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Single-cell microfluidics in combination with chlorophyll *a* fluorescence measurements to assess the lifetime of the *Chlamydomonas* PSBO protein

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Abstract

PSBO is an essential subunit of the oxygen-evolving complex and we recently demonstrated that its lifetime depends on environmental conditions in *Chlamydomonas reinhardtii*. To assess PSBO lifetime with a high time resolution, we employed (*1*) a microfluidic platform enabling the trapping of single cells and the parallel measurement of photosynthetic activity, and (*2*) a nitrate-inducible *PSBO* amiRNA line. Our microfluidic platform allowed the rapid replacement of the nutrient solution necessary for induction. It also enabled the precise monitoring of the decline in the F_v/F_m value, reflecting PSBO loss. We found that in the dark, at medium and high light intensity, the F_v/F_m value decreased with halftimes of about 25, 12.5, and 5 h, respectively. We also observed that photosynthetic activity was better sustained upon carbon limitation. In the absence of acetate, the halftimes of F_v/F_m diminishment doubled to quadrupled compared with the control, acetate-supplied cultures.

Keywords: carbon availability; chlorophyll fluorescence; oxygen-evolving complex; photoinhibition; protein lifetime.

Introduction

In a light-driven cycle, the manganese cluster (Mn_4CaO_5) of photosystem II (PSII) splits water molecules into oxygen, protons, and electrons. In plants and green algae, the Mn-cluster is shielded on the luminal side of the thylakoid membrane by the extrinsic proteins PSBO, PSBP, and PSBQ, with apparent molecular masses of 33, 23, and 17 kDa, respectively (reviewed by Ifuku and Noguchi 2016, Roose *et al.* 2016). These proteins stabilize the Mn-cluster and optimize oxygen evolution by regulating the access and retention of Ca^{2+} and Cl^- (Vinyard and Brudvig 2017). The extrinsic OEC subunits also protect the Mn-cluster from reductants (Popelkova

Highlights

- Microfluidics enables measuring the photosynthetic activity of single cells for days
- Assessment of the lifetime of PSBO became possible with high-time resolution
- PSBO lifetime varies by a factor of 10, depending on light and carbon availability

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Abbreviations: amiRNA – artificial microRNA; Chl – chlorophyll; *EV31* – empty vector control strain #31; F_0 – minimal fluorescence yield of the dark-adapted state; F_m – maximal fluorescence yield of the dark-adapted state; F_v/F_m – Chl *a* fluorescence parameter, reflecting photosynthetic integrity; OEC – oxygen-evolving complex; PSBO – 33 kD OEC extrinsic protein; TAP – Tris–acetate–phosphate medium; TP – Tris–phosphate medium.

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Conflict of interest: The authors declare that they have no conflict of interest.

et al. 2011, Bricker *et al.* 2012), including luminal ascorbate (Podmaniczki *et al.* 2021). The major subunit, PSBO, may also regulate the access of water to the Mn-cluster and proton removal *via* a hydrogen-bond network (Ho and Styring 2008, Offenbacher *et al.* 2013).

Due to its crucial role in photosynthetic oxygen evolution, PSBO is essential in vascular plants and for the photoautotrophic growth of algae (Mayfield *et al.* 1987, Liu *et al.* 2009, Pigolev and Klimov 2015). In *Arabidopsis thaliana*, PSBO has two isoforms. T-DNA knockout mutants of PSBO1 (*psbo1*) exhibit malfunction on both the donor and acceptor sides of PSII, high susceptibility to photoinhibition, and a severe growth phenotype (Allahverdiyeva *et al.* 2009). On the other hand, the absence of PSBO2 hardly affects PSII activity and plant growth (Lundin *et al.* 2007, Allahverdiyeva *et al.* 2009). In the green alga, *Chlamydomonas reinhardtii*, PSBO is encoded by a single gene (*Cre09.g396213*), and it is indispensable for oxygen evolution (Mayfield *et al.* 1987, Pigolev and Klimov 2015).

