

# Plant functional types and their ecological responses to salinization in saline grasslands, Northeastern China

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## Abstract

Photosynthetic pathways ( $C_3$ ,  $C_4$ , and CAM) and morphological functional types (e.g. shrubs, high perennial grasses, short perennial graminaceous plants, annual grasses, annual forbs, perennial forbs, halophytes, and hydrophytes) were identified for the species from salinity grasslands in Northeastern China, using the data from both stable carbon isotope ratios ( $\delta^{13}\text{C}$ ) and from the references published between 1993 and 2002. 150 species, in 99 genera and 37 families, were found with  $C_3$  photosynthesis, and most of these species are dominants [e.g. *Leymus chinensis* (Trin.) Tzvel., *Calamagrostis epigeios* (L.), *Suaeda corniculata* (C.A. Mey.) Bunge]. 40 species in 25 genera and 8 families were identified with  $C_4$  photosynthesis [e.g. *Chloris virgata* Sw., *Aeluropus littoralis* (Gouan) Parlat] and 1 species with CAM photosynthesis. *Gramineae* is the leading family with  $C_4$  photosynthesis (27 species), *Chenopodiaceae* ranks the second (5 species). The significant increase of  $C_4$  proportions with intense salinity suggested this type plant is remarkable response to the grassland salinization in the region. 191 species were classified into eight morphological functional types and the changes of most of these types (e.g. PEF, HAL, and HPG) were consistent with habitats and vegetation dynamics in the saline grassland. My findings suggest that the photosynthetic pathways, combined with morphological functional types, are efficient means for studying the linkage between species and ecosystems in this type of saline grassland in Northeastern China.

*Additional key words:*  $C_3$ ,  $C_4$ , CAM species; ecological responses; morphological functional types; saline grasslands.

## Introduction

Photosynthetic pathways ( $C_3$ ,  $C_4$ , and CAM) and morphological types are the main composition of plant functional types (PFTs), and the popular means for studying the logical links between physiological and life-history strategies at plant level, as well as ecological process at ecosystem and global levels (Chapin 1993, Paruelo and Lauenroth 1996, Wang 2003). PFTs are defined based on the relevant traits, e.g. plant morphology, physiology, life history, and bio-climatic tolerance, depending on the intentions and studying scales (Wang 2003). Some studies proved that grassland management decisions must take into account photosynthetic and morphological functional types, because they are closely related to seasonal development patterns (warm season *versus* cool season) (Williams and Markley 1973) and grazing succession (Wang 2002a).

Saline grasslands, one of main vegetation types, cover an area of about 17 000 km<sup>2</sup> in the central part of Northeastern China. The usual growing conditions of the grasslands produce herbage superior both in quality and in

quantity, so that this type of grassland is one of the best suited for the rangeland industry in Northeastern China (Wang 2002a). There has, however, been deterioration and salinization of the grasslands since the 1960's, because of the increasing demand for agricultural land and need for more livestock. The substantial reduction in canopy cover throughout much of the grasslands, combined with soil salinization, has led to considerable changes in vegetation, e.g. plant composition, distribution, and plant functional types. Studies on the grasslands, e.g. community classification, plant biomass, and grazing effects, have been conducted in the region (Zheng and Li 1993, Wang 2002a), but the patterns of photosynthetic and morphological functional types remain unclear. Present study investigated the composition of photosynthetic pathways and morphological functional types and their ecological roles in the saline grasslands. The results could be important for the interpretation of relationships between the changes of plant functional types and grassland use.

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## Materials and methods

This study was conducted in the Songnen grasslands ( $43^{\circ}30' \sim 48^{\circ}40'N$ ,  $121^{\circ}30' \sim 127^{\circ}00'E$ ), typical saline grasslands in Northeastern China. The grasslands are widely distributed at the eastern end of the Eurasian steppe zone, on average 141 m above sea level, varying from 137.8 to 144.9 m. Most of the region has meadow chernozem soil, with 3.5 to 6.0 % organic matter in the surface layer. Average soil pH for this soil type is about 8.7, rising as high as 10.0 in the spring (Wang and Ripley 1997). For at least 20 years prior to 2003, the grasslands were grazed and mowed, and transient floods may occur in autumn. In winter, the climate of the area is dominated by the Mongolian anticyclone, which produces a westerly flow of cold, dry air and little snowfall. As the anticyclone breaks down in spring, the region comes increasingly under the influence of moist Pacific air masses, reaching a climax in the summer monsoon, which lasts 2–3 months. The mean annual air temperature is about 5 °C, varying from –18 °C in January to 23 °C in July. Annual precipitation varies from 300 to 600 mm, falling mainly during the summer monsoon. Precipitation is not distributed evenly over growing season, of which 70 % falls between June

and August in this region.

