originating from a positive  $N_L$ -intercept (Schmitt and Edwards 1981, Sage and Percy 1987, Sinclair and Horie 1989, Meinzer and Zhu 1998). As  $N_L$  increases, the photosynthetic rate converges onto an asymptotic maximum value. This curvilinear relationship arises as a direct consequence of various rate limiting processes which play a role in setting the upper limit for the maximum possible values obtainable for photosynthetic parameters such as the photon-saturated net rate of photosynthesis ( $P_{Nmax}$ ) and the quantum yield efficiency ( $\alpha$ ) as  $N_L$  increases. For example, the maximum value for  $P_{Nmax}$  is directly proportional to the flux of CO<sub>2</sub> from the atmosphere. The rate of CO<sub>2</sub> diffusion into the leaf is directly proportional to the atmosphere-leaf CO<sub>2</sub> concentration gradient, the stomatal conductance, the mesophyll conductance, and the carboxylation efficiency of the leaf (Charles-Edwards

1978, Fisher *et al.* 1981, Evans and Caemmerer 1996, Katul *et al.* 2000). The carboxylation efficiency ( $C_e$ ) is dependent on the concentration of the various enzyme catalytic sites available for the different reactions involved in photosynthetic metabolism. Theoretically the upper limit to the maximum value for  $P_{Nmax}$  which also determines the plant potential photosynthetic capacity, is proportional to the number of active catalytic sites in chloroplasts that are involved in the reductive assimilation of CO<sub>2</sub>. The total number of these catalytic sites involved in CO<sub>2</sub> assimilation is dependent on the proportion of the total  $N_L$  allocated to the chloroplasts.

The asymptotic relationship between  $P_{Nmax}$  or  $\alpha$  and  $N_L$  indicates that as the number of catalytic units increases, other processes become in turn rate limiting. As  $N_L$  increases, these processes will in turn fix the upper limit of the plant N efficiency for biomass production. In this respect, the rate of carbon dioxide diffusion into the leaf will be a factor limiting the plant N dependent

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**Abbreviations:**  $C_a$  – ambient CO<sub>2</sub> concentration;  $C_e$  – carboxylation efficiency;  $C_i$  – intercellular CO<sub>2</sub> concentration;  $N_e$  – tissue nitrogen concentration;  $N_L$  – leaf nitrogen content;  $N_s$  – nitrogen supply concentration;  $N_t$  – total elemental nitrogen accumulated by the plant;  $P_{Nmax}$  – photon-saturated rate of photosynthesis;  $R_f$  – root mass fraction;  $r_m$  – mesophyll resistance;  $\alpha$  – quantum yield efficiency;  $\delta_L$  – specific leaf area;  $\mu_w$  – specific growth rate.

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capacity for biomass production (Evans and Caemmerer 1996, Katul *et al.* 2000). Other transport processes such as the rate of phosphate recycling between the chloroplast and cytosol also play a prominent role in fixing the upper limit of the plant capacity for biomass production (Cockburn *et al.* 1967a,b, Usuda and Edwards 1982, Pradet and Raymond 1983, Mächler *et al.* 1984, Rao and Terry 1989, 1995, Rao *et al.* 1989a,b).

If the concentration of nitrogen supply ( $N_s$ ) to the roots remains constant while the plant exponentially grows, then steady-state contents of N within the plant can only be maintained if the capacity for N accumulation increases exponentially during plant growth (Hirsose 1986, Garnier *et al.* 1989). In this context, the term balance exponential growth has been used to describe the situation where under constant conditions of nutrient supply, moisture availability, and saturating irradiance, all extensive variables (*e.g.* plant biomass and leaf area) increase exponentially at a constant specific growth rate ( $\mu_w$ ). Under these conditions all variables that are either ratios or rates (*e.g.* plant N concentration)

## Materials and methods

**Plants:** We used the cold-hardy *Vicia faba* L. cv. Aquadulce Claudia. Early spring or autumn planted cold-hardy cultivars require approximately 130–150 d for crop development, while over-wintering cultivars require approximately 240 d to reach dry seed maturity. Faba beans germinate and grow well in cool soil. For out-door experimental pot trials seeds were sown on the 23<sup>rd</sup> February and all measurements were done in April. During April, the daily average extraterrestrial global irradiance was 28.7 MJ m<sup>-2</sup>, average photoperiod 11.99 h, average daily maximum temperature 21.6 °C, average daily minimum temperature 9.1 °C, and average A-pan evaporation 6.3 mm d<sup>-1</sup>.

