

# Variable responses of mesophyll conductance to substomatal carbon dioxide concentration in common bean and soybean

J.A. BUNCE

*Crop Systems and Global Change Laboratory, USDA-ARS, Beltsville Agricultural Research Center, 10300 Baltimore Avenue, 20705-2350 Beltsville, MD, USA*

## Abstract

Some reports indicate that mesophyll conductance ( $g_m$ ) to carbon dioxide varies greatly with the substomatal carbon dioxide concentration ( $C_i$ ) during the measurement, while other reports indicate little or no change in  $g_m$  with  $C_i$ . I used the oxygen sensitivity of photosynthesis to determine the response of  $g_m$  to  $C_i$  over the range of about 100 to 300  $\mu\text{mol mol}^{-1}$   $C_i$  at constant temperature in common bean (*Phaseolus vulgaris*) and soybean (*Glycine max*) grown over a range of temperatures and photosynthetic photon flux densities (PPFD). In soybean grown and measured at high PPFD there was only a slight, approximately 15% decrease in  $g_m$  with  $C_i$  over the range of 100 to 300  $\mu\text{mol mol}^{-1}$ . With lower PPFD during the measurement of  $g_m$ , and especially with low PPFD during plant growth, there was a larger decrease in  $g_m$  with  $C_i$  in soybean. In common bean, the same range in  $C_i$  resulted in about a 60% decrease in  $g_m$  for plants grown and measured at high PPFD, with an even larger decrease for plants at low growth or measurement PPFD. Growth temperatures of 20 to 30°C had little influence on the response of  $g_m$  to  $C_i$  or its absolute value in either species. It is concluded that these two species differed substantially in the sensitivity of  $g_m$  to  $C_i$ , and that PPFD but not temperature during leaf development strongly affected the response of  $g_m$  to  $C_i$ .

*Additional key words:* *Glycine max*; light; *Phaseolus vulgaris*; temperature.

## Introduction

With the recognition that there is a significant difference in the carbon dioxide concentration between the substomatal airspace and the site of carbon dioxide fixation within chloroplasts during photosynthesis, there has been renewed interest in estimating the “mesophyll conductance” ( $g_m$ ) to the movement of carbon dioxide along this path and determining the structural, metabolic and environmental factors which affect  $g_m$  (Evans and von Caemmerer 1996, Flexas *et al.* 2008, Warren 2008a). Variation in  $g_m$  with the carbon dioxide concentration in the substomatal airspace ( $C_i$ ) would greatly complicate the biochemical modeling of photosynthesis until the basis of such variation is understood. Variation in  $g_m$  with  $C_i$  has been observed in some species using both the carbon isotope and the fluorescence methods of estimating  $g_m$  (During 2003, Flexas *et al.* 2007a, Hassiotou *et al.* 2009, Vrabl *et al.* 2009), but did not occur in other studies (Loreto *et al.* 1992, Bunce 2008, 2009, Tazoe *et al.* 2009). Evans (2009) suggested that errors in the fluorescence methods could create the

appearance of a change in  $g_m$  with  $C_i$ , although fluorescence methods have been implemented somewhat differently in various studies. It remains unclear whether the different responses of  $g_m$  to  $C_i$  result from species differences or from differences in measurement techniques.

The method of using the oxygen sensitivity of photosynthesis to estimate  $g_m$  (Bunce 2009) is well suited to determining the  $C_i$  response of  $g_m$  over the lower range of  $C_i$ , where photosynthesis is limited by carboxylation capacity. Because  $\text{O}_2$  and  $\text{CO}_2$  compete for RuBp at Rubisco, and this competition determines the rate of net photosynthesis as long as neither substrate is saturating, the sensitivity of  $\text{CO}_2$ -limited photosynthesis to a change in  $[\text{O}_2]$  provides information on the  $[\text{CO}_2]$  at Rubisco ( $C_c$ ). The main assumptions of the method are that competition at Rubisco described by the Farquhar-type  $C_3$  photosynthesis model fully explains  $[\text{O}_2]$  effects on  $\text{CO}_2$  fixation and that respiration in the light is unchanged over the  $[\text{O}_2]$  range used. The resolution of the method is greatest when photosynthesis is limited by carboxylation

Received 29 April 2010, accepted 6 August 2010.

