

# Photosynthetic pathway types in rangeland plant species from Inner Mongolia, North China

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## Abstract

Photosynthetic pathway types, based on  $\delta^{13}\text{C}$  measurements, were determined for 125 species in 95 genera and 32 families growing in rangelands from Inner Mongolia. Of the total species, 4 species from 3 genera and 2 families had  $\text{C}_4$  photosynthesis (2 species in *Gramineae* and 2 in *Chenopodiaceae*) and 118 species from 90 genera and 31 families had  $\text{C}_3$  photosynthesis. The number of  $\text{C}_4$  species differed significantly among four rangeland sites, 4 species in desert, 3 species in steppe, but no  $\text{C}_4$  species were identified in meadow and dune. Six species [e.g. *Agriophyllum arenarium* Bieb., *Bassia dasyphylla* O. Kuntze, *Saussurea japonica* (Thunb.) DC.] earlier identified as  $\text{C}_4$  species using the enzyme ratio method were found as  $\text{C}_3$  species using the carbon isotope ratios ( $\delta^{13}\text{C}$ ). Hence the enzyme ratio method for  $\text{C}_3$  and  $\text{C}_4$  identification may not always be reliable. The  $\delta^{13}\text{C}$  values of 3 species of *Crassulaceae*, which had been considered as CAM species, differed remarkably [ $-25.79\text{‰}$  for *Sedum aizoon* L.,  $-24.42\text{‰}$  for *Osostachys fimbriatus* (Turcz.) Berger, and  $-16.97\text{‰}$  for *O. malacophyllus* (Pall.) Fisch], suggesting that the use of  $\delta^{13}\text{C}$  method as a diagnosis for CAM photosynthetic pathway type may not always be reliable and supplementary measurements are needed.

*Additional key words:*  $\text{C}_3$ ,  $\text{C}_4$ , and CAM plants; carbon isotope ratio; Xilingol steppe;  $\delta^{13}\text{C}$ .

## Introduction

Photosynthetic pathway type is not only the basis for studying plant functional types, but also relates with ecological processes, e.g. community succession, climate changes (Collatz *et al.* 1998, Wang 2003d), as well as bridges the micro-ecology and macro-ecology.  $\text{C}_4$  plant identification began with the works of Downton and Tregunna (1968) and Black (1971), and more than 1 700  $\text{C}_4$  plants have been identified worldwide (Li 1993). It is expected that approximately one-half of the 10 000 grass species and a thousand of the 165 000 *Dicotyledoneae* in the world have  $\text{C}_4$  photosynthetic pathway (Hattersley 1987, Ehleringer *et al.* 1997) indicating that only 1/3 of the  $\text{C}_4$  species was identified and the other 2/3 remains unclear. Various methods, e.g. carbon isotope ratio ( $\delta^{13}\text{C}$ ), "Kranz" type leaf anatomy, low  $\text{CO}_2$  compensation concentration, and photosynthetic enzyme ratio, have been used for photosynthetic pathways classification, but many studies suggested that the carbon isotope ratio method is most reliable (Redmann *et al.* 1995).

Desert and steppe are two typical rangeland vegetations in Inner Mongolia, North China. Superior both in quality and quantity of herbage production in the usual growing condition of the rangelands make them best suited for livestock grazing. Many studies of local flora, community classification, plant biomass, and photosynthetic pathways have been conducted in the region, but systematic research on identification of  $\text{C}_3$ ,  $\text{C}_4$ , and CAM species by using carbon isotope ratio in the rangelands has not been done. Photosynthetic pathways are important linkages between plant species and ecosystems, and rangeland management decisions must take into account their knowledge which is closely related to seasonal development patterns (warm season *versus* cool season).

The objective of this study was to determine  $\text{C}_3$  and  $\text{C}_4$  photosynthetic pathways from rangelands in Inner Mongolia by using carbon isotope ratio and to uncover the relationship between pathway type and plant habitats.

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The results may help in studying the correlation of rangeland species composition with ecosystem, climate varia-

bles, and grazing management.