Floristic species were obtained from the sampling data conducted from 1986 to 2003 and from references published from 1993 to 2002 (Zheng and Li 1993, Wang 2002a). Photosynthetic pathway types for the forage species were determined by stable carbon isotope ratio ( $\delta^{13}\text{C}$ ) and from the references published between 1985 and 2002 (e.g. Takeda and Hakoyama 1985, Li 1993, Redmann *et al.* 1995, Yin and Li 1997, Pyankov *et al.* 2000, Wang 2002c,d). Plants with  $\delta^{13}\text{C}$  values above –19 ‰ were considered to have the  $\text{C}_4$  photosynthesis, and  $\delta^{13}\text{C}$  values lower than –21 ‰ to have  $\text{C}_3$  photosynthesis (Redmann *et al.* 1995). Plants identified in the region were classified into both photosynthetic pathway types by physiological attributes ( $\delta^{13}\text{C}$ ) and morphological functional types, e.g. shrubs (SHR), high perennial grasses (HPG), short perennial graminaceous plants (grasses and sedge) (SPG), annual grasses (ANG), annual forbs (ANF), perennial forbs (PEF), halophytes (HAL), and hydrophytes (HYD) by morphological attributes (Table 1).

Table 1. Photosynthetic pathway ( $\text{C}_3$  or  $\text{C}_4$ ) and morphological functional types in saline grasslands, Northeastern China. Nomenclature follows Kitagawa (1979). Plant functional types: SHR = shrubs, HPG = high perennial grasses, SPG = short perennial graminaceous plants (grasses and sedge), ANG = annual grasses, ANF = annual forbs, PEF = perennial forbs, HAL = halophytes, HYD = hydrophytes. Habitat types: DB = disturbed and cultivated land, DU = dune, LS = light salinity, MS = moderate salinity, HS = heavy salinity, SM = saline marsh.

	Species	$\text{C}_3 / \text{C}_4$	PTF type	Habitat
<i>Gymnospermae</i>				
<i>Ephedraceae</i>	<i>Ephedra sinica</i> Staph.	$\text{C}_3$	SHR	DU
<i>Angiospermae</i>				
<i>Dicotyledoneae</i>				
<i>Amaranthaceae</i>	<i>Amaranthus ascendens</i> Lois. <i>A. blitoides</i> S. Watson <i>A. retroflexus</i> L.	$\text{C}_4$ $\text{C}_4$ $\text{C}_4$	ANF ANF ANF	DB DB DB
<i>Apocynaceae</i>	<i>Apocynum lancifolium</i> Russan.	$\text{C}_3$	HAL	MS HS
<i>Asclepiadaceae</i>	<i>Cynanchum amplexicaule</i> Hemsl. <i>C. chinensis</i> R. Br. <i>Pycnostelma paniculatum</i> K. Schum.	$\text{C}_3$ $\text{C}_3$ $\text{C}_3$	PEF PEF PEF	LS DU DU
<i>Bignoniaceae</i>	<i>Incarvillea sinensis</i> Lamark.	$\text{C}_3$	ANF	LS DU
<i>Boraginaceae</i>	<i>Cynoglossum divaricatum</i> Steph. <i>Lappula echinata</i> Gillb. <i>L. intermedia</i> (Ledeb.) M. Pop.	$\text{C}_3$ $\text{C}_3$ $\text{C}_3$	PEF PEF PEF	DU DU LS DU
<i>Campanulaceae</i>	<i>Adenophora stenophylla</i> Hemsley <i>Platycodon grandiflorum</i> D.C.	$\text{C}_3$ $\text{C}_3$	PEF PEF	DU LS DU
<i>Caryophyllaceae</i>	<i>Dianthus amurensis</i> Jacq. <i>Silene repens</i> Pall.	$\text{C}_4$ $\text{C}_3$	PEF PEF	DU LS DU
<i>Chenopodiaceae</i>	<i>Agriophyllum arenarium</i> Bieb. <i>Axyris amaranthoides</i> L. <i>Bassia dasypylla</i> O. Kuntze <i>Chenopodium acuminatum</i> Willd.	$\text{C}_4$ $\text{C}_3$ $\text{C}_4$ $\text{C}_3$	ANF ANF ANF ANF	DU DB DU DB

Table 1 (continued)