Faba bean seeds were planted in 1 600 cm<sup>3</sup> pots (150 mm diameter) filled with autoclaved river sand (medium grained with sieve mesh 2.5 mm). Two seeds were planted in each pot and after germination one seedling per pot was selected so as to give an initial uniform population of seedlings. All pots were watered every second day with tap water until emergence of cotyledons. Once the seedlings had emerged, they were watered every second day with a modified Long Ashton nutrient solution. The modifications involved the application of the nitrogen in the form KNO<sub>3</sub> at the following concentrations of N: 0, 0.714, 1.786, 3.571, 7.143, 17.857, and 35.714 mM N [equivalent to 0, 10, 25, 50, 100, 250, and 500 g(N) m<sup>-3</sup>]. All zero nitrogen treatments were watered using N-free Long Ashton nutrient solution. Dry biomass, nitrogen, and photosynthesis were measured for three harvest intervals: 44 (T<sub>1</sub>, 8<sup>th</sup> April), 51 (T<sub>2</sub>, 15<sup>th</sup> April), and 58 (T<sub>3</sub>, 22<sup>nd</sup> April) d after planting (DAP).

**Biomass determination:** At each harvesting interval, the roots were rinsed carefully with tap water to remove sand. The plants were divided into leaf, stem, and root

remain constant (Thornley 1998). Under balanced exponential growth, it is expected that  $\mu_w$  or specific photosynthetic rate would be determined by the plant steady-state  $N_c$ . This was an important consideration in this study. Under balanced exponential growth, it would also be expected that ratios such as the root mass fraction ( $R_f$ ) and the specific leaf area ( $\delta_L$ ) would remain constant. Information on these predictions does not exist for *Vicia faba*. For *V. faba* there are also no reports on how  $N_L$  influences  $P_{Nmax}$  and  $\alpha$ . Therefore the objectives of this study were: (1) determination of the asymptotic values and associated intercept values for  $P_{Nmax}$  and  $\alpha$  with respect to  $N_L$ ; (2) provision of information on the minimum value of  $N_L$  below which  $P_{Nmax}$  and  $\alpha$  equal zero; (3) establishing whether intensive variables or ratios such as  $P_{Nmax}$ ,  $\alpha$ ,  $N_c$ ,  $N_L$ ,  $\delta_L$ , and  $R_f$  remain constant under steady-state  $N_s$ . The latter information will help decide whether these intensive variables or ratios can be treated as unchanging plant growth parameters whose values are governed by the steady-state  $N_s$ .

components, respectively. Before drying, leaf area was measured using a Li-Cor 3100 area meter (Li-Cor, Lincoln, NE, USA). The leaves, stems, and roots were dried in the oven at 105 °C for 15 min and then at 65 °C for 3 d for dry mass determinations.

**Tissue N content analysis:** After the determinations of dry mass, the dried leaves, stems, and roots were milled for N analysis; 0.100±0.001 g milled plant material were digested by the Kjeldahl procedure. The total N content of the digests was determined using colorimetric assays (Dorich and Nelson 1983, Anderson and Ingram 1993). For the Kjeldahl digestion mixture, 0.42 g of selenium powder and 14 g lithium sulphate was added to 350 cm<sup>3</sup> of 30 % hydrogen peroxide, and 450 cm<sup>3</sup> of concentrated sulphuric acid was added slowly to the mixture that was kept cool in an ice bath.

For the colorimetric determination of ammonium nitrogen, two reagent solutions were used. The first reagent solution was made up of 34 g sodium carbonate, 25 g sodium citrate, and 25 g sodium tartarate dissolved together in 750 cm<sup>3</sup> H<sub>2</sub>O. To this mixture 0.12 g sodium nitroprusside was added and when dissolved the solution was made up to 1 000 cm<sup>3</sup> with H<sub>2</sub>O. The second reagent solution was made of 30 g sodium hydroxide dissolved in 750 cm<sup>3</sup> H<sub>2</sub>O. After cooling, 10 cm<sup>3</sup> of sodium hypochlorite was added and the solution was brought up to 1 000 cm<sup>3</sup>. For the ammonium assay, a 0.1 cm<sup>3</sup> ammonium sample was added to a 25 cm<sup>3</sup> Erlenmeyer flask and 5.0 cm<sup>3</sup> of both the first and second reagent solutions were added. After 1 h the absorbance was measured at 665 nm.

**Gas exchange measurements:** Photosynthetic gas exchange rates were determined at the three harvest

intervals on equal-aged cohorts. All plants were fully acclimated to full sunlight. For the measurements of  $C_i$  and  $C_e$ , the portable *CIRAS-1*, *PP IR* gas analysis system (*PP system*, Hitchin, Hertfordshire, U.K.) was used. For the estimation of  $P_{Nmax}$  and  $\alpha$ , an *ADC* infra-red gas analyser (*IRGA*) *225-2B-SS* (*Analytical Development*, Hoddesdon, Hertfordshire, England) on differential mode was used. These values were measured at 44, 51, and 58 DAP. Leaf surfaces were exposed to photosynthetically active photon flux densities (PPFD) ranging from 75 to 1 800  $\mu\text{mol}(\text{quantum})\text{ m}^{-2}\text{ s}^{-1}$  by adjusting the height of a 400 W halide (*Power Star HQ1*) lamp above the leaf surface. The PPFD at the leaf surface was measured with a *LI-188B* quantum meter.  $P_N$  was measured on the youngest fully expanded leaves. The trifoliate leaflet was clamped into and sealed in a *Perspex* leaf chamber that allowed the irradiation of 0.63  $\text{cm}^2$  of leaf area. The chamber was surrounded by a water jacket connected to a temperature regulated water bath water (*SS-CD-5*, *Specht Scientific*, Johannesburg, SA). Leaf temperature was continuously monitored by means of a thermocouple