Phone: 301-504-7629, fax: 301-504-5823, e-mail: James.Bunce@ars.usda.gov

*Abbreviations:*  $C_c$  –  $[\text{CO}_2]$  at Rubisco;  $C_i$  –  $[\text{CO}_2]$  in the substomatal (intercellular) airspace;  $g_m$  – mesophyll conductance to  $\text{CO}_2$ ;  $P_N$  – net photosynthetic rate; PPFD – photosynthetic photon flux density; Rubisco – ribulose-1,5-bisphosphate carboxylase/oxygenase; RuBp – ribulose bisphosphate;  $V_{\text{Cmax}}$  – the maximum rate of carboxylation of Rubisco.

capacity rather than electron transport (Bunce 2009).

In some surveys of species (Evans and von Caemmerer 1996, Hassiotou *et al.* 2009),  $g_m$  was found to scale approximately with maximum  $P_N$ , but there is less information on whether  $g_m$  scales with maximum  $P_N$  within a species grown under conditions which produce a range of maximum  $P_N$ . A few studies have measured the response of  $g_m$  to temperature, and found the

responses to vary among species (reviewed in Bunce 2008, Warren 2008b). However, there is little information on the response of  $g_m$  to measurement PPFD. In this study, I used the oxygen sensitivity of  $P_N$  to estimate  $g_m$  in soybean and common bean over a range of  $C_i$  values, for plants grown over a range of temperatures and PPFD levels and measured at different PPFD levels at constant temperature.

## Materials and methods

Soybeans (*Glycine max* L. Merr., cv. Kent) and common beans (*Phaseolus vulgaris* L., cv. Dark Red Kidney) were grown in controlled environment chambers. Plants were grown singly in 20 cm diameter pots filled with vermiculite and flushed daily with a complete nutrient solution containing 14.5 mM nitrogen. Plants were grown at 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  photosynthetic photon flux density (PPFD) for 12 h per day, with constant air temperatures of 20, 25, or 30°C. Plants were also grown at 25°C with a PPFD of 150  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . In all growth conditions, the  $[\text{CO}_2]$  was controlled at 370  $\mu\text{mol mol}^{-1}$ , and the relative humidity was 60%. Light was supplied by a mixture of high-pressure sodium and metal halide lamps. Leaf gas exchange measurements were made on second trifoliolate leaves a few days after they reached maximum area.

The response of  $g_m$  to  $C_i$  over the range of 100 to 300  $\mu\text{mol mol}^{-1}$  was measured at 25°C, 1500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD and a leaf to air vapor pressure deficit of 1.2 to 1.4 kPa for three or four leaves of both species grown under all of the environmental conditions given earlier. Leaf gas-exchange measurements were made using a LI-6400 portable photosynthesis system (LI-Cor, Inc., Lincoln, Nebraska). Steady-state rates of  $P_N$  in both 2% and 21%  $[\text{O}_2]$  were recorded at six different external  $[\text{CO}_2]$ , ranging from about 100 to 800  $\mu\text{mol mol}^{-1}$ . The system software was used to correct the output of the infrared analyzers for the background  $[\text{O}_2]$  and to calculate  $C_i$ . Gas-exchange measurements were conducted inside a controlled environment chamber in which the water vapor pressure was controlled to match

( $\pm 0.1$  kPa) that inside the cuvette. This was found to eliminate the need to correct for water vapor leakage into or out of the cuvette (Rodeghiero *et al.* 2007). Corrections for  $\text{CO}_2$  leakage through cuvette materials were made based on the difference between the chamber and cuvette  $[\text{CO}_2]$ , using the manufacturer's protocol. Possible leakage of  $\text{CO}_2$  between the leaf surface and the gasket was tested by increasing the  $[\text{CO}_2]$  around the cuvette to several thousand  $\mu\text{mol mol}^{-1}$  and observing changes in  $\text{CO}_2$ -exchange rate. No significant leaks were detected.

A Farquhar-type  $C_3$  photosynthesis model with updated kinetic parameters (Sharkey *et al.* 2007) was used to estimate  $g_m$  from  $P_N$  and  $C_i$  at 2 and 21%  $[\text{O}_2]$ , using the method in Bunce (2009). This was done from composite  $P_N$  vs.  $C_i$  curves from three or four leaves for each species and growth condition by determining by trial and error values for  $V_{\text{Cmax}}$  and  $g_m$  which fit the observed rates of  $P_N$  at both 21 and 2%  $[\text{O}_2]$  interpolated at  $C_i$  values of 100 to 300  $\mu\text{mol mol}^{-1}$ , with steps of 50  $\mu\text{mol mol}^{-1}$ . Estimates of  $g_m$  at higher  $C_i$  were often not feasible because  $P_N$  at 2%  $[\text{O}_2]$  became insensitive to increases in  $C_i$  above 300  $\mu\text{mol mol}^{-1}$ . The composite  $P_N$  vs.  $C_i$  curves consisted of measurements on four leaves per species and growth condition. Interpolated values of  $P_N$  at the steps in  $C_i$  was determined by fitting curves for an exponential rise to a maximum of the form  $P_N = P_{N0} + a(1 - e^{-bC_i})$ , using the nonlinear regression utility of *SigmaPlot v. 11* software. Statistical tests were conducted using *JMP v. 5* (SAS Institute, Cary, North Carolina).