## Materials and methods

The study was conducted in typical desert and steppe vegetations in Xilingol rangelands (41°35'–46°46'N, 111°09'–119°58'E), which is part of Mongolian plateau, and covers about 200 000 km<sup>2</sup> area, with average 1 200 m above sea level, varying from 900 m in the north-east to 1 300 m in the south-west. Most of the rangelands have chestnut, chernozem, and sandy soils. Located in semi-arid area of mid-temperate zone, the rangelands have the characteristics of temperate steppe climate, with air temperature and precipitation varying sharply all round the year. Mongolia anticyclone and moist Pacific air mass are the two major factors determining the climate type in this region. The sharp gradient between the high-pressure and Aleutian low-pressure system bring a strong westerly flow of cold, dry continental air at the surface. As the anticyclone breaks down in spring, the rangeland region comes increasingly under the influence of moist Pacific air mass, reaching a climax in the summer monsoon, which lasts 2–3 months. The typical climate characteris-

tics of the region are: cold dry and frequently windy spring; warm and wet summer, but with possible seasonal droughts, early autumn frosts, and long cold winter with less snowfall. The mean annual air temperature ranges from 0 to 3 °C, varying from –21 °C in January to 21 °C in July. Moisture gradient is very steep, with annual precipitation varying from 100 mm in the western desert to 400 mm in the eastern meadow. Precipitation mainly falls between June and August over the growing season.

Four types of sites, *e.g.* meadow, steppe, desert, and dune, were selected for the plant sampling. Shoot tissue was collected from field-grown plants, dried at 80 °C for 48 h, and ground. Stable carbon isotope ratio ( $\delta^{13}\text{C}$ ) in the plant tissue was determined by using *Delta<sup>plus</sup> XP* mass spectrograph. Plants with  $\delta^{13}\text{C}$  values above –19 ‰ were considered to have C<sub>4</sub> photosynthetic pathway, and with  $\delta^{13}\text{C}$  values less than –21 ‰ to have C<sub>3</sub> photosynthesis (Smith and Brown 1973) (Table 1).

Table 1. Photosynthetic pathway types (C<sub>3</sub>, C<sub>4</sub>) in species from rangelands of Inner Mongolia, North China. Nomenclature follows Redmann *et al.* (1995). Habitats; ME = meadow, TS = typical steppe, DS = desert and DU = dune.

Species	$\delta^{13}\text{C}$ [‰]	C <sub>3</sub> /C <sub>4</sub>	Habitat
<i>Gymnospermae</i>			
<i>Ephedraceae</i> <i>Ephedra sinica</i> Stapf	–24.449	C <sub>3</sub>	DS
<i>Angiospermae</i>			
<i>Dicotyledoneae</i>			
<i>Asclepiadaceae</i> <i>Cynanchum thesioides</i> (Freyn) K. Schum.	–25.167	C <sub>3</sub>	DS
<i>Boraginaceae</i> <i>Lappula redowskii</i> (Horn) Green	–28.594	C <sub>3</sub>	TS
<i>Campanulaceae</i> <i>Adenophora stenanthina</i> (Ledeb.) Kitag.	–26.560	C <sub>3</sub>	ME
<i>Caryophyllaceae</i> <i>Dianthus chinensis</i> L.	–25.734	C <sub>3</sub>	ME
<i>Silene repens</i> Patr.	–25.765	C <sub>3</sub>	ME
<i>Chenopodiaceae</i> <i>Agriophyllum arenarium</i> Bieb.	–25.340	C <sub>3</sub>	DU
<i>Axyris amarantoides</i> L.	–26.691	C <sub>3</sub>	TS
<i>Bassia dasyphylla</i> O. Kuntze	–25.113	C <sub>3</sub>	DS
<i>Chenopodium aristatum</i> L.	–26.148	C <sub>3</sub>	TS
<i>Ch. glaucum</i> L.	–27.213	C <sub>3</sub>	TS
<i>Corispermum declinatum</i> Steph. ex Stev.	–26.841	C <sub>3</sub>	DU
<i>Kochia prostrata</i> (L.) Schrad.	–14.036	C <sub>4</sub>	TS DS
<i>Salsola collina</i> Pall.	–11.400	C <sub>4</sub>	TS DS
<i>Compositae</i> <i>Artemisia annua</i> L.	–27.306	C <sub>3</sub>	ME
<i>A. dracunculus</i> L.	–26.941	C <sub>3</sub>	ME
<i>A. eriopoda</i> Bunge	–27.669	C <sub>3</sub>	ME
<i>A. frigida</i> Willd.	–28.125	C <sub>3</sub>	TS DS ME
<i>A. intramongolica</i> H.C. Fu.	–25.840	C <sub>3</sub>	DU
<i>A. oxycephala</i> Kitag.	–25.783	C <sub>3</sub>	DU
<i>A. pectinata</i> Pall. [ <i>Neopallasia pectinata</i> (Pall.) Polsak.]	–24.204	C <sub>3</sub>	DS
<i>A. scoparia</i> Waldst et Kit	–27.869	C <sub>3</sub>	DS
<i>Aster alpinus</i> L.	–28.003	C <sub>3</sub>	ME