PSBO is relatively stable *in vitro* (Hashimoto *et al.* 1996). However, by employing nitrate-inducible PSBO artificial microRNA (amiRNA) lines, we showed that in *C. reinhardtii* PSBO has a significant turnover that is dependent on light intensity and carbon availability (Vidal-Meireles *et al.* 2023). Here, we aimed to obtain more specific data with improved time resolution on the lifetime of PSBO. To this end, we employed single-cell microfluidics. Microfluidic systems enable studying individual cells in a well-defined environment, *in situ* and in real time (*e.g.*, Kim *et al.* 2007, Barber and Emerson 2008, Song *et al.* 2020). Microfluidic technology is also suitable for precisely exchanging culture media without centrifugation or other treatments that may represent stress effects for the algal cells. Recently, we developed the so-called 'Tulip' microfluidics platform to capture and immobilize individual *C. reinhardtii* cells without binding them to a solid support surface (Széles *et al.* 2022); our system also enables the long-term measurement of the photosynthetic activity of single cells *via* chlorophyll (Chl) *a* fluorescence (Széles *et al.* 2022).

Materials and methods

Algal strains and growth conditions of bulk cultures: A nitrate-inducible *PSBO* amiRNA transformant, *PSBO-A58*, and its empty vector control *EV31* were used (Vidal-Meireles *et al.* 2023). The *PSBO* amiRNA transformant contains a *NIT1* promoter that drives the expression of the *PSBO* amiRNA construct; it is induced upon changing the nitrogen source from ammonium to nitrate in the growth medium (Schmollinger *et al.* 2010).

Cultures were grown in 50-ml Erlenmeyer flasks for three days in Tris–acetate–phosphate (TAP) medium at 22°C and 80 µmol(photon) m^{-2} s^{-1}, and they were supplemented with 1.87 mM NH_4Cl_2 on day two of culturing to prevent unwanted induction of the *PSBO* amiRNA construct. For the induction in bulk cultures, cells were washed four times with nitrogen-free Tris–phosphate (TP) medium and transferred to nitrate-containing (7.48 mM) TAP or TP medium in 250-ml Erlenmeyer flasks. The Chl (*a*+*b*) content was set to 5 µg mL^{-1}, determined according to Porra *et al.* (1989).

Chlorophyll a fluorescence measurements on bulk cultures: Chlorophyll *a* fluorescence measurements were performed as described in Nagy *et al.* (2018). Briefly, *C. reinhardtii* cultures were dark-adapted for about 15 min, and then 3 ml of cell suspension was filtered onto a *Whatman* glass microfibre filter (GF/B) and measured with a *Handy PEA* instrument (*Hansatech Instruments Ltd.*, UK).

Immunoblot analysis of PSBO: Sample collection was performed as described by Vidal-Meireles *et al.* (2023). Protein separation and Western blotting were carried out as described by Podmaniczki *et al.* (2021). A specific polyclonal antibody against PSBO was purchased from *Agrisera AB* (AS 06 142-33).

Construction and operation of the 'Tulip' microfluidic device: The microfluidic device was constructed using standard photolithography and soft lithography techniques (Whitesides *et al.* 2001). The 'Tulip' microfluidic device was designed in *KLayout*, an open-source software (www.klayout.de), and it was made of polydimethylsiloxane (*Sylgard 184*, *Dow Corning*), as described in Széles *et al.* (2022).

Liquid cell cultures and solutions were introduced into the microfluidic chips *via* tubing plugged into the inlet holes of the microfluidic device. Fluid flow was ensured and controlled by syringe pumps (*New Era Pump Systems Inc.*, *Model No. 4000*).

The fluid flow properties within the microfluidic device were modeled with *Comsol Multiphysics 4.3a* software. The velocity magnitude profiles were calculated by the 'Laminar flow' model. We applied a 'Shallow channel' approximation to get a quasi-3D model of the streamlines. The 'Tulip' device consists of three parallel channels that were taken into account when building the flow model. We used a 26.7 µl h^{-1} flow rate in the calculations.

Cell loading and culturing in microfluidic devices: The pre-cultures of *EV31* and *PSBO-A58* were grown in 25-ml Erlenmeyer flasks in TAP medium, at 22°C under 80 µmol(photon) m^{-2} s^{-1} for three days. Using the pre-culture as an inoculum, a 'main culture' was grown in TAP medium for three days, under the above-mentioned conditions. The Chl (*a*+*b*) content was set to 0.1 µg ml^{-1}, determined according to Porra *et al.* (1989). Both the pre-culture and the 'main culture' were supplemented with 1.87 mM NH_4Cl_2 on day two of cultivation.