(Model *BAT-12*, *Bailey Instruments*, Saddlebrook, NJ, USA) touching the underside of the lamina within the leaf chamber. The leaf temperature was maintained at 25 °C for all PPFDs. Average barometric pressure during photosynthetic measurements was 83.5 kPa.

**Experimental design and statistical analysis:** The experiment involved a completely randomised 3×7 factorial design. The experimental factors were three different harvest intervals ( $T_1$ ,  $T_2$ , and  $T_3$ ) and seven N treatments. A replicate consisted of one faba bean plant per container. The appropriate data were analysed using *ANOVA*. A Tukey's multiple comparison test was performed to determine which treatments differed. When appropriate, means±SE (standard error) were calculated, and when the F ratio was significant, the least significant differences were evaluated by the Tukey HSD-test to examine differences among the harvesting intervals ( $T_1$ ,  $T_2$ , and  $T_3$ ) in each treatment. *SYSTAT* version 8.0 was used for all data analyses.

## Results

**Effects of N supply on biomass production and N accumulation:** For each nitrogen supply concentration ( $N_s$ ) the nitrogen contents ( $N_c$ ) in leaves, stems, and roots remained constant over the three harvest intervals (Fig. 1). While the total quantity of elemental N accumulated per plant ( $N_t$ ) increased with increasing  $N_s$  (Fig. 3),  $N_c$  for the different tissue components always remained constant over time for each  $N_s$  (Fig. 1). Hence the plant capacity to accumulate N increased exponentially or kept pace with the rate of biomass production. Also under constant  $N_s$ , the specific leaf nitrogen ( $N_L$ ), specific leaf area ( $\delta_L$ ), root mass fraction ( $R_f$ ), and  $P_{Nmax}$  all remained constant over the three harvest intervals (Fig. 2). All these results are consistent with the balanced exponential growth hypothesis. Increasing  $N_s$  promoted increases in the production of total biomass, total plant leaf area, and total accumulation of nitrogen (Fig. 3).

**Effects of  $N_s$  on  $R_f$ ,  $\delta_L$ , and  $N_L$ :** Both  $R_f$  and  $\delta_L$  declined slightly with increasing  $N_s$  (Fig. 2). However, these slightly declining differences in  $R_f$  and  $\delta_L$  in response to increasing  $N_s$  were significantly different at the 5 % level. There were no significant differences at the 5 % level among three harvest intervals within the specific N treatments. Even though both total biomass and leaf area production increased significantly with increasing  $N_s$ , the values for  $R_f$  and  $\delta_L$  remained constant over the three harvest intervals.  $\delta_L$  also declined slightly with increasing  $N_L$ .  $N_L$  expressed on a leaf area basis also remained constant over time for a given steady state  $N_s$  (no significant differences at the 5 % level among three harvest intervals within the  $N_L$  treatments). These results are consistent with the balanced exponential growth hypothesis that predicts that if  $N_s$  is constant then intensive variables

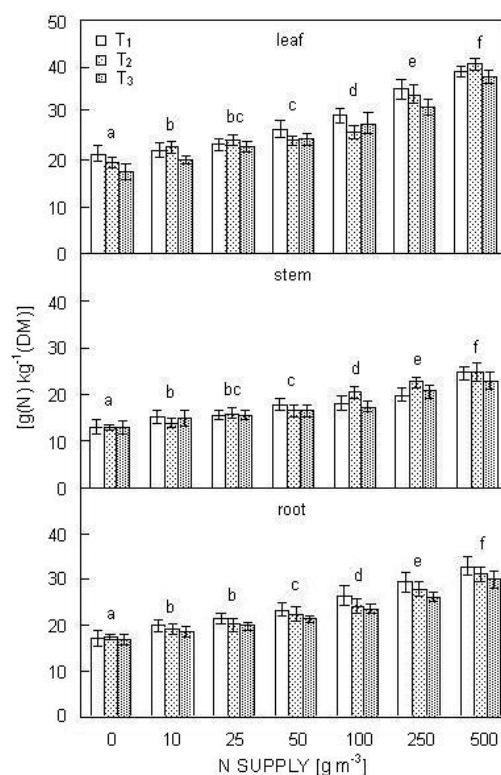


Fig. 1. Steady-state leaf, stem, and root N contents ( $N_c$ ) at the three harvests ( $T_1$ ,  $T_2$ , and  $T_3$ ) in response to different N supply (0, 10, 25, 50, 100, 250, and 500  $\text{g m}^{-3}$ ). *ANOVA* results for tissue  $N_c$  did not indicate significant differences (at the 5 % level) among the harvest intervals within the individual N treatments, but the differences among the N treatments were significant. Different letters indicate significant difference among treatments assessed by Tukey HSD-test ( $p < 0.05$ ).