Table 1 (continued)

	Species	$\delta^{13}\text{C}$ [‰]	C <sub>3</sub> /C <sub>4</sub>	Habitat
Compositae	<i>Filifolium sibiricum</i> (L.) Kitam.	-26.835	C <sub>3</sub>	ME
	<i>Heteropappus altaicus</i> (Willd.) Novopokr.	-26.895	C <sub>3</sub>	DS
	<i>Hieracium umbellatum</i> L.	-27.424	C <sub>3</sub>	ME
	<i>Ixeris chinensis</i> (Thunb.) Nakaisubsp <i>graminifolia</i> (Ledeb.) Kitag.	-26.835	C <sub>3</sub>	DU
	<i>Ligularia mongolica</i> (Turcz.) DC.	-26.900	C <sub>3</sub>	ME
	<i>Olgaea leucophylla</i> (Turcz.) Iijin	-27.205	C <sub>3</sub>	TS ME
	<i>Saussurea japonica</i> (Thunb.) DC.	-27.258	C <sub>3</sub>	TS ME
	<i>Scorzonera austriaca</i> Willd.	-25.373	C <sub>3</sub>	ME
	<i>Serratula coronata</i> L.	-28.079	C <sub>3</sub>	TS ME
	<i>Stemmacantha uniflora</i> (L.) Dittrich	-27.720	C <sub>3</sub>	ME
	<i>Youngia tenuifolia</i> (Willd.) Babc.	-23.829	C <sub>3</sub>	DS
Convolvulaceae	<i>Convolvulus ammannii</i> Desr.	-25.271	C <sub>3</sub>	DS
	<i>C. arvensis</i> L.	-25.271	C <sub>3</sub>	DS
Crassulaceae	<i>Osostachys fimbriatus</i> (Turcz.) Berger	-24.420	C <sub>3</sub>	ME
	<i>O. alacophyllus</i> (Pall.) Fisch	-16.965	C <sub>4</sub>	TS DU
	<i>Sedum aizoon</i> L.	-25.788	C <sub>3</sub>	ME
Cruciferae	<i>Dontostemon micranthus</i> C.A. Mey.	-25.534	C <sub>3</sub>	TS
	<i>Lepidium apetalum</i> Willd.	-25.478	C <sub>3</sub>	TS
	<i>Ptilotrichum tenuifolium</i> (Steoh) C.A. Mey.	-26.524	C <sub>3</sub>	TS
Dipsacaceae	<i>Scabiosa comosa</i> Fisch. ex Roemer Schult	-26.600	C <sub>3</sub>	DU
	<i>S. tschiliensis</i> Crrunning	-27.039	C <sub>3</sub>	ME
Labiatae	<i>Lagochilus ilicifolius</i> Bungein	-25.895	C <sub>3</sub>	DS
	<i>Phlomis tuberosa</i> L.	-25.942	C <sub>3</sub>	TS
	<i>Schizonepeta multifida</i> (L.) Briq.	-27.520	C <sub>3</sub>	TS ME
	<i>Scutellaria baicalensis</i> Georgi	-26.989	C <sub>3</sub>	ME