	Species	C <sub>3</sub> / C <sub>4</sub>	PTF type	Habitat
<i>Chenopodiaceae</i> (cont.)	<i>Chenopodium album</i> L.	C <sub>3</sub>	ANF	DB
	<i>Ch. glaucum</i> L.	C <sub>3</sub>	HAL	DB HS
	<i>Kochia prostrata</i> Schrad.	C <sub>4</sub>	SHR	DB
	<i>K. scoparia</i> (L.) Schrad.	C <sub>4</sub>	ANF	DB
	<i>K. sieversiana</i> (Pall.) C.A. Mey.	C <sub>4</sub>	HAL	MS SM
	<i>Salsola collina</i> Pall.	C <sub>4</sub>	ANF	DB DU
	<i>S. foliosa</i> (L.) Schrad.	C <sub>4</sub>	ANF	DB
	<i>Suaeda corniculata</i> (C.A. Mey.) Bunge	C <sub>3</sub>	HAL	HS SM
	<i>S. glauca</i> Bunge	C <sub>3</sub>	HAL	HS SM
	<i>S. heteroptera</i> Kitag.	C <sub>3</sub>	HAL	HS SM
<i>Compositae</i>	<i>Artemisia anethoides</i> Mattf.	C <sub>3</sub>	ANF	DB LS
	<i>A. anethifolia</i> Weber	C <sub>3</sub>	HAL	HS
	<i>A. annua</i> L.	C <sub>3</sub>	ANF	DU LS
	<i>A. argyi</i> Lev. et Vant.	C <sub>3</sub>	PEF	DU LS
	<i>A. desertorum</i> Spreng.	C <sub>3</sub>	PEF	DU
	<i>A. frigida</i> Willd.	C <sub>3</sub>	SHR	LS MS
	<i>A. integrifolia</i> L.	C <sub>3</sub>	PEF	LS
	<i>A. japonica</i> Thunb. var. <i>manshurica</i> Kom.	C <sub>3</sub>	PEF	DU LS
	<i>A. laciniata</i> Willd.	C <sub>3</sub>	PEF	LS
	<i>A. mongolica</i> Fisch.	C <sub>3</sub>	PEF	LS MS
	<i>A. scoparia</i> Waldst. et Kit.	C <sub>3</sub>	HAL	DB LS MS
	<i>A. sieversiana</i> Willd.	C <sub>3</sub>	PEF	DB
	<i>Filifolium sibiricum</i> Kitam.	C <sub>3</sub>	PEF	DU
	<i>Heteropappus altaicus</i> (Willd.) Novopokr.	C <sub>3</sub>	HAL	LS MS DU
	<i>Hypochaeris grandiflora</i> Ledeb.	C <sub>3</sub>	PEF	LS
	<i>Inula britanica</i> L.	C <sub>3</sub>	PEF	LS DB
	<i>I. japonica</i> Thunb.	C <sub>3</sub>	PEF	LS
	<i>Ixeris chinensis</i> Nakai subsp. <i>graminifolia</i> Kitag.	C <sub>3</sub>	PEF	DB
	<i>I. chinensis</i> Nalai var. <i>intermedia</i> Kitag.	C <sub>3</sub>	PEF	DB
	<i>I. sonchifolia</i> Hance	C <sub>3</sub>	PEF	DB
	<i>Lactuca indica</i> L.	C <sub>3</sub>	PEF	LS
	<i>Leontopodium leontopodioides</i> Beauv.	C <sub>3</sub>	PEF	DU LS
	<i>Ligularia mongolica</i> (Turcz.) DC.	C <sub>3</sub>	PEF	MS LS
	<i>Neopallasia pectinata</i> (Pall.) Poljak	C <sub>3</sub>	ANF	LS DU
	<i>Picris japonica</i> Thunb.	C <sub>3</sub>	PEF	LS DB
	<i>Saussurea glomerata</i> Poir.	C <sub>3</sub>	HAL	MS HS
	<i>S. runcinata</i> D.C.	C <sub>3</sub>	HAL	MS HS
	<i>Scorzonera glabra</i> Rupr.	C <sub>3</sub>	PEF	LS
	<i>Senecio ambracens</i> Turcz.	C <sub>3</sub>	PEF	LS MS
	<i>S. integrifolius</i> Claiville.	C <sub>3</sub>	PEF	LS
	<i>Serratula yamatsutana</i> Kitag.	C <sub>3</sub>	PEF	DU LS MS
	<i>Taraxacum mongolicum</i> Hand.	C <sub>3</sub>	HAL	MS HS
	<i>T. ohwianum</i> Kitam.	C <sub>3</sub>	HAL	MS HS
	<i>Xanthium strumarium</i> L.	C <sub>3</sub>	ANF	DB LS
<i>Convolvulaceae</i>	<i>Calystegia pellita</i> Ledeb.	C <sub>3</sub>	PEF	LS
<i>Crassulaceae</i>	<i>Sedum aizoon</i> L.	CAM	PEF	DU
<i>Cruciferae</i>	<i>Dontostemon micranthus</i> C.A. M.	C <sub>3</sub>	ANF	DU LS
	<i>Lepidium apetalum</i> Willd.	C <sub>3</sub>	ANF	DB LS
	<i>Ptilotrichum elongatum</i> C.A. May	C <sub>3</sub>	SHR	DU
<i>Dipsacaceae</i>	<i>Scabiosa comosa</i> L.	C <sub>3</sub>	PEF	DU
<i>Euphorbiaceae</i>	<i>Euphorbia esula</i> L.	C <sub>3</sub>	PEF	DU
	<i>E. humifusa</i> Willd.	C <sub>4</sub>	ANF	DB
<i>Fabaceae</i>	<i>Astragalus melilotoides</i> Pall.	C <sub>3</sub>	PEF	DU
	<i>Gueldensfaedtia stenophylla</i> Bunge	C <sub>3</sub>	PEF	LS
	<i>Kummerowia stipulacea</i> (Maxim.) Makino	C <sub>3</sub>	ANF	DB
	<i>K. striata</i> (Thunb.) Schindler	C <sub>3</sub>	ANF	DB

Table 1 (continued)