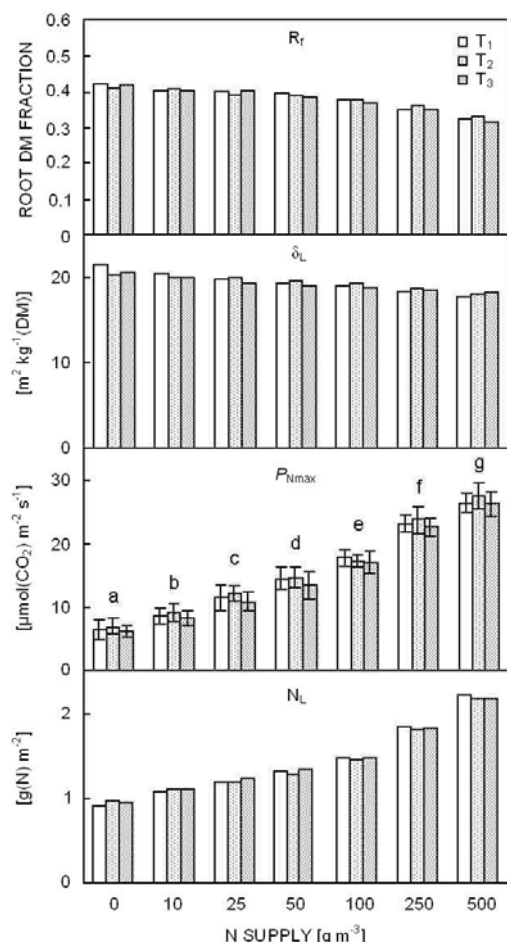


Fig. 2. The influence of increasing N supply on root dry mass fraction ( $R_f$ ), specific leaf area ( $\delta_L$ ), photon-saturated net photosynthetic rate ( $P_{Nmax}$ ), and specific leaf N content ( $N_L$ ) at the three harvest intervals ( $T_1$ ,  $T_2$ , and  $T_3$ ). ANOVA results did not indicate significant differences (at the 5 % level) among the harvest intervals for individual N treatments, but the differences among the N treatments were significant. For  $P_{Nmax}$  results, different letters indicate significant difference among treatments assessed by Tukey HSD-test ( $p < 0.05$ ).

such as  $R_f$ ,  $\delta_L$ , and  $N_L$  will also remain constant during plant growth and the capacity to accumulate N also increases exponentially. Therefore in modelling N uptake during plant growth  $R_f$ ,  $\delta_L$ , and  $N_L$  should be used as parameters.

**$\alpha$  versus  $N_s$ :** How does leaf  $N_c$  affect  $\alpha$ , given the fact that the total leaf area per plant and the total number of catalytic units per unit leaf area increases markedly with increasing  $N_s$  (Fig. 3)? For *V. faba*,  $\alpha$  showed a curvilinear response to increasing  $N_L$  (Fig. 4). The empirical equation of Sinclair and Horie (1989) was used for generating the asymptotic value for  $\alpha$  and the  $N_L$ -intercept value for  $\alpha = 0$ . With increasing  $N_L$ ,  $\alpha$  converged onto a maximum asymptotic value of  $0.0664 \pm 0.0049 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$ .  $\alpha$  fell to zero for  $N_L$ -intercept of below  $0.6600 \pm 0.0515 \text{ g(N)} \text{m}^{-2}$ . For a given  $N_s$ ,