Two separate units of the 'Tulip' microfluidic device were loaded with *EV31* and *PSBO* amiRNA cultures of 1 µg Chl (*a*+*b*) ml^{-1} in TAP medium, at a flow rate of 80 µl h^{-1} for 30 min, ensured by a syringe pump (*New Era Pump Systems Inc.*, *Model No. 4000*). After 30 min, TAP medium was provided, and the flow rate was increased to

360 µl h^{-1} for 4 h. Following that, the TAP medium was swapped with nitrate-containing (7.48 mM) TAP or TP medium at 360 µl h^{-1} flow rate for up to 96 h.

Continuous illumination was provided by white LED spot microscope lamps at an intensity of approx. 90 and 160 µmol(photon) m^{-2} s^{-1} on the surface of the microfluidic device or the cells were kept in the dark during the entire experiment.

Microscopy and Chl *a* fluorescence measurements: The *Microscopy* version of the *Imaging PAM M-series* Chl *a* fluorometer coupled to an *AxioScope A1* microscope (*Zeiss GmbH*) was employed to measure Chl *a* fluorescence using 20× and 63× objectives (*Zeiss*, *Fluar 20X/0.75 N.A* and *Zeiss*, *EC Plan-Neofluar 63X/1.25 N.A*, respectively). The 63× oil immersion objective was used for imaging. Bright-field images were captured by an *Axiocam 503* color CCD camera mounted on the microscope with a *60N-C 2/3" 0.63X* video adapter (*Zeiss GmbH*, Jena, Germany).

For Chl *a* fluorescence measurements, the LED light source was equipped with a timer which ensured 15-min dark adaptation before the Chl *a* fluorescence measurement performed every 2nd hour. Chl *a* fluorescence was induced by modulated blue (470 nm) measuring light and the emitted fluorescence image was captured by an *IMAG-K6 CCD* camera (*Walz GmbH*, Germany) mounted on the microscope *via* a *60N-C 2/3" 0.5X* video adapter. To determine the F_0 level of Chl *a* fluorescence, the measuring light intensity was set to level 2 and frequency to 2 Hz – using these settings, the measuring light intensity was not actinic. The gain was set to level 20 and damping to 1 and 5 for 20× and 63× objectives, respectively. F_m values were obtained by 820-ms saturating blue light pulses at an intensity level of 5. F_v/F_m, a sensitive parameter of PSII integrity, was calculated as $(F_m - F_0)/F_m$. The decay kinetics of F_v/F_m was fitted with a sigmoidal (logistic) function in *Origin* (*Microcal*) software and the halftime of F_v/F_m diminishment was determined directly from the fitting parameters (XC, the position of inflection points).

Electron microscopy: The 'Tulip' microfluidic device was imaged by scanning electron microscopy. For this purpose, the microfluidic devices were coated with a thin layer of gold by a *Quorum Q150T* sputter coater (100 mA, 120 s) and examined with a *JSM-7100F* field emission scanning electron microscope (using 5 kV acceleration voltage).

Statistical analysis: The presented data are based on at least three independent experiments. When applicable, averages and ± SE were calculated. The significance of the mean differences between the F_v/F_m values of bulk cultures of the *PSBO* amiRNA transformant and the *EV31* strain was analyzed by *Student*'s *t*-test using *GraphPad Prism5* software, and the significance levels are presented when applicable.

Results

Experiments on bulk PSBO amiRNA transformants: Each protein in a living cell has a certain lifetime ranging from minutes to weeks or even years. After this period, in a process called protein turnover, proteins are degraded and replaced by newly synthesized ones. Regarding the functionality of the photosynthetic electron transport chain, the lifetime of PSBO is particularly relevant, since it is an essential subunit of OEC (*e.g.*, Mayfield *et al.* 1987).

