

# Does elevated CO<sub>2</sub> protect photosynthesis from damage by high temperature *via* modifying leaf water status in maize seedlings?

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## Abstract

We hypothesized that decreased stomatal conductance ( $g_s$ ) at elevated CO<sub>2</sub> might decrease transpiration ( $E$ ), increase leaf water potential ( $\Psi_W$ ), and thereby protect net photosynthesis rate ( $P_N$ ) from heat damage in maize (*Zea mays* L) seedlings. To separate long-term effects of elevated CO<sub>2</sub>, plants grew at either ambient CO<sub>2</sub> or elevated CO<sub>2</sub>. During high-temperature treatment (HT) at 45°C for 15 min, leaves were exposed either to ambient CO<sub>2</sub> (380  $\mu\text{mol mol}^{-1}$ ) or to elevated CO<sub>2</sub> (560  $\mu\text{mol mol}^{-1}$ ). HT reduced  $P_N$  by 25 to 38% across four CO<sub>2</sub> combinations. However, the  $g_s$  and  $E$  did not differ among all CO<sub>2</sub> treatments during HT. After returning the leaf temperature to 35°C within 30 min,  $g_s$  and  $E$  were the same or higher than the initial values. Leaf water potential ( $\Psi_W$ ) was slightly lower at ambient CO<sub>2</sub>, but not at elevated CO<sub>2</sub>. This study highlighted that elevated CO<sub>2</sub> failed in protecting  $P_N$  from 45°C *via* decreasing  $g_s$  and  $\Psi_W$ .

*Additional key words:* heat stress, leaf water potential; net photosynthesis; stomatal conductance; vapor pressure deficit.

## Introduction

High air temperatures are a serious threat to crop production worldwide (Mearns *et al.* 1984). Heat stress affects photosynthetic and metabolic processes that ultimately influence the production of biomass, fruits, and grains (Hay and Walker 1989, Orbovic and Poff 2007). Rising atmospheric concentrations of CO<sub>2</sub> are expected to increase the frequency of extreme high-temperature events (Groisman and Knight 2008). Nevertheless, research on plant heat tolerance is not focused on the effects of elevated CO<sub>2</sub>.

In a few C<sub>3</sub> species, elevated CO<sub>2</sub> improved the tolerance of photosynthesis to high temperatures (Taub *et al.* 2000, Hamilton *et al.* 2008, Gutiérrez *et al.* 2009). Hamilton *et al.* (2008) exposed whole plants of maize and other species to high-temperature stress at two growth CO<sub>2</sub> concentrations. They found that elevated CO<sub>2</sub> did not always protect  $P_N$  from damage. Moreover, Wang *et al.* (2008) reported that elevated CO<sub>2</sub> even decreased the

tolerance to high temperatures in some cases. In C<sub>4</sub> species,  $P_N$  is usually near or at saturation for CO<sub>2</sub> at the current atmospheric concentration, thus, no positive effect of elevated CO<sub>2</sub> on  $P_N$  should be anticipated at high temperatures (Kim *et al.* 2007) as it is in C<sub>3</sub> species (Taub *et al.* 2000). However, indirect effects of CO<sub>2</sub> concentration could influence the tolerance of high temperatures even in C<sub>4</sub> species because of changes in  $g_s$ ,  $E$ , and  $\Psi_W$  at elevated CO<sub>2</sub>.

In the field, high-temperature stress is usually accompanied by high evaporative demand for water vapor. Thus, leaf desiccation can be involved in the damage caused by high temperatures. If desiccation is involved, then partial stomata closure caused by elevated CO<sub>2</sub> could mitigate the damage by high temperatures. Therefore, we hypothesized that decreased  $g_s$  at elevated CO<sub>2</sub> might protect  $P_N$  in C<sub>4</sub> species from damage caused by high-temperature stress, reducing  $E$ , and increasing leaf  $\Psi_W$ .