	<i>S. scordifolia</i> Fisch. ex Schrank	-27.544	C <sub>3</sub>	DU
Leguminosae	<i>Astragalus melilotoides</i> Pall.	-26.466	C <sub>3</sub>	ME
	<i>Caragana microphylla</i> Lam.	-24.483	C <sub>3</sub>	TS
	<i>C. stenophylla</i> Pojark.	-24.572	C <sub>3</sub>	DS
	<i>Hedysarum fruticosum</i> Pall. var. <i>lignosum</i> (Trautv.) Kitag.	-27.416	C <sub>3</sub>	DU
	<i>Medicago falcate</i> L.	-27.096	C <sub>3</sub>	ME
	<i>Melilotoides ruthenia</i> (L.) Sojak	-26.956	C <sub>3</sub>	DU
	<i>Oxytropis myriophylla</i> (Pall.) DC.	-27.481	C <sub>3</sub>	ME
	<i>Thermopsis lanceolata</i> R. Br.	-25.873	C <sub>3</sub>	TS ME
	<i>Trifolium lupinaster</i> L.	-25.314	C <sub>3</sub>	ME
	<i>Vicia multicaulis</i> Ldb.	-25.629	C <sub>3</sub>	ME
	<i>V. unijuga</i> R. Br.	-24.735	C <sub>3</sub>	ME
Linaceae	<i>Linum perenne</i> L.	-25.604	C <sub>3</sub>	TS
Urticaceae	<i>Urtica cannabina</i> L.	-26.725	C <sub>3</sub>	DU
Papaveraceae	<i>Papaver nudicaule</i> L.	-28.050	C <sub>3</sub>	DU
Plumbaginaceae	<i>Limonium bicolor</i> (Bunge) Q. Kuntze	-30.125	C <sub>3</sub>	TS
Polygonaceae	<i>Polygonum divaricatum</i> L.	-26.855	C <sub>3</sub>	TS ME DU
	<i>Polygonum viviparum</i> L.	-27.345	C <sub>3</sub>	SS
	<i>Rheum undulatum</i> L.	-28.628	C <sub>3</sub>	ME
Ranunculaceae	<i>Clematis hexapetala</i> Pall.	-24.343	C <sub>3</sub>	ME
	<i>Pulsatilla turczaninovii</i> Kryl. et Serg.	-26.770	C <sub>3</sub>	ME
	<i>Thalictrum petaloideum</i> L.	-27.685	C <sub>3</sub>	TS ME
	<i>Th. squarrosum</i> Steh. ex Willd.	-26.134	C <sub>3</sub>	ME
	<i>Delphinium grandiflorum</i> L.	25.878	C <sub>3</sub>	ME
Rosaceae	<i>Prunus sibirica</i> L.	-25.360	C <sub>3</sub>	DU
	<i>Potentilla acaulis</i> L.	-27.263	C <sub>3</sub>	TS ME
	<i>P. betonicaefolia</i> Poir	-25.330	C <sub>3</sub>	ME
	<i>P. bifurca</i> L.	-25.565	C <sub>3</sub>	ME TS

Table 1 (continued)