## Results

In soybean grown and measured at high PPFD,  $g_m$  decreased only slightly with increases in  $C_i$  (Fig. 1). Values at 300 averaged about 85% of those at 100  $\mu\text{mol mol}^{-1}$ . Growth temperatures from 20 to 30°C had little effect on the magnitude of  $g_m$ , or its response to  $C_i$ . Low PPFD during the measurement of  $g_m$  did not affect the  $g_m$  measured at low  $C_i$ , but reduced it by about 25% at the higher  $C_i$  values (Fig. 1). At low measurement PPFD,  $g_m$  decreased relatively more with  $C_i$  than at high PPFD, and the value of  $g_m$  at 300  $\mu\text{mol mol}^{-1}$  was about 65% of that at 100  $\mu\text{mol mol}^{-1}$   $C_i$ . Low PPFD during plant growth

resulted in lower  $g_m$  values over the whole range of  $C_i$  (Fig. 1), and increased the relative reduction in  $g_m$  with  $C_i$ .  $P_N$  at a  $C_i$  of 250  $\mu\text{mol mol}^{-1}$  decreased slightly with growth temperature and was reduced by low growth or measurement PPFD (Table 1).

The decrease in  $g_m$  with  $C_i$  in beans grown and measured at high PPFD was larger than in soybean (Fig. 2), with values at 300  $\mu\text{mol mol}^{-1}$   $C_i$  about 40% of those at 100  $\mu\text{mol mol}^{-1}$  for all three growth temperatures. Even larger decreases in  $g_m$  with  $C_i$  occurred in bean leaves either grown or measured at lower PPFD (Fig. 2).  $P_N$

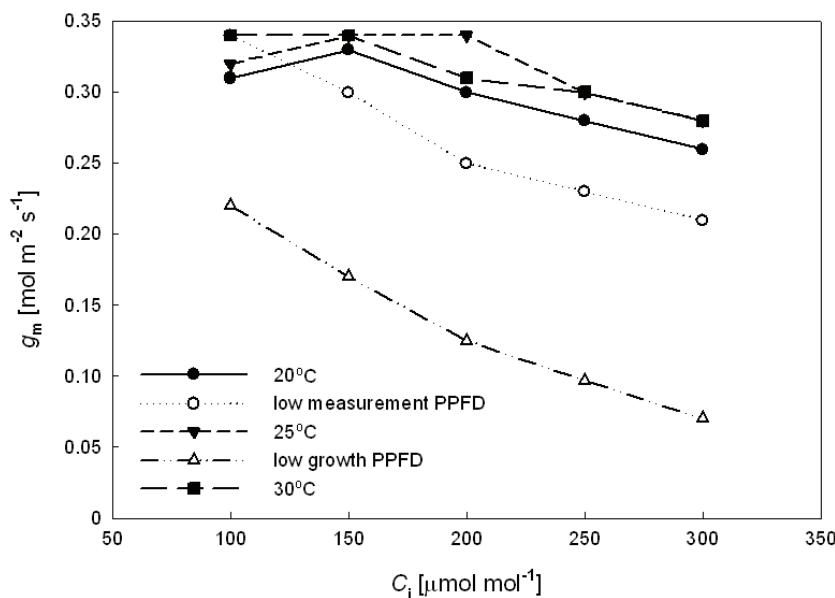


Fig. 1. Responses of  $g_m$  to  $C_i$  in soybean leaves measured at 25°C and 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD for plants grown at 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD at 20, 25, and 30°C, and for plants grown at 25°C at 150  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , measured at 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD, and for plants grown at 25°C and 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD but measured at 300  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD. Estimates of  $g_m$  are based on gas-exchange measurements on three or four leaves per species and growth condition.