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**Abbreviations:** AC – ambient CO<sub>2</sub> concentration; C<sub>i</sub> – intercellular CO<sub>2</sub> concentration; CT – control; E – transpiration rate; EC – elevated CO<sub>2</sub> concentration;  $g_s$  – stomatal conductance; HT – high-temperature treatment;  $P_N$  – net photosynthetic rate;  $R_I$  – relative injury;  $T_{leaf}$  – leaf temperature; VPD – water vapor pressure deficit; WJC – water-jacketed cuvette; WUE – water-use efficiency;  $\Psi_W$  – water potential.

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## Materials and methods

**Growth:** Maize (*Zea mays L.* cv. Silver Queen) plants were grown in two environment-controlled chambers under either ambient CO<sub>2</sub> (AC, 380  $\mu\text{mol mol}^{-1}$ ) or elevated CO<sub>2</sub> (EC, 560  $\mu\text{mol mol}^{-1}$ ) concentrations. Plants were grown in vermiculite and irrigated daily with a complete nutrient solution containing 14.5 mM total N. Day/night air temperatures were 29/17°C, while soil temperature averaged 25.7  $\pm$  0.33°C/14.8  $\pm$  0.41°C. Light was provided by a mixture of high-pressure sodium and metal halide lamps (*Shanghai Yahong Electrical Lighting Co., Ltd*, China) at a photosynthetic photon flux density (PPFD) of 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , with a photoperiod of 12 h. Air humidity was 60% during the daytime. When the 4<sup>th</sup> leaf numbered from the bottom was fully expanded (14 d after emergence), a single leaf was exposed to a high-temperature treatment described below.

**High-temperature treatment (HT):**  $P_N$ ,  $g_s$ , and intercellular CO<sub>2</sub> concentration ( $C_i$ ) of nonstressed leaves (control, CT) were measured over a range of CO<sub>2</sub> concentrations and temperatures with a portable photosynthesis system (*CIRAS-2, PP-Systems*, Amesbury, MA, USA) equipped with a LED light source, at the growth PPFD of 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . In a preliminary test,  $P_N$  were significantly reduced after exposure to 45°C for 15 min, but recovered to almost the initial rates after 24 h. Thus, HT of 45°C for 15 min was chosen for this study, because it produced significant, but slowly reversible damage to photosynthesis.

For each measurement, single, intact 4<sup>th</sup> leaf attached to the plants, numbered from the bottom, with leaf area of *ca.* 40  $\text{cm}^2$  was placed inside a water-jacketed cuvette (WJC) with an internal fan (Bunce 2006) in the following arrangement:

Combination	Growth	During HT
AA	AC	AC
AE	AC	EC
EA	EC	AC
EE	EC	EC

Leaf temperature was measured using a miniature thermister (NTC 5k, *ET Enterprises Ltd.*, UK) pressed against the lower leaf surface. These leaf sections were then exposed to gradually increasing leaf temperatures ( $T_{\text{leaf}}$ ) from 35 to 45°C, held at 45°C for 15 min, and then  $T_{\text{leaf}}$  was gradually decreased again to 35°C (Fig. 1). The whole HT cycle was completed within 1 h. During this cycle, light intensities were maintained at 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD, and the water vapor content of air in the WJC was held constant. A *Ciras-1* portable photosynthesis system (*PP-Systems*, Amesbury, MA) programmed to use an external air supply and leaf chamber was used to record the

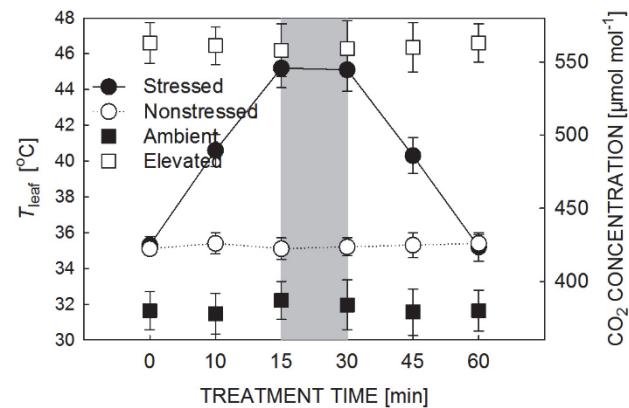


Fig. 1. Time courses of leaf temperatures ( $T_{\text{leaf}}$ ) and CO<sub>2</sub> concentrations during the heat stress cycles. During the whole temperature cycle, light was maintained at PPFD of 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . Vertical bars represent SE.