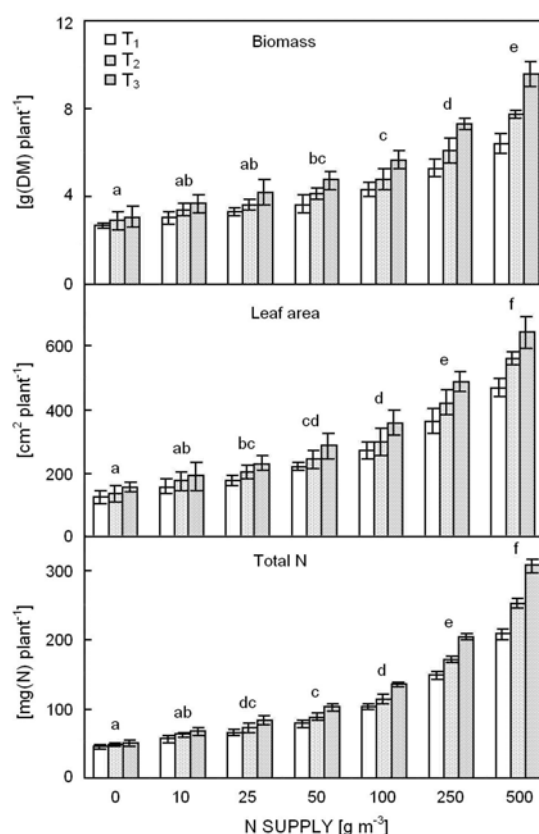


Fig. 3. The influence of increasing N supply on total biomass production, leaf area production, and total plant N accumulation ( $N_L$ ) measured at the three harvests ( $T_1$ ,  $T_2$ , and  $T_3$ ). ANOVA for all the results indicated (1) significant differences (at the 5 % level) among the N treatments, and (2) significant differences among the harvest intervals within the N treatments. Different letters indicate significant difference among treatments assessed by Tukey HSD-test ( $p < 0.05$ ).

$\alpha$  remained constant over all three harvest intervals (values not shown); this confirms the validity of the premise that a photosynthetic parameter such as  $\alpha$  will also remain constant under steady-state or constant  $N_s$ . The curvilinear relationship between  $\alpha$  and  $N_L$  for faba bean was quantitatively very similar to those reported for soybean, wheat, and maize by Sinclair and Horie (1989).

**$P_{Nmax}$  versus  $N_s$ :** Again it may be asked: How does  $N_L$  affect  $P_{Nmax}$ , given the fact that the total leaf area per plant and the total number of catalytic units per unit leaf area increases markedly with increasing  $N_s$ ?  $P_{Nmax}$  increased in a curvilinear fashion with respect to  $N_L$  (Fig. 4) and the empirical equation of Sinclair and Horie (1989) was used for deriving the asymptotic value for  $P_{Nmax}$  and the  $N_L$ -intercept value for  $P_{Nmax} = 0$ . With increasing leaf  $N_c$ ,  $P_{Nmax}$  converged onto a maximum asymptotic value of  $33.400 \pm 2.563 \mu\text{mol}(\text{CO}_2) \text{m}^{-2} \text{s}^{-1}$ . Maximum asymptotic values with increasing  $N_L$  measured for maize, soybean, and rice were 57, 36, and  $34 \mu\text{mol}(\text{CO}_2) \text{m}^{-2} \text{s}^{-1}$ , respectively (Sinclair and Horie 1989).

**$C_i$  and  $C_e$  versus  $N_s$ :** In *V. faba* as  $N_L$  increased,  $C_i$  declined exponentially at an exponential of  $3.53 \pm 0.29 \mu\text{mol mol}^{-1} [\text{g(N) m}^{-2}]^{-1}$  to an asymptotic minimum  $C_i$  of  $115.80 \pm 1.59 \mu\text{mol mol}^{-1}$ . A strongly rectangular hyper-

## Discussion

**Relationship between  $\alpha$  and  $N_L$ :** In this study  $\alpha$  with respect to increasing  $N_L$  reached a maximum of  $0.0664 \pm 0.0049 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$ . Meinzer and Zhu (1998) reported that  $\alpha$  for  $\text{CO}_2$  uptake in sugarcane increased linearly from 0.042 to  $0.075 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$  with leaf  $N_c$ . Approximately doubling as leaf  $N_c$ , it was increased from 0.63 to  $1.54 \text{ g m}^{-2}$ . This dependence of  $\alpha$  on leaf  $N_c$  was similar for different sugarcane clones. Hence any dynamic model of leaf photosynthesis should not assume that the value of  $\alpha$  is not influenced by tissue  $N_c$ . Other  $\alpha$  measurements for  $\text{CO}_2$  uptake [measured at leaf temperatures of  $30^\circ\text{C}$ ,  $330 \text{ cm}^3(\text{CO}_2) \text{ m}^{-3}$ , and  $21 \text{ kPa}(\text{O}_2)$ ] in various monocot and dicot plants, with different photosynthetic pathways, give the following range of  $\alpha$  values [ $\mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$ ]:  $C_3$  dicots,  $0.052 \pm 0.001$ ;  $C_3$  grasses,  $0.053 \pm 0.001$ ;  $C_4$  (NAD-ME) dicots,  $0.053 \pm 0.001$ ;  $C_4$  (NAD-ME) grasses,  $0.060 \pm 0.001$ ;  $C_4$  (PCK) grasses,  $0.060 \pm 0.002$ ;  $C_4$  (NADP-ME) dicots,  $0.061 \pm 0.002$ ;  $C_3$  (NADP-ME) grasses,  $0.065 \pm 0.001$ ;  $C_4$  (ME-MIX) dicot, 0.057 (Percy and Ehleringer 1984). Ehleringer and Björkman (1977) found  $\alpha$  value for  $C_3$  photosynthesis of  $0.081 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$  when oxygenase activity was suppressed (low  $\text{O}_2$  or high  $\text{CO}_2$  concentrations). At a leaf temperature of  $20^\circ\text{C}$ ,  $\alpha$  of 0.066 and  $0.044 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$  was found for field grown ryegrass and clover, respectively (unpublished data of Papadopoulos). It is thus reasonable to assume that depending on leaf  $N_c$ ,  $\alpha$  for  $\text{CO}_2$  uptake in  $C_3$  and  $C_4$  plants can range from 0.016 to  $0.075 \mu\text{mol}(\text{CO}_2) \mu\text{mol}(\text{quantum})^{-1}$ .