Table 1. Rates of  $P_N$  at 250  $\mu\text{mol mol}^{-1}$   $C_i$ , 25°C in bean and soybean leaves grown at a range of temperatures and PPFDs and measured at either 1,500 or 300  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD.  $P_N$  values are means of three or four leaves.

Growth temperature [°C]	Growth PPFD [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ]	Measurement PPFD [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ]	$P_N$ [ $\mu\text{mol m}^{-2} \text{s}^{-1}$ ]	Bean	Soybean
20	1,000	1,500	24.8	26.3	
25	1,000	1,500	25.7	25.1	
30	1,000	1,500	24.0	23.8	
25	1,000	300	11.4	12.3	
25	150	1,500	15.8	15.2	

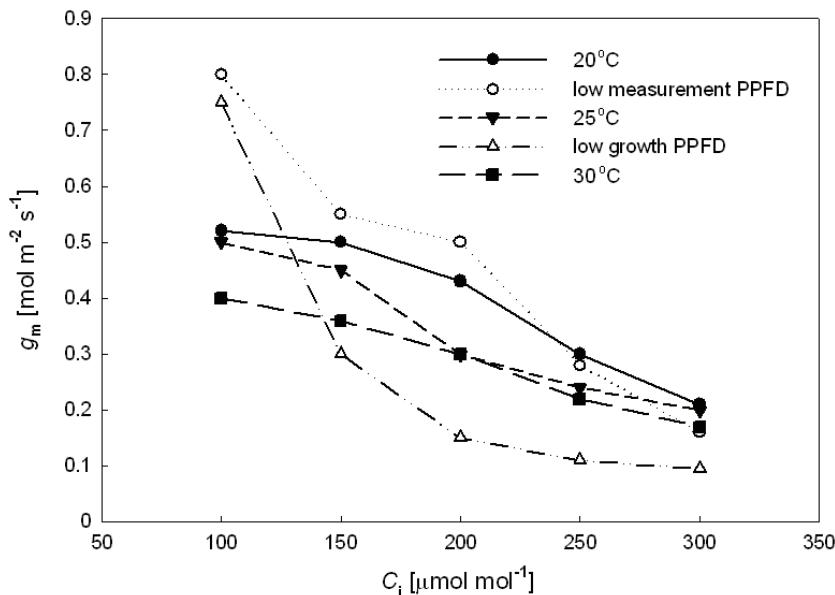


Fig. 2. Responses of  $g_m$  to  $C_i$  in common bean leaves measured at 25°C and 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD for plants grown at 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD at 20, 25, and 30°C, and for plants grown at 25°C at 150  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , measured at 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD, and for plants grown at 25°C and 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD but measured at 300  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD. Estimates of  $g_m$  are based on gas-exchange measurements on three or four leaves per species and growth condition.

at a  $C_i$  of 250  $\mu\text{mol mol}^{-1}$  in bean were similar to soybean for the same growth and measurement conditions (Table 1).

In both species the ratio of the CO<sub>2</sub> level at Rubisco compared to the substomatal air space ( $C_c$  to  $C_i$ ) decreased with  $C_i$  for plants grown and measured at high