CO<sub>2</sub> concentration,  $P_N$ ,  $g_s$ ,  $C_i$ ,  $T_{\text{leaf}}$ , and water vapour pressure deficit (VPD) for the intact leaf inside the WJC throughout the temperature cycle (Bunce 2006). The reduction in photosynthesis due to the HT was quantified as: (1) the differences between  $P_N$  before and after HT for each temperature, and (2) the percentage decrease in  $P_N$  after HT for each temperature calculated as:  $[(P_{\text{NHT}} - P_{\text{NC}})/P_{\text{NC}}] \times 100$ , where  $P_{\text{NHT}}$  is  $P_N$  after HT treatments and  $P_{\text{NC}}$  is  $P_N$  before HT treatments for each temperature, as in Ameye *et al.* (2012). Water-use efficiency (WUE) was calculated as a ratio of  $P_N$  to  $E$ . Nine leaves from 9 different plants were subjected to HT for each of the four combinations of growth and measurement CO<sub>2</sub> concentration. For  $P_N$  measurements, 9 leaves per treatment were sampled.

**Water potential ( $\Psi_w$ ) and electrolyte leakage:** Leaf samples were collected from the control and stressed, 4<sup>th</sup> leaves after the HT. For the measurement of leaf  $\Psi_w$ , leaf disc of a 6-mm diameter was removed from the leaf section in the WJC and immediately sealed in a sample chamber for determination of  $\Psi_w$  using a *Wescor HR-33* dew point hygrometer (*Wescor, Inc.*, Logan, UT), as previously described by Sicher and Barnaby (2012). Electrolyte leakage was expressed as relative injury ( $R_l$ ) according to the procedure of Warren *et al.* (1996) with minor modification. Five leaf discs taken from the leaf inside the WJC, washed 3 times, and placed in glass tubes containing 30 mL of distilled water. The tubes were kept at room temperature (about 20°C) for 16 h. The tubes were then gently shaken for 15 min and electrical conductivity was measured with an *Okaton 510* conductivity meter (*Eutech Instrumenta*, Singapore). The conductivity values were designated as  $T_1$ . The tubes were then boiled in a microwave oven for 3 min to release all electrolytes. After

cooling to room temperature, the tubes were briefly shaken and the conductivity was measured as the total ion content ( $T_2$ ). Average conductivity of distilled water served as the basic ion content ( $T_0$ ). The  $R_I$  after HT was calculated by  $[(T_1 - T_0)/(T_2 - T_0)] \times 100$ . Because of the limited size of the leaf material inside the WJC, discs for water potential and electrolyte leakage measurements were taken from different plants. Before sampling, they were subjected to

HT in a similar fashion as those for the  $P_N$  measurements.

**Statistical analysis:** For the  $\Psi_W$  and electrolyte leakage analyses,  $n = 4$ . Two-way analysis of variance (*ANOVA*) was used to test the effects of growth CO<sub>2</sub>, exposure CO<sub>2</sub>, and their interaction for the leaf gas exchange data. Means were compared using *Fisher's* protected LSD test. A program used for calculations is *StatView* (SAS Institute Inc., USA).