**Relationship between  $P_{N_{\max}}$  and  $N_L$ :** For faba bean, the  $N_L$ -intercept for  $P_{N_{\max}}$  was  $0.7100 \pm 0.0345 \text{ g(N) m}^{-2}$ . In other studies the linear response of  $P_{N_{\max}}$  to  $N_L$  rose from: zero at  $1.0 \text{ g(N) m}^{-2}$  to about  $2.4 \text{ g(N) m}^{-2}$  for soybean, zero at  $0.3$ – $1.6 \text{ g(N) m}^{-2}$  for rice, and 0 at  $0.2$ – $0.6 \text{ g(N) m}^{-2}$  for maize (Sinclair and Horie 1989). In *Chenopodium album* ( $C_3$ ) and *Amaranthus retroflexus* ( $C_4$ ), the  $P_{N_{\max}}$  intercept of the  $N_L$  axis ranged between  $0.64$ – $0.78 \text{ g(N) m}^{-2}$  (Sage and Percy 1987), for a given N supply rate the values for the corresponding  $P_{N_{\max}}$  remained constant over all three harvest intervals (Fig. 2). This confirms the validity of the premise that a photosynthetic parameter such as  $P_{N_{\max}}$  will also remain constant under steady-state  $N_s$ .

**Relationship between  $C_i$ ,  $C_e$ , and  $N_L$ :** The steady-state functional  $C_i/C_a$  ratio is not only determined by plant water status and stomatal conductance, the  $C_i/C_a$  ratio is also influenced by  $N_L$ . With regard to benchmarking  $C_i$  values, it is generally accepted that at  $25^\circ\text{C}$ , under

bolic relationship was observed between  $C_e$  and  $N_L$  (Fig. 4) with a maximum  $C_e$  asymptotic value of  $1.645 \pm 0.054 \mu\text{mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1} \text{ Pa}(\text{CO}_2)^{-1}$  and with an  $N_L$ -intercept equal to  $0.596 \pm 0.096 \text{ g(N) m}^{-2}$ .

saturating PPFD and ambient  $\text{CO}_2$ ,  $C_i$  is approximately  $100 \mu\text{mol mol}^{-1}$  for  $C_4$  species and  $250 \mu\text{mol mol}^{-1}$  for  $C_3$  species. The  $\text{CO}_2$  compensation concentration ( $\Gamma$ ) is close to 0 for  $C_4$  versus  $5.0 \mu\text{mol mol}^{-1}$  for  $C_3$  species (Evans and Caemmerer 1996).  $C_i$  values for  $C_3$  plants such as *Pinus pinaster* range from 103 to  $266 \mu\text{mol mol}^{-1}$  (Warren *et al.* 2000, Warren and Adams 2001).

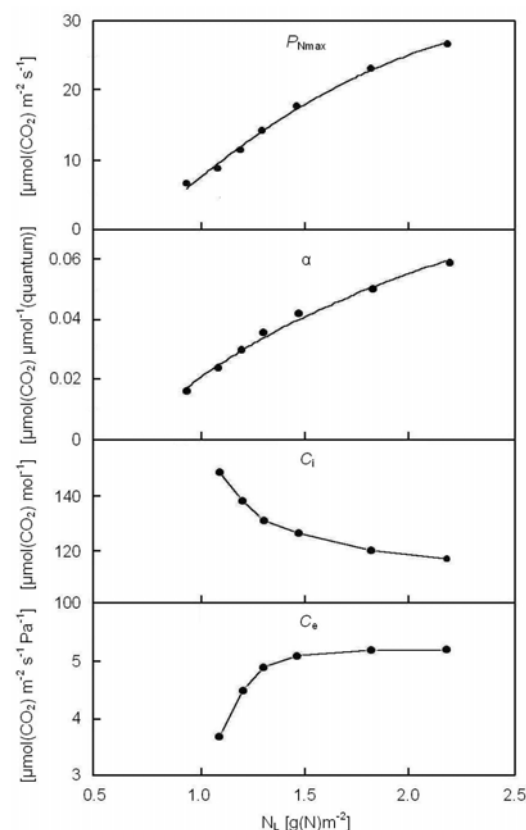


Fig. 4. The influence of increasing specific leaf nitrogen ( $N_L$ ) on photon saturated  $\text{CO}_2$  assimilation rate ( $P_{N_{\max}}$ ), quantum yield efficiency ( $\alpha$ ), leaf intercellular  $\text{CO}_2$  concentration ( $C_i$ ), and the carboxylation efficiency ( $C_e$ ).