	Species	C <sub>3</sub> / C <sub>4</sub>	PTF type	Habitat
<i>Fabaceae</i> (cont.)	<i>Lespedeza davurica</i> Schindler	C <sub>3</sub>	SHR	DU
	<i>Melilotus suaveolens</i> Ledeb.	C <sub>3</sub>	ANF	DB
	<i>Oxytropis hirta</i> Bunge	C <sub>3</sub>	PEF	DU
	<i>O. myriophylla</i> D.C.	C <sub>3</sub>	PEF	DU
	<i>Thermopsis lanceolata</i> R. Br.	C <sub>3</sub>	PEF	LS
<i>Geraniaceae</i>	<i>Erodium stephanianum</i> Willd.	C <sub>3</sub>	ANF	DU
<i>Labiate</i>	<i>Phlomis tuberosa</i> L.	C <sub>3</sub>	PEF	LS MS
	<i>Schizonepeta multifida</i> (L.) Briq.	C <sub>3</sub>	ANF	DU LS MS
	<i>Scutellaria baicalensis</i> Georgi	C <sub>3</sub>	PEF	LS
	<i>S. ikonnikovii</i> Juz.	C <sub>3</sub>	PEF	LS
	<i>S. scordifolia</i> Fisch.	C <sub>3</sub>	PEF	LS
	<i>Thymus serpyllum</i> L.	C <sub>3</sub>	SHR	DU
<i>Linaceae</i>	<i>Linum stellatum</i> Planchon	C <sub>3</sub>	ANF	DU
<i>Orobanchaceae</i>	<i>Orobanche caerulescens</i> Stephan	C <sub>3</sub>	ANF	DU
<i>Plantaginaceae</i>	<i>Plantago asiatica</i> L.	C <sub>3</sub>	PEF	MS LS
	<i>P. depressa</i> Willd.	C <sub>3</sub>	PEF	MS LS
<i>Plumbaginaceae</i>	<i>Limonium bicolor</i> O. Kuntze	C <sub>3</sub>	HAL	HS SM
<i>Polygalaceae</i>	<i>Polygala tenuifolia</i> Willd.	C <sub>3</sub>	PEF	LS
<i>Polygonaceae</i>	<i>Polygonum aviculare</i> L.	C <sub>3</sub>	ANF	DB
	<i>P. divaricatum</i> L.	C <sub>3</sub>	PEF	DB DU
	<i>P. sibiricum</i> Laxm.	C <sub>3</sub>	HAL	HS SM
<i>Portulacaceae</i>	<i>Portulaca oleracea</i> L.	C <sub>4</sub>	ANF	DB
<i>Primulaceae</i>	<i>Glaux maritima</i> L.	C <sub>3</sub>	HAL	LS HS
<i>Ranunculaceae</i>	<i>Clematis hexapetala</i> Pall.	C <sub>3</sub>	PEF	DU LS
	<i>Pulsatilla chinensis</i> (Bunge) Rgl.	C <sub>3</sub>	PEF	DU LS
	<i>Ranunculus japonicus</i> Thumb.	C <sub>3</sub>	PEF	LS
	<i>Thalictrum petaloedeum</i> L. var. <i>supr.</i> (Nakai) Kitag.	C <sub>3</sub>	PEF	LS DU
	<i>Th. simplex</i> L.	C <sub>3</sub>	PEF	LS
	<i>Th. squarrosum</i> Steph.	C <sub>3</sub>	PEF	LS
<i>Rosaceae</i>	<i>Potentilla anserina</i> L.	C <sub>3</sub>	HAL	LS HS
	<i>P. betonicaefolia</i> Poiret	C <sub>3</sub>	PEF	DU
	<i>P. chinensis</i> Seringe	C <sub>3</sub>	HAL	LS MS
	<i>P. discolor</i> Bunge	C <sub>3</sub>	PEF	LS
	<i>P. Filipendula</i> Willd.	C <sub>3</sub>	PEF	LS MS
	<i>P. flagellaris</i> Willd.	C <sub>3</sub>	PEF	LS MS
	<i>P. verticillaris</i> Steph.	C <sub>3</sub>	PEF	DU LS
	<i>Sanguisorba officinalis</i> L.	C <sub>3</sub>	PEF	LS
	<i>S. tenuifolia</i> Fisch	C <sub>3</sub>	PEF	LS
<i>Rubiaceae</i>	<i>Galium verum</i> L.	C <sub>3</sub>	PEF	LS
<i>Rutaceae</i>	<i>Haplophyllum dauricum</i> Juss	C <sub>3</sub>	PEF	DU
<i>Scrophulariaceae</i>	<i>Cymbalaria dahurica</i> L.	C <sub>3</sub>	PEF	DU LS
	<i>Veronica linearifolia</i> Pall.	C <sub>3</sub>	PEF	LS
<i>Solanaceae</i>	<i>Solanum nigrum</i> L.	C <sub>3</sub>	ANF	DB
<i>Thymelaeaceae</i>	<i>Diarthron linifolium</i> Turcz.	C <sub>3</sub>	ANF	DU
	<i>Stellera chamajiasme</i> L.	C <sub>3</sub>	PEF	DU
<i>Umbelliferae</i>	<i>Bupleurum scorzonerifolium</i> Willd.	C <sub>3</sub>	PEF	LS
	<i>Sphallerocarpus gracilis</i> (Bess.) K.	C <sub>3</sub>	PEF	DB
	<i>Siler divaricatum</i> Benth. et Hook	C <sub>3</sub>	PEF	LS
<i>Violaceae</i>	<i>Viola dissecta</i> Ledeb.	C <sub>3</sub>	PEF	DU
<i>Zygophyllaceae</i>	<i>Tribulus terrestris</i> L.	C <sub>4</sub>	ANF	LS

Table 1 (continued)