To study PSBO lifetime, we used a nitrate-inducible amiRNA line targeting *PSBO*, called *PSBO-A58*, characterized in our recent work (Vidal-Meireles *et al.* 2023). The *PSBO-A58* amiRNA transformant had an equal PSBO level under noninducing conditions as the *EV31* control strain (Vidal-Meireles *et al.* 2023). Under inducing conditions, *i.e.*, upon transferring the cultures from ammonia-containing to nitrate-containing TAP media, the amount of PSBO decreased slowly in the dark, more rapidly at normal light [approx. 100 µmol(photon) m^{-2} s^{-1}], and at a particularly high rate at high light [approx. 530 µmol(photon) m^{-2} s^{-1}] (Fig. 1). We also observed that in TP medium without carbon supply, the PSBO levels diminish more slowly than that in TAP medium (Fig. 1) or in TP medium with CO_2 supplementation (Vidal-Meireles

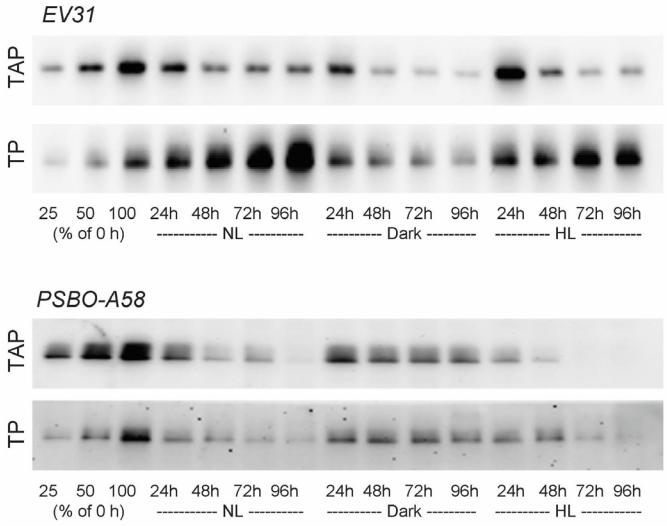

Fig. 1. Representative immunoblots to monitor the changes in PSBO levels in the empty vector *EV31* strain and the nitrate-inducible *PSBO-A58* amiRNA transformant in the nitrate-containing TAP and TP medium (*i.e.*, without acetate or other carbon supply). The induction was performed on bulk cultures at normal light [100 µmol(photon) m^{-2} s^{-1}, NL], in the dark (D), and at high light [530 µmol(photon) m^{-2} s^{-1}, HL]. The samples were loaded based on equal cell numbers. The 0 h samples represent the noninduced controls, with cell numbers of 25, 50, and 100%.

et al. 2023). On the other hand, we found that the relative *PSBO* amiRNA transcript abundance was equal, and the *PSBO* transcript level strongly diminished in all growth conditions (Vidal-Meireles *et al.* 2023). This means that the induction of the *PSBO* amiRNA construct was effective, irrespective of light intensity and carbon supply.

PSBO is essential to maintain OEC activity in green algae, and its loss entails donor-side induced photoinhibition in the light (Chen *et al.* 1995, Jegerschöld and Styring 1996), resulting in a substantial diminishment of the F_v/F_m value, a sensitive parameter of PSII integrity (Tóth *et al.* 2011, Vidal-Meireles *et al.* 2023). Donor-side induced photoinhibition is a fast process, occurring on a timescale of a few minutes; therefore, the F_v/F_m value is suitable for tracking the loss of PSBO in *C. reinhardtii.* Under noninducing conditions (*i.e.*, at time 0, Fig. 2), the F_v/F_m value was equal in the *PSBO-A58* and the *EV31* strains. In nitrate-containing TAP medium, the F_v/F_m value remained relatively high in the *EV31* strain (Fig. 2*A*), whereas in the *PSBO-A58* strain, it decreased severely (Fig. 2*B*), in parallel with the decline of the PSBO level (Fig. 1). In nitrate-containing TP medium, the F_v/F_m remained in the *EV31* strain (Fig. 2*C*), and it declined in the *PSBO-A58* strain albeit much more slowly than in nitrate-TAP (Fig. 2*D*). We also observed a clear light intensity dependence of the diminishment of PSBO (Fig. 2), in agreement with the immunoblot data (Fig. 1, *see* also Vidal-Meireles *et al.* 2023).