## Results

The HT reduced  $P_N$  in all combinations of growth and measurement CO<sub>2</sub> concentration (Table 1).  $P_N$  decreased after HT by 24–38% compared with the CT in all CO<sub>2</sub> combinations. The decrease in  $P_N$  was significantly lesser when the HT occurred at AC rather than at EC (Table 1). In the plants grown and exposed to AC,  $P_N$  was the highest at the beginning of HT and at the end of HT in all combinations of CO<sub>2</sub>. It suggested that EC did not protect  $P_N$  from the damage caused by HT (Fig. 2). Both growth

CO<sub>2</sub> and its interaction with exposure CO<sub>2</sub> had no significant effect on the decrease of  $P_N$  (Table 1). Exposure to AC rather than EC prior to the HT resulted in higher  $g_s$  and  $E$  at both 35°C and 40°C in plants from both growth CO<sub>2</sub> concentrations (Fig. 3A). During the HT,  $g_s$ ,  $E$ , VPD, WUE, and  $\Psi_W$  increased in all CO<sub>2</sub> combinations, while  $P_N$  declined (Figs. 2,3,4). After returning the  $T_{leaf}$  to 40°C and then to 35°C within 30 min,  $g_s$  and  $C_i$  were the same or higher than the initial values at those temperatures, while  $P_N$

Table 1. Effects of CO<sub>2</sub> concentration during growth and during exposure to high temperature and their effects on the decreases in  $P_N$  after a heat stress treatment, compared with values at each temperature measured before the heat stress. Within each temperature, values with the same *lowercase letters* are not significantly different using *Fisher's* LSD test. ns – not significant; \* –  $P \leq 0.05$ ; \*\* –  $P \leq 0.01$ . G, E – growth and exposure to CO<sub>2</sub> concentrations, respectively; AC, EC – ambient and elevated CO<sub>2</sub>, respectively.  $T_{leaf}$  – leaf temperature. Values are  $\pm$  SE for  $n = 9$ .

CO <sub>2</sub> treatments	Decrease in $P_N$ [ $\mu\text{mol}(\text{CO}_2) \text{m}^{-2} \text{s}^{-1}$ ]			Decrease in $P_N$ [%]		
	$T_{leaf}$ [°C]			$T_{leaf}$ [°C]		
	35	40	45	35	40	45
GAC-EAC	9.6 $\pm$ 2.2 <sup>b</sup>	10.9 $\pm$ 2 <sup>b</sup>	7.5 $\pm$ 1.6 <sup>a</sup>	26.6 $\pm$ 2.8 <sup>b</sup>	31.2 $\pm$ 2.7 <sup>b</sup>	25.5 $\pm$ 2.5 <sup>b</sup>
GAC-EEC	13.3 $\pm$ 2.0 <sup>a</sup>	15.5 $\pm$ 2.2 <sup>a</sup>	9.5 $\pm$ 2.0 <sup>a</sup>	35.0 $\pm$ 2.5 <sup>ab</sup>	41.6 $\pm$ 2.8 <sup>ab</sup>	35.1 $\pm$ 3.8 <sup>ab</sup>
GEC-EAC	11.9 $\pm$ 1.5 <sup>a</sup>	12.3 $\pm$ 2.3 <sup>a</sup>	6.1 $\pm$ 1.9 <sup>b</sup>	34.0 $\pm$ 1.9 <sup>ab</sup>	36.9 $\pm$ 2.7 <sup>ab</sup>	24.8 $\pm$ 3.8 <sup>b</sup>
GEC-EEC	14.3 $\pm$ 3.9 <sup>a</sup>	16.9 $\pm$ 3.4 <sup>a</sup>	9.8 $\pm$ 1.9 <sup>a</sup>	37.6 $\pm$ 4.9 <sup>a</sup>	44.7 $\pm$ 4.4 <sup>a</sup>	38.1 $\pm$ 4.6 <sup>a</sup>
G	ns	ns	ns	ns	ns	ns
E	*	**	*	ns	*	**
Interaction	ns	ns	ns	ns	ns	ns