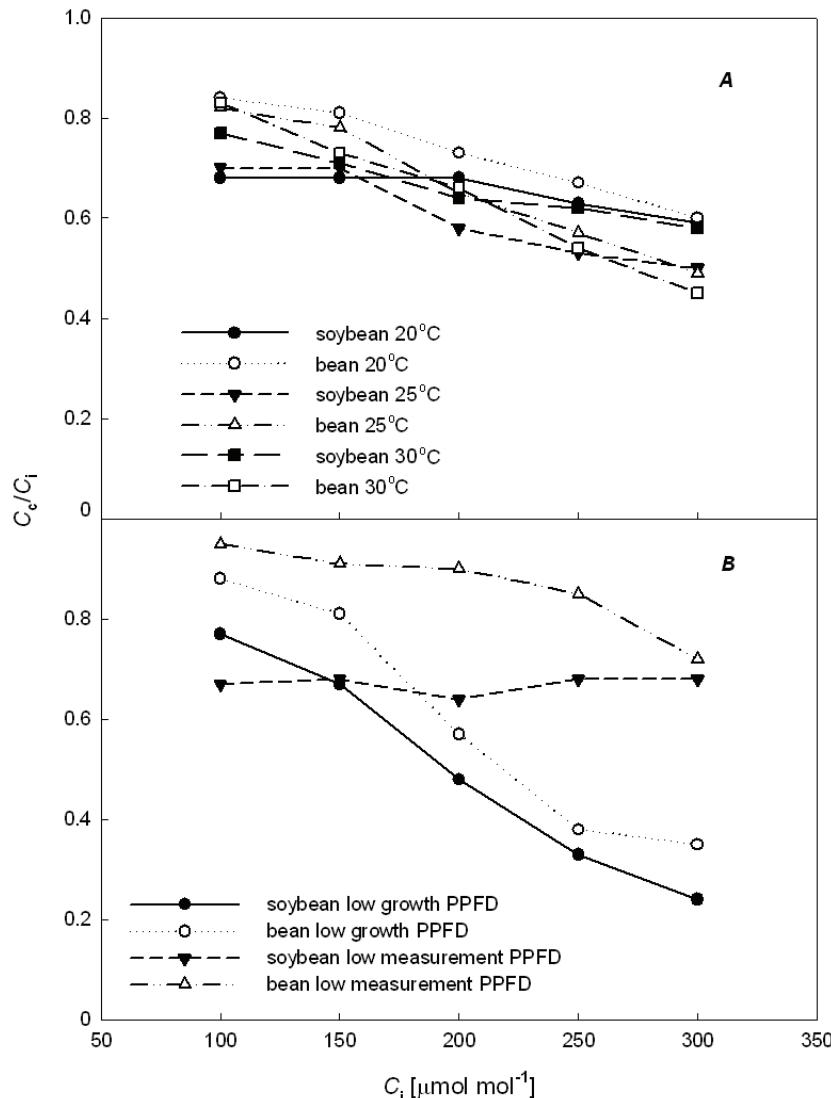


Fig. 3. The ratio of  $C_c$  to  $C_i$  at different  $C_i$  values for soybean and common bean *A*: grown at 20, 25, or 30°C at 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD and measured at 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD and *B*: for plants grown at 25°C and 150  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD, measured at 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD or grown at 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD but measured at 300  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD. Estimates of the ratio are based on gas-exchange measurements on three or four leaves per species and growth condition.

PPFD (Fig. 3*A*). On average the decrease in the  $C_c$  to  $C_i$  ratio was larger in bean than in soybean (Table 2), reflecting the larger decrease in  $g_m$  with  $C_i$  in bean. In bean, low measurement PPFD raised the  $C_c$  to  $C_i$  ratio

at all  $C_i$  values (Fig. 3*B*). In both bean and soybean grown at low PPFD, the  $C_c$  to  $C_i$  ratio decreased much more strongly with increasing  $C_i$  than for plants grown at higher PPFD (Fig. 3*B*).

## Discussion

These results indicate that bean and soybean, despite having similar leaf  $P_N$  under the same conditions, had contrasting responses of  $g_m$  to  $C_i$  when grown and measured at high PPFD, with very little change in  $g_m$  with  $C_i$  in soybean, but a large decrease in  $g_m$  with  $C_i$  in bean. The estimates of  $g_m$  in both cases used the same methodology and instrumentation, thus the differences in response were not caused by different measurement techniques in this case. The range of  $C_i$  over which  $g_m$  was determined covered the normal operating range for the current ambient atmospheric carbon dioxide concentration, including values typical of plants with low stomatal conductance because of water stress.

While several studies have tested for effects of water stress on  $g_m$ , effects of growth temperature and growth PPFD have been less studied. The small effect of growth temperature on  $g_m$  observed here in both species agrees with the few prior studies which have tested this (Yamori *et al.* 2005, Warren and Dreyer 2006, Bunce 2008, Warren 2008b), although minor changes were observed in some cases. Our results additionally indicate that a range of growth temperatures did not substantially affect the response of  $g_m$  to  $C_i$ , whether  $g_m$  was highly sensitive to  $C_i$ , as in bean or relatively insensitive, as in soybean.

The lower  $g_m$  in leaves developed at low PPFD observed here in both species is consistent with the lower

Table 2. Linear regression equations for the responses of the ratio  $C_c/C_i$  to  $C_i$  over the range of 100 to 300  $\mu\text{mol mol}^{-1}$   $C_i$  in leaves of bean and soybeans grown at three temperatures, measured at 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD.