$C_e$  has been expressed as the reciprocal of mesophyll resistance  $r_m$  (Edwards and Walker 1983). The following range of  $r_m$  values [ $\text{s cm}^{-1}$ ] have been reported for  $C_3$  plants: *Glycine max* 2.0–3.0, *Atriplex hastata* 2.6, *Phaseolus* spp. 2.6, *Triticum aestivum* 2.8, *Solanum tuberosum* 5.4, and *Medicago sativa* 2.8. Thus the carboxylation efficiencies for the above  $C_3$  plants range between  $0.002$ – $0.005 \text{ m s}^{-1}$ . In other experiments the following carboxylation efficiencies [ $\text{m s}^{-1}$ ] have been reported for  $C_3$  plants: *Helianthus* 0.0063, *Xanthium* 0.0048, *Vigna* 0.0036–0.0047, *Tilia* 0.0035, *Ficus*

0.0031, and *Citrus* 0.0018–0.0024 (Laisk and Loreto 1996).

Assuming a temperature of 20 °C and air pressure of 101.315 Pa [under these conditions CO<sub>2</sub> per 1 000 cm<sup>3</sup> = 41.6×10<sup>-6</sup> mol(CO<sub>2</sub>) m<sup>-3</sup> = 0.101 Pa(CO<sub>2</sub>)] the range of C<sub>e</sub> values reported for various C<sub>3</sub> plants can be recalculated to give the following equivalent values of 0.741–2.595

mol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup> Pa(CO<sub>2</sub>)<sup>-1</sup>. In general, C<sub>e</sub> ranged between 1.046–1.478 μmol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup> Pa(CO<sub>2</sub>)<sup>-1</sup> for C<sub>3</sub> *Cyperus* species and between 2.529–4.123 μmol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup> Pa(CO<sub>2</sub>)<sup>-1</sup> for C<sub>4</sub> *Cyperus* species (Li 1993). Thus the C<sub>e</sub> values estimated for faba bean fell within the C<sub>3</sub> benchmark range of values.