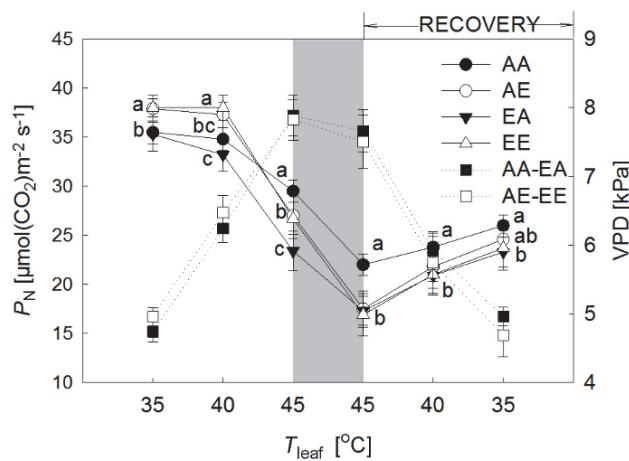


Fig. 2. Net photosynthetic rate ( $P_N$ ) in response to the high-temperature treatment of 4 combinations of growth and exposure to CO<sub>2</sub> for maize seedlings, and the water vapor pressure deficit (VPD), to which leaves were exposed.  $P_N$  and VPD are depicted with solid and dotted lines, respectively. AA – growth and exposure at the ambient carbon dioxide concentration (AC, 380  $\mu\text{mol mol}^{-1}$ ), AE – growth at AC and exposure at the elevated concentration (EC, 560  $\mu\text{mol mol}^{-1}$ ), EE – growth and exposure at EC, and EA – growth at the EC and exposure at the AC. AA-EE – exposure to heat stress at AC across growth CO<sub>2</sub>, AE-EE – exposure to heat stress at EC across growth CO<sub>2</sub>. Within each temperature point, values among different [CO<sub>2</sub>] treatments with same *lowercase letter* are not significantly different ( $P > 0.05$ ). Grey area represents the 15-min period of exposure to high temperature. Vertical bars represent SE for  $n = 9$ .  $T_{leaf} = 45^\circ\text{C}$ .

was lower than initial values (Figs. 2,3). Leaf  $\Psi_w$  was slightly lower after the HT at AC, but not at EC (Fig. 4A).

There were higher values of  $R_I$  from leaf tissue after the

## Discussion

Our results underlined that the damage to  $P_N$  due to 45°C was greater under EC, regardless of EC used for plant growth, compared to the plants grown and exposed to AC. Thus, EC failed in protecting  $P_N$  from 45°C in this maize cultivar. Hamilton *et al.* (2008) showed that EC increased

HT at both CO<sub>2</sub> concentrations, but the values of  $R_I$  were lower at AC than at EC (Fig. 4C).