Species	Growth temperature [°C]	Intercept	Slope	$r^2$
Bean	20	0.98	-1,200	0.99
	25	1.01	-1,700	0.98
	30	1.02	-1,900	0.99
Soybean	20	0.74	-460	0.79
	25	0.83	-1,100	0.90
	30	0.85	-940	0.96

$g_m$  in *Arabidopsis thaliana* grown at low PPFD (Flexas *et al.* 2007b) compared with the same species grown at high PPFD (Bunce 2008), and with the comparison of sun- and shade leaves of *Fagus sylvatica* (Warren *et al.* 2007). At least as importantly, low growth PPFD greatly increased the relative decrease in  $g_m$  with  $C_i$  in both species examined here, which makes comparisons of  $g_m$  sensitive to the measurement  $C_i$  (e.g. Fig. 2). Because the sensitivity of  $g_m$  to  $C_i$  was affected by growth PPFD, this could potentially account for some of the variation in responses of  $g_m$  to  $C_i$  in the literature. However, as shown here and in some other studies (e.g. Flexas *et al.* 2007a, and Hassiotou *et al.* 2009), high sensitivity of  $g_m$  to  $C_i$

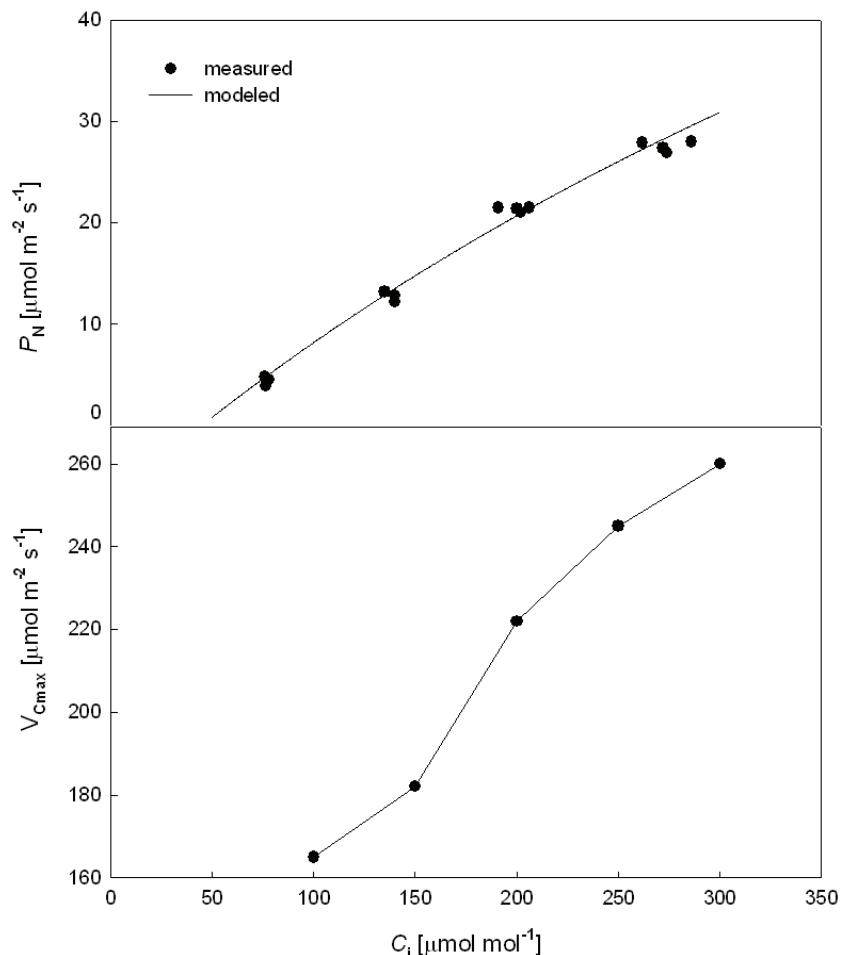


Fig. 4. Measured and modeled responses of  $P_N$  to  $C_i$  in common bean leaves, assuming infinite  $g_m$  and a single value of  $V_{C\max}$ , and values of  $V_{C\max}$  needed to predict measured  $P_N$  values of those leaves using measured values of  $g_m$ . Plants were grown at 25°C, 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD, and leaves of four plants were measured at 25°C and 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD.

can occur in some species even when grown at high light.

Tazoe *et al.* (2009) reported no effect of measurement PPFD on  $g_m$  in wheat leaves measured over the range of 200 to 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , while Hassiotou *et al.* (2009) found a 22% decrease in  $g_m$  at 500 vs. 1,500  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . Our results for bean and soybean measured at 150  $\mu\text{mol m}^{-2} \text{s}^{-1}$  similarly indicate only minor effects of measurement PPFD on the absolute value of  $g_m$  when averaged over the range of  $C_i$  measured. However, for both bean and soybean, low measurement PPFD substantially increased the sensitivity of  $g_m$  to  $C_i$ .