	Species	C <sub>3</sub> / C <sub>4</sub>	PTF type	Habitat
<i>Monocotyledoneae</i>				
<i>Commelinaceae</i>	<i>Commelina communis</i> L.	C <sub>4</sub>	PEF	DB
<i>Cyperaceae</i>	<i>Bolboschoenus compactus</i> Heffm.	C <sub>3</sub>	HYD	SM
	<i>B. maritimus</i> (L.) Pall.	C <sub>3</sub>	HYD	SM
	<i>Carex duriuscula</i> C.A.M.	C <sub>3</sub>	HAL	HS MS
	<i>C. pediformis</i> C.A.M.B.	C <sub>3</sub>	SPG	LS DU
	<i>Heleocharis intersita</i> Zinesrl.	C <sub>3</sub>	HYD	SM
	<i>Scirpus tabernaemontani</i> Gmel.	C <sub>3</sub>	HYD	SM
<i>Gramineae</i>	<i>Achnatherum avinoides</i> (Honda) Chang	C <sub>4</sub>	HPG	LS MS
	<i>A. sibiricum</i> (L.) Keng	C <sub>4</sub>	HAL	HS SM
	<i>Aeluropus littoralis</i> (Gouan) Parlat	C <sub>4</sub>	HAL	HS SM
	<i>Agropyron cristatum</i> (L.) Gaertner	C <sub>3</sub>	HPG	DU LS
	<i>Arthraxon hispidus</i> (Thunb.) Makino	C <sub>4</sub>	HPG	DB
	<i>Arundinella hirta</i> (Thunb.) Tanaka	C <sub>4</sub>	HPG	LS
	<i>Avena fatua</i> L.	C <sub>3</sub>	HPG	DB
	<i>Calamagrostis epigeios</i> (L.) Roth	C <sub>3</sub>	HPG	LS SM
	<i>Chloris virgata</i> Sw.	C <sub>4</sub>	HAL	HS MS SM
	<i>Cleistogenes chinensis</i> (Maxim.) Keng	C <sub>4</sub>	SPG	LS
	<i>C. squarrosa</i> (Trin.) Keng	C <sub>4</sub>	SPG	LS
	<i>Clinelymus sibiricus</i> (L.) Nevski	C <sub>3</sub>	HPG	LS MS
	<i>Digitaria ciliaris</i> (Rotz.) Koel	C <sub>4</sub>	ANG	DB
	<i>D. ischaemum</i> (Schreb.) Schreb. ex Muhl.	C <sub>4</sub>	ANG	DB
	<i>D. lineis</i> (Krock.) Crep	C <sub>4</sub>	ANG	DB
	<i>D. sanguinalis</i> (L.) Scop.	C <sub>4</sub>	ANG	DB
	<i>Echinochloa caudata</i> Roshev.	C <sub>4</sub>	ANG	SM HS
	<i>E. crus galli</i> (L.) Beauv.	C <sub>4</sub>	ANG	SM HS
	<i>Eragrostis cilianensis</i> (All.) Link.	C <sub>4</sub>	HPG	DB
	<i>E. filiformis</i> Link.	C <sub>4</sub>	HPG	DB
	<i>E. pilosa</i> (L.) P. B.	C <sub>4</sub>	HPG	DB
	<i>E. poaeoides</i> Beauv.	C <sub>4</sub>	HPG	DB LS
	<i>Eriochloa villosa</i> (Thunb.) Kunth L.	C <sub>4</sub>	HPG	DU LS
	<i>Festuca ovina</i> L.	C <sub>3</sub>	HPG	LS
	<i>Hemarthria sibirica</i> (Gand.) Ohwi	C <sub>4</sub>	HYD	SM
	<i>Hierochloe glabra</i> Trin.	C <sub>3</sub>	HPG	LS DB
	<i>Hordeum brevisubulatum</i> (Trin.) Link	C <sub>3</sub>	HAL	HS SM
	<i>H. brevisubulatum</i> var. <i>hirtellum</i> Cheng ex Skv.	C <sub>3</sub>	HPG	HS SM
	<i>Imperata cylindrica</i> (L.) P.B.	C <sub>4</sub>	HPG	DU
	<i>Koeleria cristata</i> (L.) Pers.	C <sub>3</sub>	HPG	LS
	<i>Leymus chinensis</i> (Trin.) Tzvel.	C <sub>3</sub>	HPG	LS MS
	<i>Miscanthus sacchariflorus</i> (Maxim.) Hack.	C <sub>4</sub>	HPG	LS
	<i>Panicum ruderale</i> (Kitag.) Cheng	C <sub>4</sub>	SPG	DB
	<i>Pennisetum flaetidum</i> Griseb.	C <sub>4</sub>	HPG	DU
	<i>Phragmites communis</i> Trin.	C <sub>3</sub>	HYD	HS SM
	<i>Poa pratensis</i> L.	C <sub>3</sub>	SPG	LS
	<i>Puccinellia chinampoensis</i> Ohwi.	C <sub>3</sub>	HAL	HS SM
	<i>P. jeholensis</i> Kitag.	C <sub>3</sub>	HAL	HS SM
	<i>P. tenuiflora</i> (Turcz.) Scrib. et Merr.	C <sub>3</sub>	HAL	HS SM
	<i>Setaria lutescens</i> (Weigel) F. T. Hubb.	C <sub>4</sub>	ANG	DB
	<i>S. viridis</i> (L.) Beauv.	C <sub>4</sub>	ANG	DB
	<i>Spodiopogon sibiricus</i> Trin.	C <sub>4</sub>	HPG	DU
	<i>Stipa baicalensis</i> Roshev.	C <sub>3</sub>	HPG	DU LS
	<i>S. grandis</i> P. Smirn.	C <sub>3</sub>	HPG	LS
<i>Liliaceae</i>	<i>Allium condensatum</i> Turcz.	C <sub>3</sub>	PEF	DU
	<i>A. macrostemon</i> Bunge	C <sub>3</sub>	PEF	DB LS
	<i>A. polyyrrhizum</i> Turcz.	C <sub>3</sub>	HAL	HS
	<i>A. senescens</i> L.	C <sub>3</sub>	PEF	MS HS
	<i>A. tenuissimum</i> L.	C <sub>3</sub>	PEF	DU