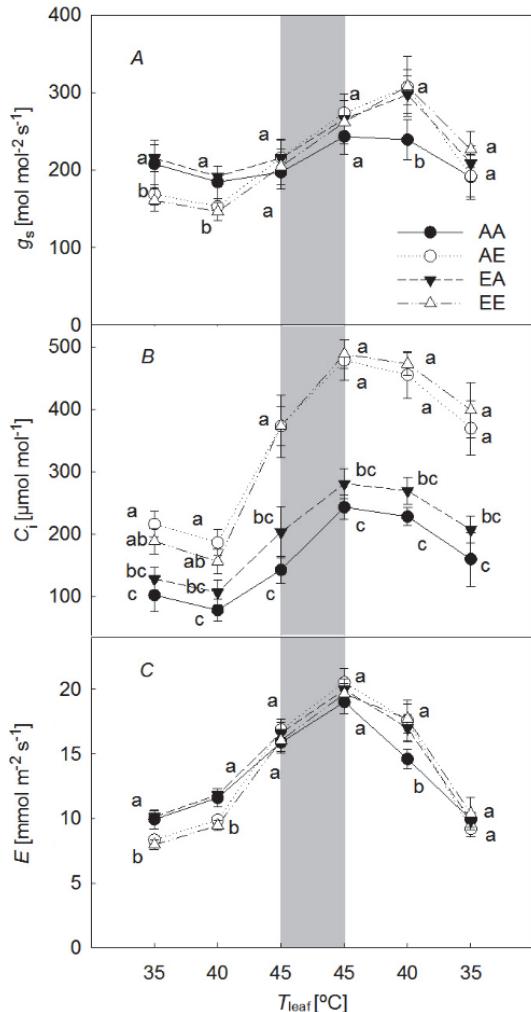


Fig. 3. Effects of a high-temperature treatment on *A*: stomatal conductance ( $g_s$ ), *B*: intracellular CO<sub>2</sub> ( $C_i$ ) and *C*: transpiration ( $E$ ) under 4 combinations of growth and exposure CO<sub>2</sub> concentrations. AA – growth and exposure at the ambient carbon dioxide concentration (AC, 380  $\mu\text{mol mol}^{-1}$ ), AE – growth at AC and exposure at the elevated concentration (EC, 560  $\mu\text{mol mol}^{-1}$ ), EE – growth and exposure at EC, and EA – growth at EC and exposure at AC. Within each temperature point, values among different CO<sub>2</sub> treatments with same lowercase letter are not significantly different using Fisher's LSD test ( $P > 0.05$ ). The grey area represents the high-temperature treatment of 15 min. Vertical bars represent SE for  $n = 9$ .

the damage to  $P_N$  caused by the HT in maize at 50°C. In their study, maize, pigweed, lambs quarters, and pea were subjected to 3 different temperature treatments. Results showed that the negative EC effects occurred mainly at the highest temperature. In maize, CO<sub>2</sub> effects similar to those

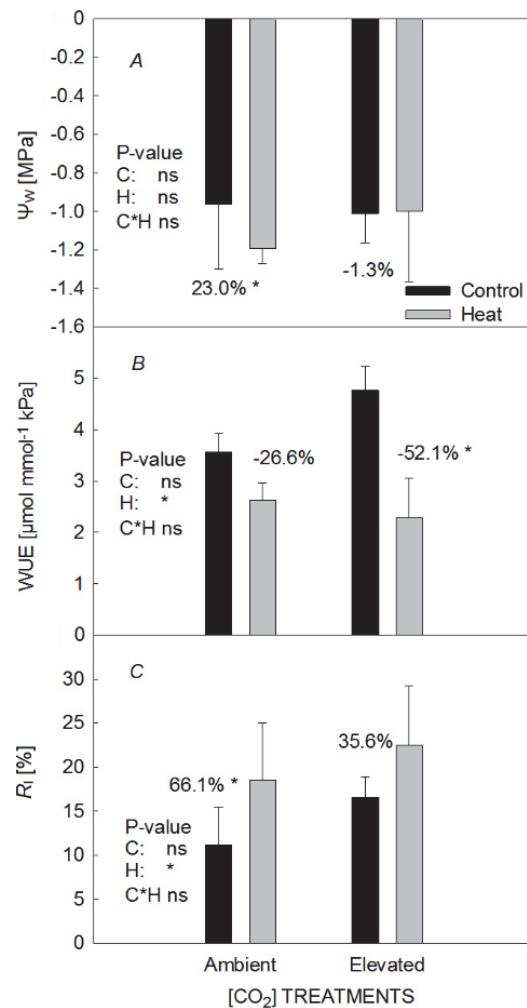


Fig. 4. High-temperature treatment induced changes in *A*: water potential ( $\Psi_w$ ), *B*: water-use efficiency (WUE), and *C*: leaf relative injury ( $R_I$ ) in leaves grown and measured at ambient and elevated CO<sub>2</sub>. CO<sub>2</sub> effect (C), heat stress effect (H), and their interaction effect were inserted into each panel. ns – not significant; \* –  $P < 0.05$ . Vertical bars represent SE, with  $n = 4$  for  $\Psi_w$  and  $R_I$  and  $n = 9$  for WUE.

observed here occurred also at 45°C (Wang *et al.* 2008). The lower  $P_N$  (Fig. 2), and higher  $g_s$  (Fig. 3A), and  $C_i$  (Fig. 3B) indicated that nonstomatal inhibition of  $P_N$

occurred after the HT. This was also found after HT stress in grapes (Luo *et al.* 2011) and in birch (Ranney and Peet 1994).