There has been debate whether  $g_m$  scales with maximum  $P_N$  such that the  $C_c$  to  $C_i$  ratio varies little among species (Evans and von Caemmerer 1996, Loreto *et al.* 2002, Warren and Adams 2006). The data for bean and soybean indicate that the  $C_c$  to  $C_i$  ratio often decreased with measurement  $C_i$ . The overall range in the ratio was reasonably small for plants grown at high PPFD, but quite large for both species when grown at low PPFD.

The value of  $g_m$  has been found to depend on chloroplast position (Tholen 2008) and on aquaporin activity (Terashima and Ono 2002, Flexas *et al.* 2006,

Miyazawa *et al.* 2008), as well as being temperature-dependent in some cases (Yamori *et al.* 2005, Warren and Dryer 2006, Warren 2008b), but it is not known how those responses may relate to changes in  $g_m$  with  $C_i$ , or with effects of growth and measurement PPFD.

Simple biochemical models of  $C_3$  photosynthesis which assumed infinite  $g_m$  have been extremely useful in diverse applications in agriculture and ecology. Farquhar-type biochemical models of  $C_3$  photosynthesis can generally be fit to  $P_N$  vs.  $C_i$  data in the  $C_i$  range of 100 to 300  $\mu\text{mol mol}^{-1}$  with a single value of  $V_{C\max}$  or a single value of  $V_{C\max}$  combined with a single value of  $g_m$  (Sharkey *et al.* 2007). However, because  $C_c$  to  $C_i$  ratio decreased with  $C_i$  in most cases in bean and soybean

(Fig. 3), the  $V_{C\max}$  values required to fit the model based on  $C_c$  with the data increased substantially with  $C_i$ . This is illustrated in Fig. 4 for bean grown and measured at 25°C and high PPFD, where a single value of  $V_{C\max}$  gave a satisfactory fit to the data when  $C_c$  was assumed to be equal to  $C_i$ , but  $V_{C\max}$  had to increase strongly with  $C_i$  when using the measured values of  $g_m$ . Because the ratio of  $C_c$  to  $C_i$  decreased with increasing  $C_i$  in this case (Fig. 3), a plot of  $V_{C\max}$  against  $C_c$  would be even steeper than that shown in Fig. 4. While this increase in  $V_{C\max}$  with  $C_i$  could reflect changing activation status of Rubisco (*i.e.* deactivation at low  $C_i$ ), it substantially complicates the modeling of photosynthesis until the response of  $g_m$  to  $C_i$  becomes predictable.

## References

Bunce, J.A. Acclimation of photosynthesis to temperature in *Arabidopsis thaliana* and *Brassica oleracea*. – *Photosynthetica* **46**: 517-524, 2008.

Bunce, J.A. Use of the response of photosynthesis to oxygen to estimate mesophyll conductance to carbon dioxide in water-stressed soybean leaves. – *Plant Cell Environ.* **32**: 875-881, 2009.

During, H. Stomatal and mesophyll conductances control CO<sub>2</sub> transfer to chloroplasts in leaves of grapevine (*Vitis vinifera* L.). – *Vitis* **42**: 65-68, 2003.

Evans, J.R.: Potential errors in electron transport rates calculated from chlorophyll fluorescence as revealed by a multi-layer leaf model. – *Plant Cell Physiol.* **50**: 698-706, 2009.

Evans, J.R., von Caemmerer, S. Carbon dioxide diffusion inside leaves. – *Plant Physiol.* **110**: 339-346, 1996.

Flexas, J., Ribas-Carbo, M., Hanson, D.T., Bota, J., Otto, B., Cifre, J., McDowell, N., Medrano, H., Kadenhoff, R. Tobacco aquaporin Nt AQP1 is involved in mesophyll conductance to CO<sub>2</sub> *in vivo*. – *Plant J.* **48**: 427-439, 2006.

Flexas, J., Diaz-Espejo, A., Galmes, J., Kaldenhoff, R., Medrano, H., Ribas-Carbo, M.: Rapid variations of mesophyll conductance in response to changes in CO<sub>2</sub> concentration around leaves. – *Plant Cell Environ.* **30**: 1284-1298, 2007a.

Flexas, J., Ortuno, M.F., Ribas-Carbo, M., Diaz-Espejo, A., Florez-Sarasa, I.D., Medrano, H.: Mesophyll conductance to CO<sub>2</sub> in *Arabidopsis thaliana*. – *New Phytol.* **175**: 501-511, 2007b.