## References

- Anderson, J.M., Ingram, J.S.I.: Chemical analysis. – In: Tropical Soil Biology and Fertility - A Handbook of Methods. 2<sup>nd</sup> Ed. Pp. 70-89. CAB International, Wallingford 1993.
- Charles-Edwards, D.A.: An analysis of the photosynthesis and productivity of vegetative crops in the United Kingdom. – *Ann. Bot.* **42**: 717-731, 1978.
- Cockburn, W., Baldry, C.W., Walker, D.A.: Oxygen evolution by isolated chloroplasts with carbon dioxide as the hydrogen acceptor. A requirement for orthophosphate or pyrophosphate. – *Biochim. biophys. Acta* **131**: 594-596, 1967a.
- Cockburn, W., Baldry, C.W., Walker, D.A.: Some effects of inorganic phosphate on O<sub>2</sub> evolution by isolated chloroplasts. – *Biochim. biophys. Acta* **143**: 614-624, 1967b.
- Dorich, J.A., Nelson, D.W.: Direct colorimetric measurement of ammonium in potassium chloride extracts in soil. – *Soil Sci. Soc. Amer. J.* **55**: 171-178, 1983.
- Edwards, G., Walker, D.: C<sub>3</sub>, C<sub>4</sub>: Mechanisms, and Cellular and Environmental Regulation of Photosynthesis. – Blackwell Scientific Publications, Oxford 1983.
- Ehleringer, J., Björkman, O.: Quantum yields for CO<sub>2</sub> uptake in C<sub>3</sub> and C<sub>4</sub> plants. Dependence on temperature, CO<sub>2</sub>, and O<sub>2</sub> concentrations. – *Plant Physiol.* **59**: 86-90, 1977.
- Evans, J.R., Caemmerer, S. von: Carbon dioxide diffusion inside leaves. – *Plant Physiol.* **110**: 339-346, 1996.
- Fisher, M.J., Charles-Edwards, D.A., Ludlow, M.W.: An analysis of the effects of repeated short-term soil water deficits on stomatal conductance to carbon dioxide and leaf photosynthesis by the legume *Macroptilium atropurpureum* cv. Siratro. – *Aust. J. Plant Physiol.* **8**: 347-357, 1981.
- Garnier, E., Koch, G.W., Roy, J., Mooney, H.A.: Responses of wild plants to nitrate availability: Relationships between growth rate and nitrate uptake parameters, a case study with two *Bromus* species, and a survey. – *Oecologia* **79**: 542-550, 1989.
- Hirose, T.: Nitrogen uptake and plant growth II. An empirical model of vegetative growth and partitioning. – *Ann. Bot.* **58**: 487-496, 1986.
- Katul, G.G., Ellsworth, D.S., Lai, C.T.: Modelling assimilation and intercellular CO<sub>2</sub> from measured conductance: a synthesis of approaches. – *Plant Cell Environ.* **23**: 1313-1328, 2000.
- Laisk, A., Loreto, F.: Determining photosynthetic parameters from leaf CO<sub>2</sub> exchange and chlorophyll fluorescence. Ribulose-1,5-bisphosphate carboxylase/oxygenase specificity factor, dark respiration in the light, excitation distribution between photosystems, alternative electron transport rate, and mesophyll diffusion resistance. – *Plant Physiol.* **110**: 903-912, 1996.
- Mächler, F., Schnyder, H., Nösberger, J.: Influence of inorganic phosphate on photosynthesis of wheat chloroplasts. I. Photosynthesis and assimilate export at 5 °C and 25 °C. – *J. exp. Bot.* **35**: 481-487, 1984.
- Li, M.: Leaf photosynthetic nitrogen-use efficiency of C<sub>3</sub> and C<sub>4</sub> *Cyperus* species. – *Photosynthetica* **29**: 117-130, 1993.
- Meinzer, F.C., Zhu, J.: Nitrogen stress reduces the efficiency of C<sub>4</sub> CO<sub>2</sub> concentrating system and therefore quantum yield, in *Saccharum* (sugarcane) species. – *J. exp. Bot.* **49**: 1227-1234, 1998.
- Pearcy, R.W., Ehleringer, J.: Comparative ecophysiology of C<sub>3</sub> and C<sub>4</sub> plants. – *Plant Cell Environ.* **7**: 1-13, 1984.
- Pradet, A., Raymond, P.: Adenine nucleotide ratios and adenylate energy charge in energy metabolism. – *Annu. Rev. Plant Physiol.* **34**: 199-224, 1983.
- Rao, I.M., Arulanantham, A.R., Terry, N.: Leaf phosphate status, photosynthesis and carbon partitioning in sugar beet. II. Diurnal change in sugar phosphates, adenylates, and nicotinamide nucleotides. – *Plant Physiol.* **90**: 820-826, 1989a.
- Rao, I.M., Arulanantham, A.R., Terry, N.: Diurnal change in sugar phosphates, adenylates, and nicotinamide nucleotides in sugar beet leaves. – *Photosynth. Res.* **23**: 205-212, 1989b.
- Rao, I.M., Terry, N.: Leaf phosphate status, photosynthesis and carbon partitioning in sugar beet. I. Changes in growth, gas exchange, and Calvin cycle enzymes. – *Plant Physiol.* **90**: 814-819, 1989.
- Rao, I.M., Terry, N.: Leaf phosphate status, photosynthesis, and carbon partitioning in sugar beet. IV. Changes with time following increased supply of phosphate to low-phosphate plants. – *Plant Physiol.* **107**: 1313-1321, 1995.
- Sage, R.F., Pearcy, R.W.: The nitrogen use efficiency of C<sub>3</sub> and C<sub>4</sub> plants. I. Leaf nitrogen, growth and biomass partitioning in *Chenopodium album* (L.) and *Amaranthus retroflexus* (L.). – *Plant Physiol.* **84**: 954-958, 1987.
- Schmitt, M.R., Edwards, G.E.: Photosynthetic capacity and nitrogen use efficiency of maize, wheat, and rice: a comparison between C<sub>3</sub> and C<sub>4</sub> photosynthesis. – *J. exp. Bot.* **32**: 459-466, 1981.
- Sinclair, T.R., Horie, T.: Leaf nitrogen, photosynthesis and crop radiation use efficiency: A review. – *Crop Sci.* **29**: 90-98, 1989.
- Thornley, J.H.M.: Modelling shoot:root relations: the only way forward? – *Ann. Bot.* **81**: 165-171, 1998.
- Usuda, H., Edward, G.E.: Influence of varying CO<sub>2</sub> and orthophosphate concentrations on rates of photosynthesis, and synthesis of glycolate and dihydroxyacetone phosphate by wheat chloroplasts. – *Plant Physiol.* **69**: 469-473, 1982.
- Warren, C.R., Adams, M.A.: Distribution of N, Rubisco and photosynthesis in *Pinus pinaster* and acclimation to light. – *Plant Cell Environ.* **24**: 598-609, 2001.
- Warren, C.R., Adams, M.A., Chen, Z.L.: Is photosynthesis related to concentration of nitrogen and Rubisco in leaves of Australian native plants? – *Aust. J. Plant Physiol.* **27**: 407-416, 2000.