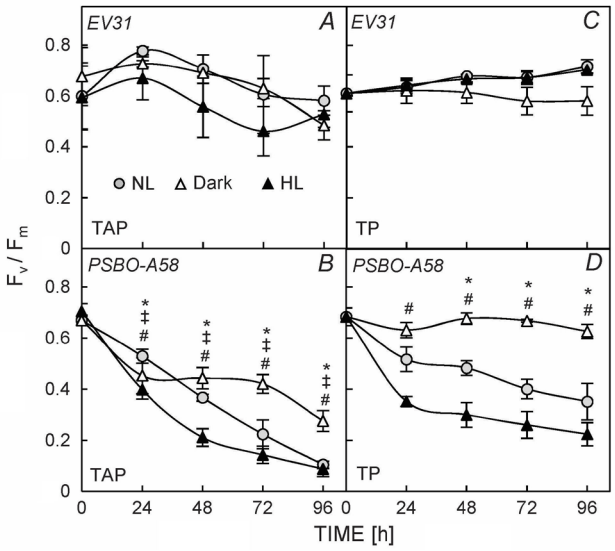

Fig. 2. The effects of downregulating *PSBO* on photosynthesis in bulk cultures, as assessed by the F_v/F_m Chl *a* fluorescence parameter, in the nitrate-inducible *PSBO-A58* amiRNA transformant and the empty vector *EV31* strain in the nitrate-containing TAP and TP medium. The induction was performed on bulk cultures at normal light [100 µmol(photon) m^{-2} s^{-1}, NL], in the dark (D), and at high light [530 µmol(photon) m^{-2} s^{-1}, HL]. Values are means ± SE of four biological replicates. Statistical significance levels are presented relative to the 0 h time point of each sample, as *, ‡, # for the NL, D, and HL samples, respectively, $p<0.05$.

Induction of the PSBO amiRNA transformants in the 'Tulip' microfluidics device: The above data demonstrate that PSBO lifetime is light-dependent and largely affected by carbon availability. To study PSBO lifetime with high-time resolution and precision, we employed single-cell microfluidics in combination with Chl *a* fluorescence measurement.

Our 'Tulip' microfluidics platform is suitable for trapping and immobilizing individual cells for several hours (Széles *et al.* 2022). Within the device, the traps are arranged in rows that are laterally shifted with respect to each other (Fig. 3*A*). The traps have a relatively wide opening (about 28 µm), a narrow middle section (about 8 µm), and an even narrower exit (about 3 µm) to prevent cells from escaping (Fig. 3*B*). The traps are relatively shallow (about 7 µm) to limit cell movement in the vertical direction thereby improving Chl *a* fluorescence imaging quality.

Computational modeling of the flow field demonstrates that in the case of empty traps, the magnitude of the flow velocity is about 6–7-fold lower at the entrance of the trap compared to the velocity between neighboring traps, and it is the highest at the narrow exits of individual traps (Fig. 3*C*). Moreover, upon trapping a cell, the flow