	Species	$\delta^{13}\text{C}$ [‰]	C <sub>3</sub> /C <sub>4</sub>	Habitat
Rosaceae	<i>P. nudicaulis</i> Willd.	-26.201	C <sub>3</sub>	ME
	<i>P. tanacetifolia</i> Willd. ex Schlecht	-26.852	C <sub>3</sub>	ME TS
	<i>P. verticillaris</i> Steph. ex Willd.	-26.609	C <sub>3</sub>	ME
	<i>Sanguisorba officinalis</i> L.	-26.758	C <sub>3</sub>	ME
	<i>Spiraea aquilegifolia</i> Pall.	-27.571	C <sub>3</sub>	TS ME ME
Rubiaceae	<i>Galium verum</i> L.	-25.810	C <sub>3</sub>	TS ME
Rutaceae	<i>Haplophyllum dauricum</i> (L.) Juss.	-27.040	C <sub>3</sub>	TS DS
Saxifragaceae	<i>Ribes diacanthum</i> Pall.	-27.934	C <sub>3</sub>	DU
Scrophulariaceae	<i>Cymbarraria dahurica</i> L.	-27.768	C <sub>3</sub>	TS ME
	<i>Linaria vulgaris</i> Mill.	-27.748	C <sub>3</sub>	ME
	<i>Pedicularis striata</i> Pall.	-27.771	C <sub>3</sub>	ME
	<i>Veronica linariifolia</i> Pall. ex Link	-27.124	C <sub>3</sub>	ME
Thymelaceae	<i>Stellera chamaejasme</i> L.	-25.780	C <sub>3</sub>	ME
Umbelliferae	<i>Bupleurum scorzonerifolium</i> Willd.	-25.800	C <sub>3</sub>	ME TS
	<i>Peucedanum rigidum</i> Bunge	-24.595	C <sub>3</sub>	DS DU
	<i>Siler divaricatum</i> Benth. et Hook.	-26.620	C <sub>3</sub>	DS ME
	<i>Sphallerocarpus gracilis</i> (Bess.) K. Pol.	-23.913	C <sub>3</sub>	ME
Valerianaceae	<i>Patrinia rupestris</i> Juss.	-26.144	C <sub>3</sub>	ME
Zygophyllaceae	<i>Peganum harmala</i> L.	-23.361	C <sub>3</sub>	DS
Monocotyledoneae				
Cyperaceae	<i>Carex duriuscula</i> C.A. Mey.	-24.354	C <sub>3</sub>	TS
	<i>C. pediformis</i> C.A. Mey.	-25.257	C <sub>3</sub>	ME
	<i>C. korshinskyi</i> Kom.	-25.831	C <sub>3</sub>	TS
Gramineae	<i>Achnatherum sibiricum</i> (L.) Keng	-25.731	C <sub>3</sub>	ME
	<i>Agropyron cristatum</i> (L.) Gaertn.	-27.157	C <sub>3</sub>	TS DS ME
	<i>Bromus inermis</i> Leyss.	-24.882	C <sub>3</sub>	ME
	<i>Cleistogenes squarrosa</i> (Trin.) Keng	-14.639	C <sub>4</sub>	TS
	<i>C. songorica</i> (Roshev.) Ohwi	-14.757	C <sub>4</sub>	DS
	<i>Elymus dahuricus</i> Turcz.	-25.145	C <sub>3</sub>	ME
	<i>Festuca rubra</i> L.	-27.089	C <sub>3</sub>	ME
	<i>Koeleria cristata</i> (L.) Pers.	-26.347	C <sub>3</sub>	ME
	<i>Leymus chinensis</i> (Trin.) Tzvel.	-26.008	C <sub>3</sub>	TS ME
	<i>Poa pratensis</i> L.	-24.635	C <sub>3</sub>	ME
	<i>Psammochloa villosa</i> (Trin.) Bor	-28.118	C <sub>3</sub>	DU
	<i>Stipa baicalensis</i> Roshev.	-25.920	C <sub>3</sub>	ME
	<i>S. grandis</i> P. Smirn.	-26.541	C <sub>3</sub>	TS
	<i>S. krylovii</i> Roshev.	-25.285	C <sub>3</sub>	DS
Iridaceae	<i>Iris dichotoma</i> Pall.	-24.938	C <sub>3</sub>	ME
	<i>I. lactea</i> Pall. var. <i>chinensis</i> (Fisch) Koidz.	-25.844	C <sub>3</sub>	DS
	<i>I. tenuifolia</i> Pall.	-26.233	C <sub>3</sub>	DU
	<i>I. ventricosa</i> Pall.	-26.078	C <sub>3</sub>	ME
Liliaceae	<i>Allium bidentatum</i> Fisch. ex Prokh.	-25.140	C <sub>3</sub>	DS
	<i>A. polyrhizum</i> Turcz.	-25.805	C <sub>3</sub>	DS
	<i>A. ramosum</i> L.	-25.562	C <sub>3</sub>	DS TS
	<i>A. senescens</i> L.	-25.179	C <sub>3</sub>	DU
	<i>A. tenuissimum</i> L.	-26.089	C <sub>3</sub>	DS
	<i>Anemarrhena asphodeloides</i> Bunge	-24.620	C <sub>3</sub>	TS
	<i>Asparagus dauricus</i> Fisch. ex Link	-24.719	C <sub>3</sub>	DS
	<i>Hemerocallis minor</i> Mill.	-25.811	C <sub>3</sub>	ME
	<i>Veratrum nigrum</i> L.	-23.762	C <sub>3</sub>	ME