Table 1 (continued)

	Species	C <sub>3</sub> / C <sub>4</sub>	PTF type	Habitat
Liliaceae (cont.)	<i>Anemarrhena asphodeloides</i> Bunge.	C <sub>3</sub>	PEF	DU
	<i>Asparagus davarica</i> Fisch.	C <sub>3</sub>	PEF	DU LS
	<i>Lilium tenuifolium</i> Fisch.	C <sub>3</sub>	PEF	LS
	<i>Scilla thunbergii</i> Miyabe et Kudo	C <sub>3</sub>	PEF	LS
Iridaceae	<i>Iris dichotoma</i> Pall.	C <sub>3</sub>	PEF	DU LS
	<i>I. pallasii</i> Fisch.	C <sub>3</sub>	HAL	HS MS
	<i>I. ruthenica</i> Ker-Gawl.	C <sub>3</sub>	PEF	DU
	<i>I. tenuifolia</i> Pall.	C <sub>3</sub>	PEF	HS MS
	<i>I. ventricosa</i> Pall.	C <sub>3</sub>	PEF	MS LS
Typhaceae	<i>Typha minima</i> Funk	C <sub>3</sub>	HYD	SM

## Results

**Floristic composition of photosynthetic pathway types:** 191 vascular plant species (about 25 % of the total species in the Songnen grasslands), in 42 families and 124 genera, were identified with C<sub>3</sub>, C<sub>4</sub>, and CAM photosynthesis (Table 1). Of these species, 1 species was in *Gymnospermae*, 190 in *Angiospermae*. 65 % (124 of 191) was found in *Dicotyledoneae*, e.g. *Compositae* (34 species), *Chenopodiaceae* (14 species), *Fabaceae* (9 species), *Rosaceae* (8 species), and *Ranunculaceae* (6 species). 35 % were in *Monocotyledoneae*, e.g. *Gramineae* (44 species), *Liliaceae* (9 species), *Cyperaceae* (6 species), and *Iridaceae* (5 species). As for photosynthetic pathway types, 150 species (79 % of the identified species in Table 1) in 99 genera and 37 families were found with C<sub>3</sub> photosynthesis, 40 species in 25 genera and 8 families with C<sub>4</sub> photosynthesis, and 1 species with CAM photosynthesis. 21 % (40 of 191) of the identified species in Table 1, or about 6 % of identified species in local flora, was found with C<sub>4</sub> photosynthesis, e.g. *Gramineae* (27 species), *Chenopodiaceae* (5 species), *Amaranthaceae* (3 species), 1 species each in *Caryophyllaceae*, *Euphorbiaceae*, *Portulacaceae*, *Zygophyllaceae*, and *Commelinaceae*. The occurrence of C<sub>4</sub> species was common in *Gramineae* (68 %) and *Chenopodiaceae* (36 %), and all the species in *Amaranthaceae*, *Portulacaceae*, *Zygophyllaceae*, and *Commelinaceae* were found with C<sub>4</sub> photosynthesis in the region.

The occurrence of C<sub>4</sub> species and C<sub>4</sub> proportion (C<sub>4</sub>/total species) were significant response to habitats in the saline grasslands in the region (Fig. 1A). Disturbed and cultivated land (DB) had the highest number of C<sub>4</sub> species (22 species) and C<sub>4</sub> proportion (47.8 %), followed by saline marsh (7 species and 29.2 %). C<sub>4</sub> proportions increased significantly from light salinity (LS) to saline marsh (SM) or with the salinization. Some C<sub>4</sub> species, e.g. *K. sieversiana* (Pall.) C.A. Mey., *Chloris virgata* Sw., *Aeluropus littoralis* (Gouan) Parlat, and *Achnatherum avinoides* (Honda) Chang., can form large patch of consociations. This suggested that the occurrence of C<sub>4</sub> species was related with grassland salinity in the region.