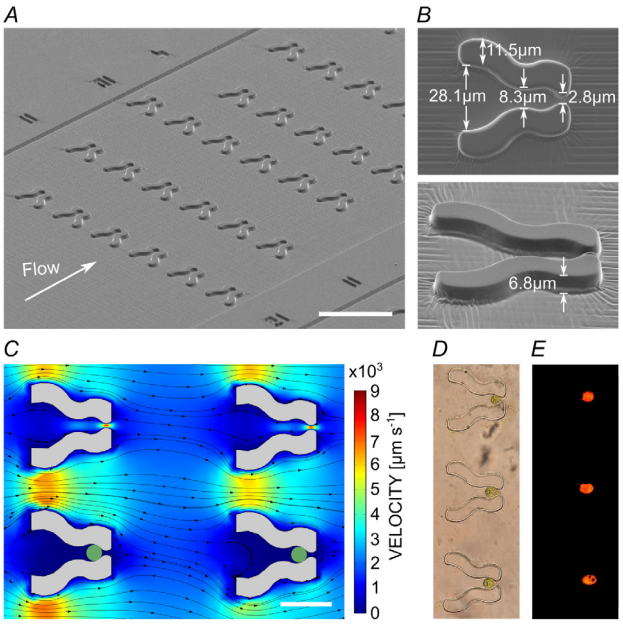

Fig. 3. 'Tulip' microfluidics platform for capturing and immobilizing single *C. reinhardtii* cells enabling the measurement of their photosynthetic activity. (*A*) Scanning electron microscopy image of the device with the direction of media flow indicated by *the arrow*. Scale bar is 100 µm. (*B*) Scanning electron microscopy images of single 'Tulip' traps from above and at a tilted angle (60°). The approximate characteristic sizes of the traps are shown. (*C*) Computational modeling of media flow in the device without (*upper line*) and with cells trapped inside (*lower row*). The density of the streamlines and the color code represent the velocity magnitude (the scale bar is shown on the right, 30 µm). (*D*) Representative bright-field microscopy image of trapped *C. reinhardtii* cells. (*E*) Maximum Chl *a* fluorescence (F_m) values of the captured cells in the 'Tulip' device.

is diminished, reducing the chance of another cell entering the trap (Fig. 3*C*). By holding the cells in their middle sections, the traps act as a physical constraint (Fig. 3*C*,*D*), enabling the measurement of photosynthetic parameters on a single cell level (Fig. 3*E*; Széles *et al.* 2022).

To investigate the lifetime of PSBO, nitrate-inducible *PSBO* amiRNA lines were introduced into the 'Tulip' microfluidic device. They were adapted to the new conditions for 4 h; following this stage, the culture media were exchanged from ammonia-containing (regular) TAP to nitrate-containing TAP medium. To investigate the dependence of PSBO lifetime on carbon availability, a nitrate-containing TP medium was used in a set of experiments.

The light dependence of PSBO lifetime was investigated under three conditions: darkness, 90 and 160 µmol(photon) m^{-2} s^{-1} of white light. We note that these light intensities were measured on the surface of the microfluidic device, *i.e.*, each cell receive this illumination continuously and uniformly during the entire course of the experiment.

In the dark, the F_v/F_m values of the *PSBO-A58* line decreased continuously in TAP medium and reached a 0 value by the 48th h of induction, with a halftime of about 24.7 h. In the TP medium, the diminishment of F_v/F_m, indicating PSBO loss, was about two-fold slower, occurring with a halftime of about 53 h (Fig. 4*A*).

In the presence of light, the diminishment of F_v/F_m was remarkably faster: in the TAP medium at 90 µmol(photon) m^{-2} s^{-1}, it occurred with a halftime of about 12.5 h, whereas in the carbon limitation (TP medium), the halftime was more than double, approx. 30 h (Fig. 4*B*).

At 160 µmol(photon) m^{-2} s^{-1}, the diminishment of F_v/F_m was further accelerated, and it reached a 0 value already after 24 h of illumination in the TAP medium, with a halftime of only 5 h. In the TP medium, the diminishment was four-fold slower, occurring with a halftime of about 20 h (Fig. 4*C*).

The above data show that the induction of the *PSBO-A58* line resulted in a very severe loss of photosynthetic activity. This was accompanied by morphological changes as well, as shown in Fig. 5; after 16 h of induction at 160 µmol(photon) m^{-2} s^{-1} in the TAP medium, the cell ultrastructure was severely altered, also substantially affecting the F_0 and F_m images.