## Results

125 vascular plant species in 95 genera and 32 families, about 20 % of the total species in Xilingol rangelands, were examined for photosynthetic pathway types. Carbon isotope ratios indicative of  $C_4$  photosynthesis were found in 4 species from 3 genera and 2 families, 2 were found in *Gramineae* [*Cleistogenes squarrosa* (Trin.) Keng and *C. songorica* (Roshev.) Ohwi] and the other two in *Chenopodiaceae* [*Kochia prostrata* (L.) Schrad. and *Salsola collina* Pall.]. 118 species from 90 genera and 31 families had  $C_3$  photosynthesis. This suggested that the abundance of  $C_4$  plants in these sample sites was very low, even the occurrence of  $C_4$  species in the desert and steppe was common. Three species, *i.e.* *Osostachys fimbriatus* (Turcz.) Berger, *O. malacophyllus* (Pall.) Fisch, and *Sedum aizoon* L. in *Crassulaceae* had been considered as CAM photosynthesis, but their  $\delta^{13}C$  values varied remarkably:  $-25.79\text{‰}$  for *Sedum aizoon* L. and  $-24.42\text{‰}$  for *O. fimbriatus* (Turcz.) Berger, while that for *O. malacophyllus* (Pall.) Fisch was  $-16.97\text{‰}$ . Thus the identification of CAM photosynthetic pathway may need other supplementary measurements.

According to the literature, the photosynthetic pathways of more than 50 % of the total 125 species in Table 1, *e.g.* *Ephedra sinica* Stapf., *Corispermum declinatum* Steph. ex Stev., *Stemmacantha uniflora* (L.) Dittrich, *Scabiosa comosa* Fisch. ex Roemer Schult., were identified for the first time (Table 1). The following species, with  $C_3$  carbon isotope ratios, had earlier been identified as  $C_4$  based on phosphoenolpyruvate carboxylase : ribulose-1,5-bisphosphate carboxylase ratios (Yin and Li 1997, Wang 2002a,c): *Agriophyllum arenarium* Bieb., *Bassia dasyphylla* O. Kuntze, *Saussurea japonica* (Thunb.) DC., *Thalictrum squarrosum* Steh. ex Willd., *Thermopsis lanceolata* R.Br., and *Carex pediformis* C.A.

## Discussion

The identification of photosynthetic pathway types is the basic knowledge for studying the logical linkages between physiological and life-history strategies at plant level, and ecological process at ecosystem and global levels. The  $\delta^{13}C$  value of plant tissue is diagnostic for photosynthetic pathways (Smith and Brown 1973) and is widely used as a criterion for  $C_3$  and  $C_4$  classification (Downton 1975, Li 1993, Redmann *et al.* 1995, Pyankov *et al.* 2000). 125 species from four sites were classified to their types of photosynthesis (Table 1): only 4 species were found with  $C_4$  photosynthesis, while most species had  $C_3$  photosynthesis. Relatively low amount of  $C_4$  species identified in this study was mainly due to the small scale of sampling area (about  $1\text{ km}^2$ ) and species abundance in the four sites, as well as the reduction of  $C_4$  species which had earlier been identified as  $C_4$  species, *e.g.* *Agriophyllum arenarium* Bieb., *Bassia dasyphylla* O. Kuntze, *Saussurea japonica* (Thunb.) DC., *Thalictrum*

Mey. This suggests that the use of the enzyme ratio method as diagnosis for photosynthetic pathway type may not always be reliable and some previous results need to be corrected.