**Morphological functional types:** According to plant morphological attributes and life-span, the identified plant species were classified into 8 functional types, e.g. SHR, HPG, SPG, ANG, ANF, PEF, HAL, and HYD (Table 1 and Fig. 1B). Of the total 191 species identified in Table 1, 44 % was PEF type, while this type makes up less than 10 % of the total forage production in the region. 16 % species was ANF type, followed by HAL (15 %), HPG (12 %), ANG (5 %), HYD (4 %), SHR (3 %), and SPG (3 %). The total amount of grass species, including HPG, ANG, SPG, and some species in HAL and HYD types, was about 23 %, and these grass types make up about 90 % forage production in this type of grassland. Most grasses are the dominant species, e.g. *Leymus chinensis* (Trin.) Tzvel., *Calamagrostis epigeios* (L.) Roth., *Chloris virgata* Sw., and *Cleistogenes squarrosa* (Trin.) Keng. 15 % of the total species in this type of grassland was halophytes (HAL), e.g. *Suaeda corniculata* (C.A. Mey.) Bunge, *S. glauca* Bunge, *Aeluropus littoralis* (Gouan) Parlat, *Chloris virgata* Sw., and most of these species were also dominant species in heavy salinity area, e.g. water supply, camp near the centre of the grassland, and in the vicinity of gates to grazed areas. There were few SHR functional type species in the grasslands, e.g. *Ephedra sinica* Staph., *Kochia prostrata* Schrad., *Artemisia frigida* Willd., and *Ptilotrichum elongatum* C.A. May. SHR type species were only about 3 % of the total identified in the Table 1.

The responses of morphological functional type compositions to grassland salinity were remarkable in the saline grasslands (Fig. 2). Species numbers of PEF, HPG, and ANF decreased with the saline intensity or from light salinity to saline marsh, while those for HAL, HYD, and ANG increased considerably. Relative more abundance for HAL and ANG indicated that the saline stress was higher in these habitats.

Photosynthetic pathway compositions varied considerably in these morphological functional types (Fig. 1B). C<sub>4</sub> proportion (C<sub>4</sub>/total species) was relatively higher in the grass functional types, e.g. HPG (52 %), ANG

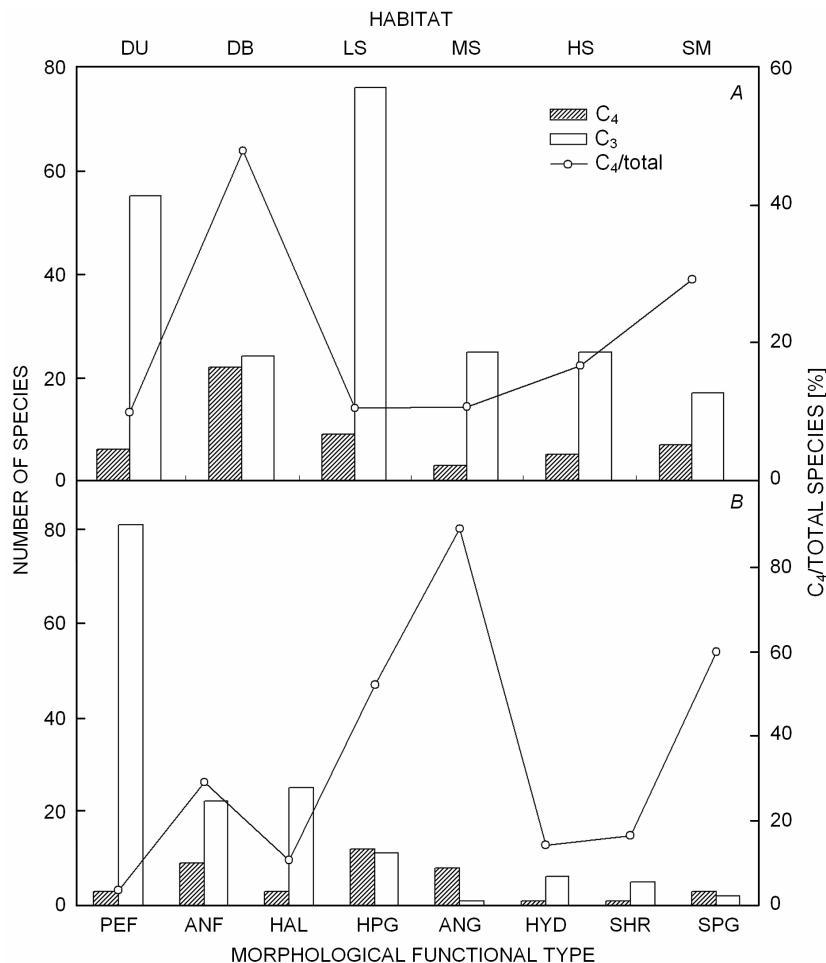


Fig. 1. The numbers of photosynthetic pathway types ( $C_3$ ,  $C_4$ , and CAM) in different habitats (A) and photosynthetic pathway compositions in morphological functional types (B) in the saline grasslands, Northeastern China. Habitats: DB = disturbed and cultivated land, DU = dune, LS = light salinity, MS = moderate salinity, HS = heavy salinity, SM = saline marsh.

(89 %), SPG (60 %), and annual forbs (ANF) type (29 %), but much lower in PEF (4 %), HAL (11 %), HYD (14 %), and SHR (17 %). Larger  $C_4$  proportion in

ANG and ANF was related with vegetation dynamics, because most annual  $C_4$  species are the pioneer species in grassland succession.