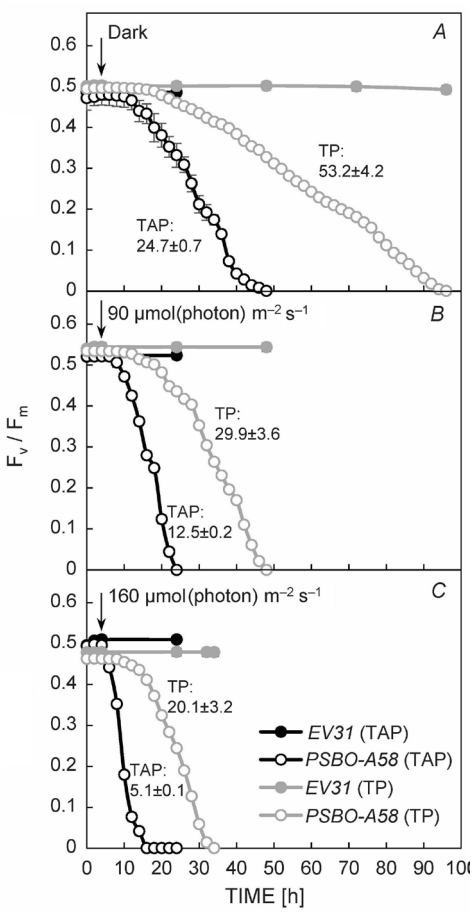

Fig. 4. The effects of downregulating *PSBO* on photosynthesis as assessed by the F_v/F_m parameter, in the nitrate-inducible *PSBO-A58* amiRNA transformant and the empty vector *EV31* strain in the nitrate-containing TAP and TP medium. The induction was performed in the 'Tulip' microfluidic device, in the dark (*A*), at normal light [90 µmol(photon) m^{-2} s^{-1}, *B*], and at high light [160 µmol(photon) m^{-2} s^{-1}, *C*]. Values are means ± SE of four biological replicates. *The arrows* indicate the start of the induction, *i.e.*, the time point of replacing the culture media with the nitrate-containing TAP or TP media. The kinetics were fitted with a Logistic equation, and the XC values, used to estimate the halftime of F_v/F_m diminishment are displayed in the panels together with the SE of the four independent experiments.

Discussion

Most studies on algal physiology have been performed on bulk cultures. The obtained parameters originate from heterogeneous cell populations, in which the individual cells may be in various cell cycle phases and each cell is subjected to continuously changing light conditions as the culture is being stirred. Microfluidics offers an attractive alternative to conventional methods of cultivating individual cells in a well-defined environment. The microfluidic technology also enables the precise control of the cellular microenvironment, including rapid and gentle media exchange; this is a notable advantage compared to bulk cultures, in which changing the culture media includes centrifugation, inferring stress effects for the cells.

Our microfluidics platform enables constant and stable exposure of individual cells to the given light intensity. Another major advantage is that the same cells can be monitored on the timescale of days. Moreover, the combination with Chl *a* fluorescence enables measuring photosynthetic parameters at high-time resolution and *in situ*. In conventional bulk cultures, determining the changes in photosynthetic activity requires frequent sampling, which could substantially affect the culture parameters. Our system is also automated, so the

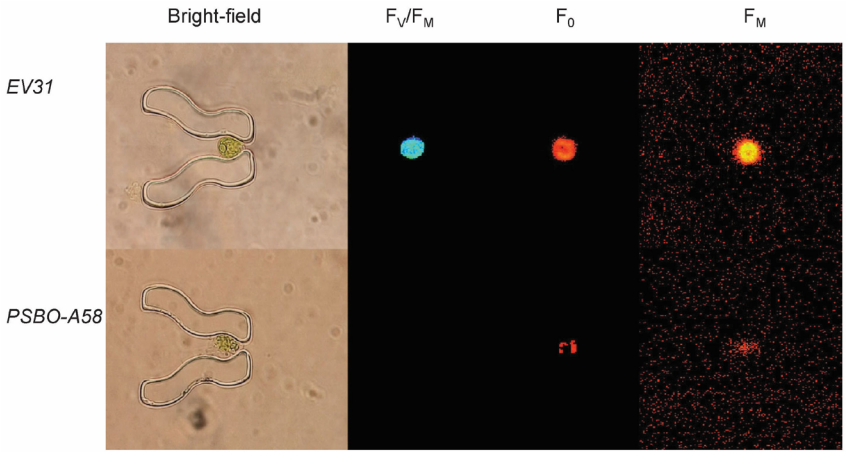

Fig. 5. Representative bright-field microscopy images, F_v/F_m, F_0, and F_m images of *PSBO-A58* amiRNA transformant and *EV31* cells in the nitrate-containing TAP medium after 16 h of induction at 160 µmol(photon) m^{-2} s^{-1} in the 'Tulip' microfluidics device.

measurements can be taken continuously for days, without human intervention.