With high  $g_s$  and VPD during the HT, the values of  $E$  were 16 to 21 mmol m<sup>-2</sup> s<sup>-1</sup>, approximately twice of those at 35°C (Fig. 4). Thus, it was remarkable that the HT did not lower  $\Psi_w$  substantially. The lowest  $\Psi_w$  observed (−1.2 MPa) was too high to cause inhibition of  $P_N$  in this cultivar (Barnaby and Sicher 2012). Thus, the HT did not produce damaging leaf  $\Psi_w$  at two CO<sub>2</sub> concentrations despite of high VPD and  $E$ . The lack of stomata response to CO<sub>2</sub> concentration and the lack of significant leaf water deficits during the HT stress explained why our hypothesis was contradicted by the experimental data. Our results indicated that damage to  $P_N$  occurred at 45°C without the involvement of low leaf  $\Psi_w$ .

The premise of our hypothesis was that part of the damage to  $P_N$  caused by HT in combination with high VPD would be caused by low leaf  $\Psi_w$  resulting from high  $E$ . In such situation, we expected EC to decrease  $g_s$  and reduce the drop in leaf  $\Psi_w$ , thus preventing the damage. This scenario did not occur for two reasons in this maize cultivar. First,  $T_{leaf}$  of 45°C was high enough to cause

damage to  $P_N$  in this species. Fig. 3 clearly indicated that during HT no differences in  $g_s$  occurred between the AC and EC. This was even true comparing CO<sub>2</sub> concentrations of 200 and 1,000 μmol mol<sup>-1</sup> (data not shown). In addition, because the HT occurred at high VPD, it could be either HT or high VPD that eliminated the stomata response to CO<sub>2</sub>. Reduced CO<sub>2</sub> effect on  $g_s$  at high VPD has been reported in other species, *e.g.*, potato and sorghum (Bunce 2003), *Bromus japonicas* (Maherali *et al.* 2003), and wheat and barley (Bunce 2000b). An increase in  $g_s$  with temperature at constant water VPD occurs in many species including soybean, sunflower, tomato, turnip, winter wheat, barley (Bunce 2000a,b), and castor bean (Dai *et al.* 1992). Current study did not find a major weakening of membrane integrity by the heat stress. At this point, Xu *et al.* (2011) also found no ion leakage in maize after heat stress.

**Conclusion:** EC did not protect photosynthesis from inhibition caused by high temperature by modifying the leaf water status at high leaf temperature in maize. Actually, EC increased the inhibition of photosynthesis during the high temperature treatment.

## References

Ameye, M., Werten, T.M., Bauweraerts, I., *et al.*: The effect of induced heat waves on *Pinus taeda* and *Quercus rubra* seedlings in ambient and elevated CO<sub>2</sub> atmospheres. – *New Phytol.* **196**: 448-46, 2012.

Barnaby, J.Y., Sicher, R.C.: Impact of carbon dioxide enrichment on the responses of maize leaf transcripts and metabolites to water stress. – *Physiol. Plantarum* **144**: 238-253, 2012.

Bunce, J. A.: Acclimation of photosynthesis to temperature in eight cool and warm climate herbaceous C<sub>3</sub> species: temperature dependence of parameters of a biochemical photosynthesis model. – *Photosynth. Res.* **63**: 59-67, 2000a.

Bunce, J. A.: Responses of stomatal conductance to light, humidity and temperature in winter wheat and barley grown at three concentrations of carbon dioxide in the field. – *Glob. Change Biol.* **6**: 371-382, 2000b.

Bunce, J. A.: Effects of water vapor pressure difference on leaf gas exchange in potato and sorghum at ambient and elevated carbon dioxide under field conditions. – *Field Crop Res.* **82**: 37-47, 2003.

Bunce, J.A.: How do leaf hydraulics limit stomatal conductance at high water vapour pressure deficits? – *Plant Cell Environ.* **29**: 1644-1650, 2006.