## Discussion

Plant functional types, including photosynthetic pathways and morphological functional types, bring more details for studying the logical linkage between species and ecosystems at both species and global levels (Collins and Jones 1985, Chapin 1993, Cramer 1997, Leemans 1997, Duckworth 2000). The occurrence of  $C_4$  species was not only related with large-scale climate variations, but also affected by local soil and community differences. Both the number of  $C_4$  species and  $C_4$  proportion in the saline grasslands were higher than in the other grassland types in Northeastern China, likely due to complex geo-relief, landscape, and soils (Wang 2002a). 40  $C_4$  species were identified in this type grassland, which was about 48 % and 74 % greater than those from steppe and desert border on the western part of this saline grassland. This may

be mainly due to the intense salinity soil with pH average about 8.7, rising as high as 10.0 in the spring, and sufficient precipitation (450 mm per year) in the region. The occurrence of halophyte and hydrophyte  $C_4$  species, e.g. *K. sieversiana* (Pall.) C.A. Mey., *Chloris virgata* Sw., *Aeluropus littoralis* (Gouan) Parlat, *Achnatherum avinoides* (Honda) Chang., and *Echinochloa caudata* Roshev., was very common (Table 1), because these  $C_4$  species had relative higher saline tolerance and water use efficiency (Redmann *et al.* 1995, Yin and Li 1997, Wang 2002c). Relatively more annual precipitation resulted in the higher abundance of  $C_4$  grasses (27 species), which is the leading family with  $C_4$  photosynthesis. This finding is much different from the work by Pyankov *et al.* (2000): they found that *Chenopodiaceae* was the leading  $C_4$

family, while the *Gramineae* ranks the second in desert and steppe in Mongolia (Pyankov *et al.* 2000). The other previous studies (Wang 2002a,b,c) also proved that the

abundance of  $C_4$  *Chenopodiaceae* species was relatively higher in the steppe and desert, which is much drier than the saline grasslands.

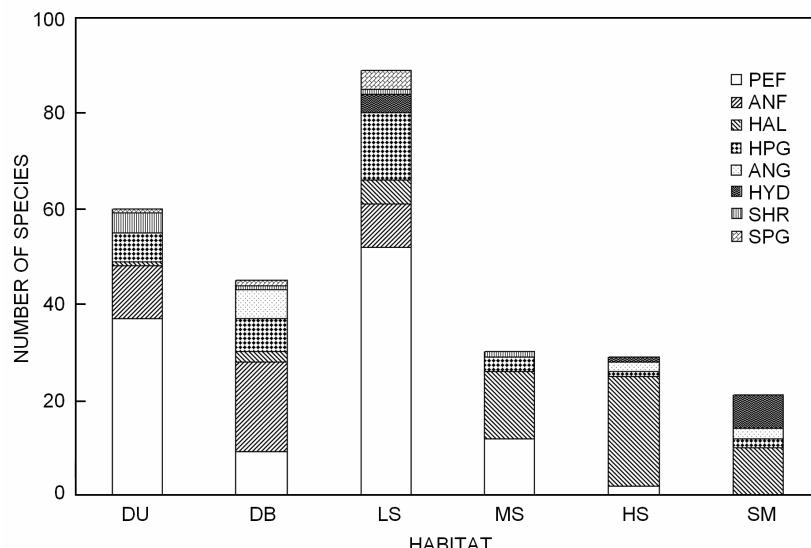


Fig. 2. The morphological functional type compositions in different habitats in the saline grasslands, Northeastern China. Habitats: DB = disturbed and cultivated land, DU = dune, LS = light salinity, MS = moderate salinity, HS = heavy salinity SM = saline marsh.

The occurrence of  $C_4$  species is consistent with grassland conditions, *e.g.* salinity, deterioration, and grazing intensities in the region (Wang 2002a,c). This is supported by the observations that  $C_4$  species number and  $C_4$  proportion increased significantly with the saline intensity or from light salinity to saline marsh (Fig. 1A). Some  $C_4$  species, *Chloris virgata* Sw., *Aeluropus littoralis* (Gouan) Parlat, and *Achnatherum avinoides* (Honda) Chang have higher salt tolerance to survive intense salinity condition and become the pioneer species in the restoration succession of salinized grassland, and can form large patch of consociations in the region. The earlier study (Wang 2002a) also proved that the  $C_4$  species had greater capacity to tolerate environment stresses (saline and drought) caused by animal grazing.

The use of morphological functional types, combined with photosynthetic pathways, has become essential to study and predict consequences of environment changes on vegetation and ecosystem processes. Morphological functional types clearly reflect spatial and temporal changes in saline grassland ecosystems in local and region scales (Fig. 1B). Comparing with the adjacent steppe, the saline grassland had more perennial forbs and hydrophyte

species, but less shrubs, short perennial graminaceous plants (grasses and sedge), and no succulents, indicating this type of grassland was more moist than steppe (Table 1, Fig. 1B; Wang 2003). Morphological functional type components varied significantly with the habitat types in the saline grasslands (Fig. 2). The number of PEF type decreased with grassland deterioration and salinity or from LS to DB, MS, HS, and SM, while that for HAL increased significantly (Fig. 2). This suggested the responses of these two functional types to the grassland salinity were different and can be used for predicting consequences of vegetation dynamics and grassland deterioration in local scale. Relatively more  $C_4$  species found in ANF, HPG, ANG, and SPG types indicate that these functional types are more fit for the saline and moist grassland in the region. This and an earlier study (Wang 2002a) clearly document the importance of plant function types (*e.g.* photosynthetic pathways and morphological functional types) in predicting vegetation dynamics in the saline grassland and the occurrence of  $C_4$  species and halophytes can be indicator plants for both diagnosis of grassland conditions and management decisions in the region.

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