Using nitrate-inducible amiRNA lines targeting the *PSBO* mRNA, we previously demonstrated that the lifetime of PSBO in *C. reinhardtii* is dependent on both light intensity and carbon source availability (Vidal-Meireles *et al.* 2023). Under noninducing conditions, our nitrate-inducible *PSBO* amiRNA transformants grow normally, and their photosynthetic activity is similar to that of the control strain. Upon induction of amiRNA expression, the cellular PSBO level diminishes, cell division is halted, and the Chl (*a*+*b*) content decreases (Vidal-Meireles *et al.* 2023). The arrest of cell division and the gradual decline in cellular PSBO content mean that PSBO has significant turnover. In Vidal-Meireles *et al.* (2023), bulk cultures were used, with a few sampling points to avoid changing culture parameters considerably. Therefore, the time course of the diminishment of PSBO level and photosynthetic activity consisted only of a few data points serving as a qualitative descriptor of PSBO lifetime (*see* also Figs. 1 and 2).

Here, with microfluidics combined with Chl *a* fluorescence-induction measurements, we could obtain high-resolution data on the decline of photosynthetic activity, reflecting the loss of PSBO. This approach is based on the fact that PSBO is essential for the biogenesis and stability of OEC (Liu *et al.* 2009, Pigolev and Klimov 2015); the diminishment of OEC activity results in a prompt decrease in F_v/F_m, and the rapidly occurring donor-side-induced photoinhibition further diminishes F_v/F_m (Tóth *et al.* 2011). Indeed, our data presented in this study (Figs. 1, 2) and our previous paper (Vidal-Meireles *et al.* 2023) show that the loss of PSBO and the decline of F_v/F_m occur in parallel in the inducible *PSBO* amiRNA lines.

We found that the diminishment of F_v/F_m in the TAP medium and 90 µmol(photon) m^{-2} s^{-1} occurs with halftime of about 12.5 h. By increasing the light intensity to 160 µmol(photon) m^{-2} s^{-1}, the diminishment of F_v/F_m became remarkably faster, occurring with halftime of about 5 h. In the dark, the F_v/F_m value decreased slowly, with halftime of about 25 h (Fig. 4).

On the other hand, keeping the cells in the TP medium (*i.e.*, carbon-limited conditions) doubled to quadrupled the lifetime of PSBO relative to the TAP medium: at 90 µmol(photon) m^{-2} s^{-1}, the F_v/F_m value diminished with halftime of about 30 h, whereas at 160 µmol(photon) m^{-2} s^{-1}, the halftime was about 20 h. In the dark, the diminishment of F_v/F_m occurred very slowly, with halftime of about 53 h.

We noted that the rate of the F_v/F_m diminishment is much faster in our microfluidic device than in bulk cultures (*cf.* Figs. 2 and 4); the reason may be that in the microfluidic device each cell is exposed continuously to the same, constant light intensity, and the cells do not shade each other. On the other hand, it is also conceivable that the exchange of culture media from the ammonium-containing to nitrate-containing ones occurs more effectively, therefore, the induction of the amiRNA construct is also faster.

Thus we could show that under photoautotrophic conditions (*i.e.*, in the TP medium), the cellular PSBO content is at least two-fold more stable in comparison with acetate-supplied cultures (Fig. 4). Acetate mitigates singlet oxygen production in *C. reinhardtii* (Roach *et al.* 2013) and excess acetate reduces the rates of CO_2 fixation and oxygen evolution without affecting PSII integrity (Heifetz *et al.* 2000), thus it is unlikely that acetate itself led to PSBO damage. Moreover, we found earlier that CO_2 supplementation also decreases the lifetime of PSBO (Vidal-Meireles *et al.* 2023), showing that PSBO lifetime in *C. reinhardtii* largely depends on carbon availability. This may be related to the fact that carbon-limited cultures are metabolically less active, which, similarly to yeast and mammals subjected to nutrient limitation, leads to lifespan prolongation (López-Otín *et al.* 2016, Sampaio-Marques *et al.* 2019, Zamzam *et al.* 2022).

Biomass accumulation can be limited in various biotechnological applications of green algae, such as in biofilm culturing systems (*e.g.*, Vajravel *et al.* 2020), and bio-hydrogen production platforms with restricted carbon assimilation (Kosourov *et al.* 2018, Nagy *et al.* 2021). Due to the high cost of establishing new algal cultures, they must be maintained for a long period to be remunerative.

Therefore, determining the lifetimes of photosynthetic subunits is highly relevant for the bio-industry. Our findings on PSBO lifetime suggest that the photosynthetic activity can be better maintained in moderate light and with a limited carbon supply.

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