Dai, Z., Ku, M.S.B., Edwards, G.E.: Control of photosynthesis and leaf conductance in *Ricinus communis* L. (castor bean) by leaf to air vapor pressure deficit. – *Plant Physiol.* **99**: 1426-1434, 1992.

Groisman, P.Y., Knight, R.W.: Prolonged dry episodes over the conterminous United States: new tendencies emerging during the last 40 years – *J. Climate* **21**: 1850-1862, 2008.

Gutiérrez, D., Gutiérrez, E., Pérez, P., *et al.*: Acclimation to future atmospheric CO<sub>2</sub> levels increases photochemical efficiency and mitigates photochemistry inhibition by warm temperatures in wheat under field chambers. – *Physiol. Plantarum* **137**: 86-100, 2009.

Hamilton, E.W., Heckathorn, S.A., Joshi, P., Wang, D., Barua, D.: Interactive effects of elevated CO<sub>2</sub> and growth temperature on the tolerance of photosynthesis to acute heat stress in C<sub>3</sub> and C<sub>4</sub> species. – *J. Integr. Plant Biol.* **50**: 1375-1387, 2008.

Hay, R.K.M., Walker, A.J.: An Introduction to the Physiology of Crop Yield. Pp. 292. Longman Scientific & Technical, New York 1989.

Kim S.H., Gitz D.C., Sicher R.C., *et al.*: Temperature dependence of growth, development, and photosynthesis in maize under elevated CO<sub>2</sub>. – *Environ. Exp. Bot.* **61**: 224-236, 2007.

Luo, H.B., Ma, L., Xi, H.F., *et al.*: Photosynthetic responses to heat treatments at different temperatures and following recovery in Grapevine (*Vitis amurensis* L.) leaves. – *PLOS ONE*: doi:10.1371/journal.pone.0023033, 2011.

Maherali, H., Johnson, H.B., Jackson, R.B.: Stomatal sensitivity to vapour pressure difference over a subambient to elevated CO<sub>2</sub> gradient in a C<sub>3</sub>/C<sub>4</sub> grassland. – *Plant Cell Environ.* **26**: 1297-1306, 2003.

Mearns, L.O., Katz, R.W., Schneider, S.H.: Extreme high temperature events: changes in their probabilities with changes in mean temperature. – *J. Clim. Appl. Meteorol.* **23**, 1601-1613, 1984.

Orbovic V., Poff, K.L.: Effect of temperature on growth and phototropism of *Arabidopsis thaliana* seedlings. – *J. Plant Growth Regul.* **26**: 222-228, 2007.

Ranney, T.G., Peet, M.M.: Heat tolerance of five taxa of birch (*Betula*): physiological responses to supraoptimal leaf temperatures – *J. Am. Soc. Hortic. Sci.* **119**: 243-248, 1994.

Sicher, R.C., Barnaby, J.Y.: Impact of carbon dioxide enrichment on the responses of maize leaf transcripts and metabolites to water stress. – *Physiol. Plantarum* **144**: 238-253, 2012.

Taub, D.R., Seemann, J.R., Coleman, J.S.: Growth in elevated CO<sub>2</sub> protects photosynthesis against high-temperature damage. – *Plant Cell Environ.* **23**: 649-656, 2000.

Wang, D., Heckathorn, S.A., Barua, D., *et al.*: Effects of elevated CO<sub>2</sub> on the tolerance of photosynthesis to acute heat stress in C<sub>3</sub>, C<sub>4</sub>, and CAM species. – Am. J. Bot. **95**: 165-176, 2008.

Warren, G.R., Marin, M.A., Teutonico, R.: Isolation of mutations affecting the development of freezing tolerance in *Arabidopsis thaliana* (L.). Heynh. – Plant Physiol. **111**: 1011-1019, 1996.

Xu, Z.Z., Zhou, G.S., Han, G.X., Li, Y.J.: Photosynthetic potential and its association with lipid peroxidation in response to high temperature at different leaf ages in maize. – J. Plant Growth Regul. **30**: 41-50, 2011.