Flexas, J., Ribas-Carbo, M., Diaz-Espejo, A., Galmes, J., Medrano, H.: Mesophyll conductance to CO<sub>2</sub>: current knowledge and future prospects. – *Plant Cell Environ.* **31**: 602-621, 2008.

Hassiotou, F., Ludwig, M., Renton, M., Venklaas E.J., Evans, J.R.: Influence of leaf dry mass per area, CO<sub>2</sub>, and irradiance on mesophyll conductance in sclerophylls. – *J. Exp. Bot.* **60**: 2303-2314, 2009.

Loreto, F., Harley, P.C., Di Marco, G., Sharkey, T.D.: Estimation of mesophyll conductance to CO<sub>2</sub> flux by three different methods. – *Plant Physiol.* **98**: 1437-1443, 1992.

Miyazawa, S.-I., Yoshimura, S., Shinzaki, Y., Maeshima, M., Miyake, C.: Deactivation of aquaporins decreases internal conductance to CO<sub>2</sub> diffusion in tobacco leaves grown under long-term drought. – *Func. Plant Biol.* **35**: 533-564, 2008.

Rodeghiero M., Niinemets U. & Cescatti A. Major diffusion leaks of clamp-on cuvettes still unaccounted: how erroneous are the estimates of Farquhar et al. model parameters? – *Plant Cell Environ.* **30**: 1006-1022, 2007.

Sharkey, T.D., Bernacchi, C.J., Farquhar, G.D., Singsaas, E.L.: Fitting photosynthetic carbon dioxide response curves for  $C_3$  leaves. – *Plant Cell Environ.* **30**: 1035-1040, 2007.

Tazoe, Y., von Caemmerer, S., Badger, M.R., Evans, J.R. Light and CO<sub>2</sub> do not affect the mesophyll conductance to CO<sub>2</sub> diffusion in wheat leaves. – *J. Exp. Bot.* **60**: 2291-2301, 2009.

Terashima, I., Ono, K.: Effects of HgCl<sub>2</sub> on CO<sub>2</sub> dependence of leaf photosynthesis: Evidence indicating involvement of aquaporins in CO<sub>2</sub> diffusion across the plasma membrane. – *Plant Cell Physiol.* **43**: 70-78, 2002.

Tholen, D., Boom, C., Noguchi, K., Uesda, S., Katase, T., Treashima, I.: The chloroplast avoidance response decreases internal conductance to CO<sub>2</sub> diffusion in *Arabidopsis thaliana* leaves. – *Plant Cell Environ.* **31**: 1688-1700, 2008.

Vrabl, D., Vaskova, M., Hrondova, M., Flexas, J., Santrucek, J.: Mesophyll conductance to CO<sub>2</sub> transport estimated by two independent methods: effect of variable CO<sub>2</sub> concentration and abscisic acid. – *J. Exp. Bot.* **60**: 2315-2323, 2009.

Warren, C.R.: Stand aside stomata, another actor deserves center stage: the forgotten role of the internal conductance to CO<sub>2</sub> transfer. – *J. Exp. Bot.* **59**: 1475-1487, 2008a.

Warren, C.R.: Does growth temperature affect the temperature responses of photosynthesis and internal conductance to CO<sub>2</sub>? A test with *Eucalyptus regnans*. – *Tree Physiol.* **28**: 11-19, 2008b.

Warren, C.R., Adams, M.A.: Internal conductance does not scale with photosynthetic capacity: implication for carbon isotope discrimination and the economics of water and nitrogen use in photosynthesis. – *Plant Cell Environ.* **29**: 192-201, 2006.

Warren, C.R., Dreyer, E.: Temperature response of photosynthesis and internal conductance to CO<sub>2</sub>: results from two independent approaches. – *J. Exp. Bot.* **57**: 3057-3067, 2006.

Warren, C.R., Low, M., Matyssek, R., Tausz, M.: Internal conductance to CO<sub>2</sub> transfer of adult *Fagus sylvatica*: Variation between sun and shade leaves and due to free-air ozone fumigation. – *Environ. Exp. Bot.* **59**: 130-138, 2007.

Yamori, W., Noguchi, K., Terashima, I.: Temperature acclimation of photosynthesis in spinach leaves: analyses of photosynthetic components and temperature dependencies of photosynthetic partial reactions. – *Plant Cell Environ.* **28**: 536-547, 2005.