As for floristic composition of the total 125 species, 1  $C_3$  species was found in *Gymnospermae*, 124 species in *Angiospermae*, 94 species (about 25 % of the total *Dicotyledoneae* species in Xilingol rangelands) in 74 genera and 27 families were found in *Dicotyledoneae*, *e.g.* *Compositae* (20 species) *Leguminosae* (11 species), *Rosaceae* (9 species), and *Chenopodiaceae* (8 species), and they were the leading families with 48 species identified (about 51 % of the total 94 species in *Dicotyledoneae*). The other 23 families had a total of 46 species, less than 50 % of the total *Dicotyledoneae* species. 30 species in 19 genera and 4 families were identified in *Monocotyledoneae*, *e.g.* *Gramineae* (14 species), *Liliaceae* (9 species), *Iridaceae* (4 species), and *Cyperaceae* (3 species). These 30 species make up 16 % of the total *Monocotyledoneae* species in this region, indicating that the species abundance of *Monocotyledoneae* was smaller than that of *Dicotyledoneae*.

Four  $C_4$  species were found in desert site and 3 in steppe site, but no  $C_4$  species was identified in meadow and dune sites. The value of  $C_4/C_3$  in desert site was two times that in the steppe site. This suggests that the occurrence of  $C_4$  species was common in the more arid region. Meadow site had most species abundance, with 67 species (about 54 % of the total 125), in, *e.g.*, *Compositae* (12 species), *Gramineae* (8 species), *Leguminosae* (7 species), and *Rosaceae* (6 species). However, only 36 species (about 29 %) and 26 species (21 %) were found in steppe and desert sites, respectively.

*squarrosum* Steh. ex Willd., *Thermopsis lanceolata* R.Br., and *Carex pediformis* C.A. Mey. This suggests that the use of stable carbon isotope ratio ( $\delta^{13}C$ ) was more reliable for  $C_3$  and  $C_4$  identification and some of the previous classifications by using the enzyme, "Kranz" type leaf anatomy, and low  $CO_2$  compensation concentration need to be modified.

Of the total 125 species,  $\delta^{13}C$  values of 3 *Crassulaceae* species, which had been classified as CAM species (Wang 2002a), were remarkably different (Table 1). The  $\delta^{13}C$  values for *Osostachys fimbriatus* (Turcz.) Berger and *Sedum aizoon* L. were less than  $-24.42\text{‰}$ , while that for *O. malacophyllus* (Pall.) Fisch was  $-16.97\text{‰}$ . We can not classify the 3 species into their photosynthetic pathway types by using the  $\delta^{13}C$  values, and supplementary measurements, *e.g.* leaf acidometric titration in the morning and in the dusk (Lin *et al.* 1988) should be taken. This suggests that the use of carbon isotope ratio ( $\delta^{13}C$ )

method as diagnosis for CAM photosynthetic pathway type may not always be reliable and supplementary measurements are needed.

The changes of plant photosynthetic pathway composition are consistent with vegetation dynamics caused by nature and human activities, especially in grassland and deserts (Wang 2002a). Four C<sub>4</sub> species were found in desert site, 3 in steppe site, and no C<sub>4</sub> species was identified in meadow and dune sites. Of the 4 C<sub>4</sub> species identified in Table 1, two were xerophytic grass species [*Cleistogenes squarrosa* (Trin.) Keng and *C. songorica* (Roshev.) Ohwi] and the other two belonged to annual

*Chenopodiaceae* [*Kochia prostrata* (L.) Schrad. and *Salsola collina* Pall.]; this suggests that these species had greater tolerance to drought. Previous studies (Williams and Markley 1973, Wang 2002a,b,c) also clearly documented the importance of photosynthetic pathway types in predicting vegetation dynamics and the occurrence of C<sub>4</sub> species could be an indicator for both diagnosis of grassland conditions and management decisions. Future studies in North China could evaluate the responses of photosynthetic pathways to land use managements.

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